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(74) Agents: OKABE, Masao et al.; No. 602, Fuji Bldg. 2-3,
Marunouchi 3-chome, Chiyoda-ku, Tokyo, 1000005 (JP).

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(71) Applicant (for all designated States except US): CANON
KABUSHIKI KAISHA [JP/JP]; 3-30-2, Shimomaruko,
Ohta-ku, Tokyo, 1468501 (JP).

(72) Inventors; and

(75) Inventors/Applicants (for US only): IMAMURA,
Takeshi [JP/JP]; c/o CANON KABUSHIKI KAISHA,
3-30-2, Shimomaruko, Ohta-ku, Tokyo, 1468501 (JP).
YANO, Tetsuya [JP/JP]; c/o CANON KABUSHIKI
KAISHA, 3-30-2, Shimomaruko, Ohta-ku, Tokyo,
1468501 (JP). HONMA, Tsutomu [JP/JP]; c/o CANON
KABUSHIKI KAISHA, 3-30-2, Shimomaruko, Ohta-ku,
Tokyo, 1468501 (JP). KOZAKI, Shinya [JP/JP]; c/o
CANON KABUSHIKI KAISHA, 3-30-2, Shimomaruko,
Ohta-ku, Tokyo, 1468501 (JP). NOMOTO, Tsuyoshi
[JP/JP]; c/o CANON KABUSHIKI KAISHA, 3-30-2,
Shimomaruko, Ohta-ku, Tokyo, 1468501 (JP). TSU-
CHITANI, Akiko [JP/JP]; c/o CANON KABUSHIKI
KAISHA, 3-30-2, Shimomaruko, Ohta-ku, Tokyo,
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(54) Title: STRUCTURED CONSTRUCT AND PRODUCING METHOD THEREFOR

(57) Abstract: There is provided a construct in which at least a part of the magnetic material is coated with polyhydroxyalkanoate (PHA), and a method for producing a construct by immobilizing a PHA synthesizing enzyme on the surface of the magnetic material thereby biosynthesizing a PHA and coating such PHA on the material.

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B. FIELDS SEARCHED

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Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

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C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
1 A	EP 1 275 728 A (CANON KK) 15 January 2003 (2003-01-15) page 4, line 50 - line 55 page 6, line 42 - page 15, line 30; claims; examples -----	1-24
1 X	EP 1 253 476 A (CANON KK) 30 October 2002 (2002-10-30) page 3, line 40 - page 4, line 22 page 9, line 49 - page 10, line 3 -----	1,2
1 X	EP 1 253 475 A (CANON KK) 30 October 2002 (2002-10-30) page 6, line 47 - page 10, line 34 page 11, line 10 - page 17, line 52 page 33, line 55 - line 58 page 37, line 29 - line 35 ----- -/--	1-19

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

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European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax (+31-70) 340-3016

Authorized officer

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C (Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
1	A US 6 133 047 A (SAUZEDDE FLORENCE ET AL) 17 October 2000 (2000-10-17) the whole document	20-24
1	A US 5 648 124 A (SUTOR JAMES J) 15 July 1997 (1997-07-15) the whole document	20-24
1	A WO 01/52612 A (ARKIS AHMED ; BIO MERIEUX (FR); DELAIR THIERRY (FR); ELAISSARI ABDELHA) 26 July 2001 (2001-07-26) page 3, line 1 - page 6, line 30; claims; examples	20-24
1	A WO 90/15666 A (OMNI QUEST CORP) 27 December 1990 (1990-12-27) the whole document	20-24
1	A US 6 521 341 B1 (ELAISSARI ABDELHAMID ET AL) 18 February 2003 (2003-02-18) column 3; claims; examples	20-24
1	A US 5 736 349 A (YOSHIOKA KATSUAKI ET AL) 7 April 1998 (1998-04-07) column 5, line 52 - line 56; claims 4,7,8; examples	20-24
1	A US 6 231 982 B1 (WANG CHENGRONG) 15 May 2001 (2001-05-15) column 2, line 64 - column 3, line 52; claims	20-24
1	A EP 1 213 587 A (COULTER INTERNAT CORP) 12 June 2002 (2002-06-12) the whole document	20-24
1	A US 6 048 515 A (LAWACZEK RUEDIGER ET AL) 11 April 2000 (2000-04-11) claims; figure 1; examples	20-24
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Box No. 1 Nucleotide and/or amino acid sequence(s) (Continuation of item 1.b of the first sheet)

1. With regard to any nucleotide and/or amino acid sequence disclosed in the international application and necessary to the claimed invention, the international search was carried out on the basis of:

a. type of material



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3. Additional comments:

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INTERNATIONAL SEARCH REPORT

Information on patent family members

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Patent document cited in search report	Publication date	Patent family member(s)	Publication date
EP 1275728 A	15-01-2003	EP 1275728 A1	15-01-2003
		JP 2004065001 A	04-03-2004
		US 2003224494 A1	04-12-2003
EP 1253476 A	30-10-2002	JP 2002327046 A	15-11-2002
		EP 1253476 A2	30-10-2002
		US 2003096115 A1	22-05-2003
EP 1253475 A	30-10-2002	JP 2003015359 A	17-01-2003
		EP 1253475 A2	30-10-2002
		US 2003118931 A1	26-06-2003
US 6133047 A	17-10-2000	FR 2749082 A1	28-11-1997
		AT 205115 T	15-09-2001
		CA 2227439 A1	04-12-1997
		DE 69706501 D1	11-10-2001
		DE 69706501 T2	16-05-2002
		EP 0840648 A1	13-05-1998
		ES 2162301 T3	16-12-2001
		WO 9745202 A1	04-12-1997
US 5648124 A	15-07-1997	WO 9637313 A1	28-11-1996
WO 0152612 A	26-07-2001	FR 2804117 A1	27-07-2001
		AU 3558001 A	31-07-2001
		CA 2412536 A1	26-07-2001
		EP 1248679 A2	16-10-2002
		WO 0152612 A2	26-07-2001
		US 2003175691 A1	18-09-2003
WO 9015666 A	27-12-1990	WO 9015666 A1	27-12-1990
US 6521341 B1	18-02-2003	FR 2773416 A1	09-07-1999
		AU 1973499 A	26-07-1999
		CA 2316824 A1	15-07-1999
		EP 1046037 A1	25-10-2000
		WO 9935500 A1	15-07-1999
		JP 2002501175 T	15-01-2002
US 5736349 A	07-04-1998	JP 2979414 B2	15-11-1999
		JP 3115862 A	16-05-1991
		US 5320944 A	14-06-1994
		AU 641687 B2	30-09-1993
		AU 6328690 A	11-04-1991
		CA 2026515 A1	30-03-1991
		DE 69021309 D1	07-09-1995
		DE 69021309 T2	01-02-1996
		EP 0420186 A2	03-04-1991
		ES 2077618 T3	01-12-1995
		KR 206159 B1	01-07-1999
		DE 69327926 D1	06-04-2000
		DE 69327926 T2	20-07-2000
		DE 585868 T1	22-09-1994
		EP 0585868 A2	09-03-1994
		ES 2052465 T1	16-07-1994
		JP 6231957 A	19-08-1994
		JP 7092168 A	07-04-1995

INTERNATIONAL SEARCH REPORT

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Patent document cited in search report	Publication date	Patent family member(s)	Publication date
US 6231982	B1	15-05-2001	DE 69815298 D1 10-07-2003
			DE 69815298 T2 06-05-2004
			EP 0961934 A1 08-12-1999
			ES 2200395 T3 01-03-2004
			JP 2001513203 T 28-08-2001
			WO 9930160 A1 17-06-1999
EP 1213587	A	12-06-2002	US 5466609 A 14-11-1995
			EP 1213587 A1 12-06-2002
			AU 5360394 A 09-05-1994
			CA 2146964 A1 28-04-1994
			DE 69331970 D1 04-07-2002
			DE 69331970 T2 19-12-2002
			EP 0665955 A1 09-08-1995
			JP 8502443 T 19-03-1996
			JP 3547437 B2 28-07-2004
			JP 2004003991 A 08-01-2004
			WO 9409368 A1 28-04-1994
			US 5639620 A 17-06-1997
			US 5776706 A 07-07-1998
			US 5707877 A 13-01-1998
US 6048515	A	11-04-2000	DE 4428851 A1 08-02-1996
			AU 703042 B2 11-03-1999
			AU 2921095 A 04-03-1996
			CA 2195318 A1 15-02-1996
			CN 1155844 A ,B 30-07-1997
			WO 9604017 A1 15-02-1996
			EP 0773796 A1 21-05-1997
			HU 77993 A2 28-04-1999
			IL 114713 A 17-02-2000
			IL 131562 A 31-08-2000
			JP 10503496 T 31-03-1998
			NO 970468 A 02-04-1997
			US 6576221 B1 10-06-2003
			US 2002141943 A1 03-10-2002
			ZA 9506005 A 22-02-1996

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(71) Applicant (*for all designated States except US*): CANON KABUSHIKI KAISHA [JP/JP]; 3-30-2, Shimomaruko, Ohta-ku, Tokyo, 1468501 (JP).

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(72) Inventors; and

(75) Inventors/Applicants (*for US only*): IMAMURA, Takeshi [JP/JP]; c/o CANON KABUSHIKI KAISHA, 3-30-2, Shimomaruko, Ohta-ku, Tokyo, 1468501 (JP). YANO, Tetsuya [JP/JP]; c/o CANON KABUSHIKI KAISHA, 3-30-2, Shimomaruko, Ohta-ku, Tokyo, 1468501 (JP). HONMA, Tsutomu [JP/JP]; c/o CANON KABUSHIKI KAISHA, 3-30-2, Shimomaruko, Ohta-ku, Tokyo, 1468501 (JP). KOZAKI, Shinya [JP/JP]; c/o CANON KABUSHIKI KAISHA, 3-30-2, Shimomaruko, Ohta-ku, Tokyo, 1468501 (JP). NOMOTO, Tsuyoshi [JP/JP]; c/o CANON KABUSHIKI KAISHA, 3-30-2, Shimomaruko, Ohta-ku, Tokyo, 1468501 (JP). TSUCHITANI, Akiko [JP/JP]; c/o CANON KABUSHIKI

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(57) Abstract: There is provided a construct in which at least a part of the magnetic material is coated with polyhydroxyalkanoate (PHA), and a method for producing a construct by immobilizing a PHA synthesizing enzyme on the surface of the magnetic material thereby biosynthesizing a PHA and coatings such PHA on the material.

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DESCRIPTION

STRUCTURED CONSTRUCT AND PRODUCING METHOD THEREFOR

5 TECHNICAL FIELD

The present invention relates to a construct characterized in including polyhydroxyalkanoate and a magnetic member and having a structure that the polyhydroxyalkanoate covers at least a part of the
10 magnetic member, and a producing method therefor.

The present invention also relates to a method for separating a specified target component contained in a specimen, a method for detecting such a target component, and a method for screening a specified
15 component utilizing the construct. More specifically, the present invention relates a method for selective separation, detection or screening of a specified target component contained in a specimen by forming a construct bearing, on a carrier surface, a molecule
20 having a specific affinitive coupling property to a specified target component, for example, a nucleic acid molecule, a protein, a peptide, a sugar, a lipid, a low-molecular compound or a composite thereof of natural origin or artificially modified nature, and
25 causing such a construct to bond with the specified target component. The present invention also relates to an apparatus exclusively utilizable for executing

the aforescribed methods.

BACKGROUND ART

Polymer materials are indispensable for modern industries and lives, and, owing to their features such as inexpensiveness, light-weight and satisfactory molding properties, are utilized in various fields such as a casing of electric of appliance, a packaging material, a cushioning material, a fiber material etc. Further, utilizing the stability of such polymer materials, various functional materials such as a liquid crystal material or a coating material are obtained by introducing substituents capable of exhibiting various functions into the molecular chain of a polymer material. Such a functional material, having a higher added value than in the polymer itself as the structural material, can expect a large market demand even with a small-scale production. Such a functional polymer material has been obtained by methods based on organic synthetic chemistry, in a polymer synthesis process or by a modification of a synthesized polymer with a substituent. The polymer, constituting the basic skeletal structure of the functional polymer material, is obtained, in most cases, from petroleum-based raw materials by methods based on organic synthetic chemistry. Typical

examples of such a polymer include polyethylene, polyethylene terephthalate, polyester, polystyrene, polyvinyl chloride and polyacrylamide.

<Multi-layered construct containing magnetic member>

5 The present inventors have investigated a multi-layered construct in which a magnetic material is coated by a polymer compound, as an elementary technology for providing a polymer compound with a high added value. Such coating of a specified
10 magnetic material with a polymer compound can provide a composite construct having an extremely useful functionality. Applications of such a construct include, for example, a carrier being a microcapsule construct containing a magnetic material in a polymer
15 compound for separating, purifying or screening a biological substance, and a magnetic recording medium formed by coating a sheet-shaped magnetic material with a polymer compound.

 Since such a microcapsule construct containing
20 magnetic material can be easily collected by magnetic force, excellent effects can be expected principally in biochemical fields, for example, as a carrier for a medical diagnostic drug, a carrier for separating germs or cells, a carrier for separating and
25 purifying nucleic acid or protein, a carrier for drug delivery, a carrier for an enzyme reaction, or a carrier for a cell culture. The capsule construct

containing a magnetic material can be synthesized, for example, by a method of dispersing an oleophilized magnetic material in a polymerizable monomer and executing suspension polymerization.

- 5 (Japanese Patent Application Laid-Open No. S59-221302), a method of dispersing an oleophilized magnetic material in a polymerizable monomer, and executing polymerization under homogenization in water with a homogenizer thereby obtaining magnetic
- 10 particles of a relative small particle size (Japanese Patent Publication No. H04-3088), or a method of precipitating and oxidizing an iron compound in the presence of porous polymer particles having a specified functional group to introduce a magnetic
- 15 material into the interior of the porous polymer particles, thereby obtaining magnetic particles of a large and uniform particle size (Japanese Patent Publication No. H05-10808).

- However, in case such a magnetic material-
- 20 containing capsule construct, obtained by these synthesizing methods, is applied, for example, to the carrier for the medical diagnostic drug, a practically acceptable performance is often not obtained because of a significant loss in the
- 25 sensitivity or by a non-specific reaction even when most of the magnetic material is present inside the capsule construct. This is presumably because the

magnetic material is partially exposed on the surface of the capsule construct or a micropath is formed between the surface of the construct and the internal magnetic material, whereby the magnetic component is dissolved out to hinder the practical performance.

The magnetic material generally is more hydrophilic than the polymer particles, and, in the conventional synthesizing methods, the magnetic material tends to be localized at the surface of the capsule construct or in the vicinity thereof, and such a fact constitutes a major reason for deterioration of the practical performance. As explained above, in the conventional magnetic material-containing capsule construct, it is difficult to suppress the elution of the magnetic component due to the exposure of the magnetic material on the surface or the micropath formation, and the application of such a capsule construct is in fact limited to fields where such elution is not problematic.

Also, various attempts have been reported to improve the surface characteristics of the magnetic material, thereby improving dispersibility of the magnetic material in a polymerized toner. Japanese Patent Application Laid-Open Nos. S59-200254, S59-200256, S59-200257 and S59-224102 propose treating a magnetic material with various silane coupling agents. Also Japanese Patent Application Laid-Open Nos. S63-

250660 and H10-239897 disclose a technology for treating silicon-containing magnetic particles with a silane coupling agent.

However, though these technologies can improve the dispersibility of the magnetic particles to a certain extent, it is difficult to attain uniform hydrophobicity of the surface of the magnetic material, and a further improvement has been desired in order to prevent mutual uniting of the magnetic particles and generation of non-hydrophobic magnetic particles and to improve the dispersibility of the magnetic material to a satisfactory level.

Also as an example of utilizing a hydrophobicized magnetic iron oxide, Japanese Patent Publication No. S60-3181 proposes a toner containing magnetic iron oxide treated with alkyltrialkoxysilane. Though the addition of such a magnetic iron oxide provides a certain improvement in the electrophotographic properties of the toner, further improvement is still desirable because the originally low surface activity of magnetic iron oxide tends to cause fused particles or uneven hydrophobicity in the course of treatment.

<PHA>

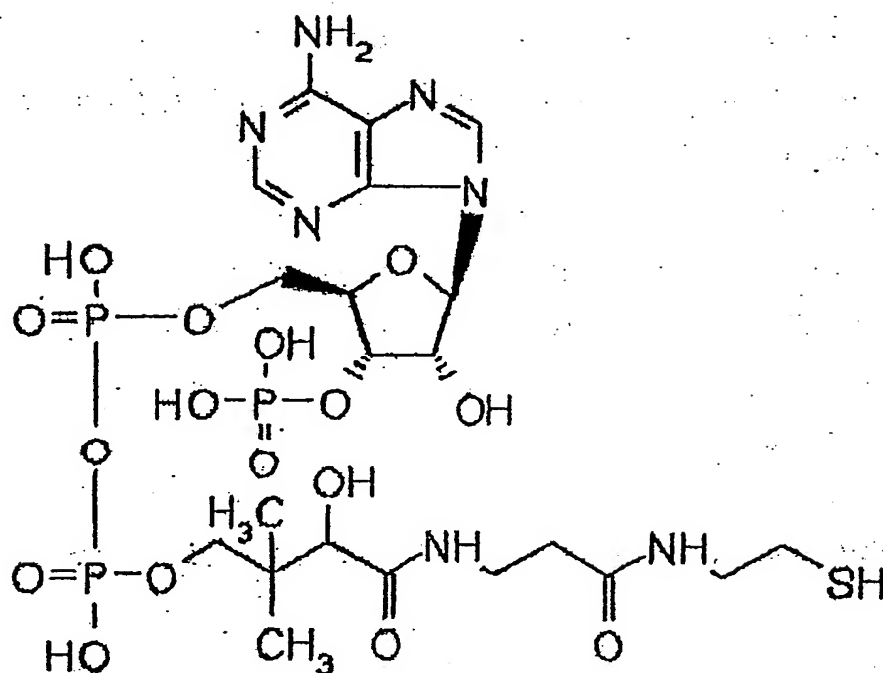
Meanwhile, researches for producing a polymer compound by a biological method have been actively carried out in recent years and are being

commercialized in part. Examples of polymer compounds derived from microorganisms include polyhydroxyalkanoate such as poly-3-hydroxy-n-butyric acid (hereinafter also abbreviated as PHB) or a
5 copolymer of 3-hydroxy-n-butyric acid and 3-hydroxy-n-valeric acid (hereinafter also abbreviated as PHB/V), a polysaccharide such as bacterial cellulose or purlan, a polyamino acid such as poly- γ -glutamic acid and polylysine. Polyhydroxyalkanoate
10 (hereinafter also represented as PHA) means polyhydroxyalkanoate containing a hydroxy alkanoic acid unit. In particular, PHA can be utilized in various products, for example, by melt-forming, like the conventional plastics. Also it shows satisfactory
15 biocompatibility and is expected as a soft material for medical use.

It has been reported that many microorganisms produce PHA and accumulate it within cells. For example, microbial production of PHB/V by *Alcaligenes eutrophus* H16 (ATCC No. 17699), *Methylobacterium* sp.,
20 *Paracoccus* sp., *Alcaligenes* sp., and *Pseudomonas* sp. has been reported (for example, Japanese Patent Application Laid-Open No. H05-74492, Japanese Patent Publication Nos. H06-15604, H07-14352, and H08-19227).
25 Furthermore, *Comamonas acidovorans* IFO 13852 produces PHA comprised of monomer units of 3-hydroxy-n-butyric acid and 4-hydroxy-n-butyric acid (Japanese Patent

Application Laid-Open No. H09-191893), and *Aeromonas caviae* produces a copolymer of 3-hydroxy-n-butyric acid and 3-hydroxyhexanoic acid (Japanese Patent Application Laid-Open Nos. H05-93049 and H07-265065).

5 Polyhydroxyalkanoate constituted of a 3-hydroxyalkanoic acid unit of a short-chain-length such as PHB or PHB/V (such PHA being hereinafter abbreviated also as scl-PHA) is synthesized by an enzymatic polymerization reaction using as a
10 substrate at least one of (R)-3-hydroxybutyryl CoA, (R)-3-hydroxypropionyl CoA and (R)-3-hydroxyvaleryl CoA that are synthesized from various carbon sources through various in vivo metabolic pathways. The enzyme that catalyzes this polymerization reaction is
15 called scl-PHA synthetase in the present invention. For example, an enzyme synthesizing PHB is called PHB synthetase (also called PHB polymerase or PHB synthase). CoA is an abbreviation for coenzyme A, and its chemical structure is represented by the
20 following chemical formula.



Recently, active studies on polyhydroxy
 alkanate comprised of 3-hydroxyalkanoic acid units
 5 of medium-chain-length (about 3 to 12 carbon atoms)
 (mcl-PHA) have been conducted.

For example, Japanese Patent No. 2642937
 discloses that *Pseudomonas oleovorans* ATCC 29347 can
 produce PHA comprised of 3-hydroxyalkanoic acid
 10 monomer units of 6 to 12 carbon atoms from non-cyclic
 aliphatic hydrocarbons. In addition, it has been
 reported, in Appl. Environ. Microbiol., 58, 746
 (1992), that *Pseudomonas resinovorans* produces PHA of
 which monomer units are 3-hydroxy-n-butyric acid, 3-

hydroxyhexanoic acid, 3-hydroxyoctanoic acid, and 3-hydroxydecanoic acid using octanoic acid as a sole carbon source, and it also produces PHA of which monomer units are 3-hydroxy-n-butyric acid, 3-hydroxyhexanoic acid, 3-hydroxyoctanoic acid, and 3-hydroxydecanoic acid using hexanoic acid as sole carbon source. Here, the 3-hydroxyalkanoic acid monomer units longer than the raw material fatty acid are considered derived from the fatty acid synthesizing pathway described below.

Int. J. Biol. Macromol., 16 (3), 119 (1994) reported that *Pseudomonas* sp. Strain 61-3 produces PHA comprised of monomer units of 3-hydroxyalkenoic acids such as 3-hydroxy-n-butyric acid, 3-hydroxyhexanoic acid, 3-hydroxyoctanoic acid, and 3-hydroxydecanoic acid, and 3-hydroxyalkenoic acids such as 3-hydroxy-5-cis-decenoic acid and 3-hydroxy-5-cis-dodecenoic acid, using sodium gluconate as a sole carbon source.

The above-described PHAs are PHAs comprised of monomer units having alkyl groups as the side chain (hereinafter also called usual-PHAs), or analogs thereof (for example, including, an alkenyl group having a double bond on a side chain other than at the end portion). However, when wider application of PHA, e.g., as a functional polymer, is intended, PHA having side chains including a substituent other than

an alkyl group (for example, a phenyl group, an unsaturated hydrocarbon, an ester group, an allyl group, a cyano group, a halogenated hydrocarbon, or an epoxide) is extremely useful (unusual-PHA).

5 As for biosynthesis of unusual-PHA having a phenyl group, it is reported that *Pseudomonas oleovorans* produces PHA having 3-hydroxy-5-phenylvaleric acid units from 5-phenylvaleric acid (Macromolecules, 24, 5256-5260 (1991), Macromol. Chem., 191, 1957-1965 (1990), Chirality, 3, 492-494 (1991)). Also Macromolecules, 29, 1762-1766 (1996) reports that *Pseudomonas oleovorans* produces PHA having 3-hydroxy-5-(4-tolyl)valeric acid units from 5-(4-tolyl)valeric acid (5-(4-methylphenyl)valeric acid). Further, Macromolecules, 32, 2889-2895 (1999) reports that *Pseudomonas oleovorans* produces PHA having 3-hydroxy-5-(2,4-dinitrophenyl)valeric acid units and 3-hydroxy-5-(4-nitrophenyl)valeric acid units from 5-(2,4-dinitrophenyl)valeric acid.

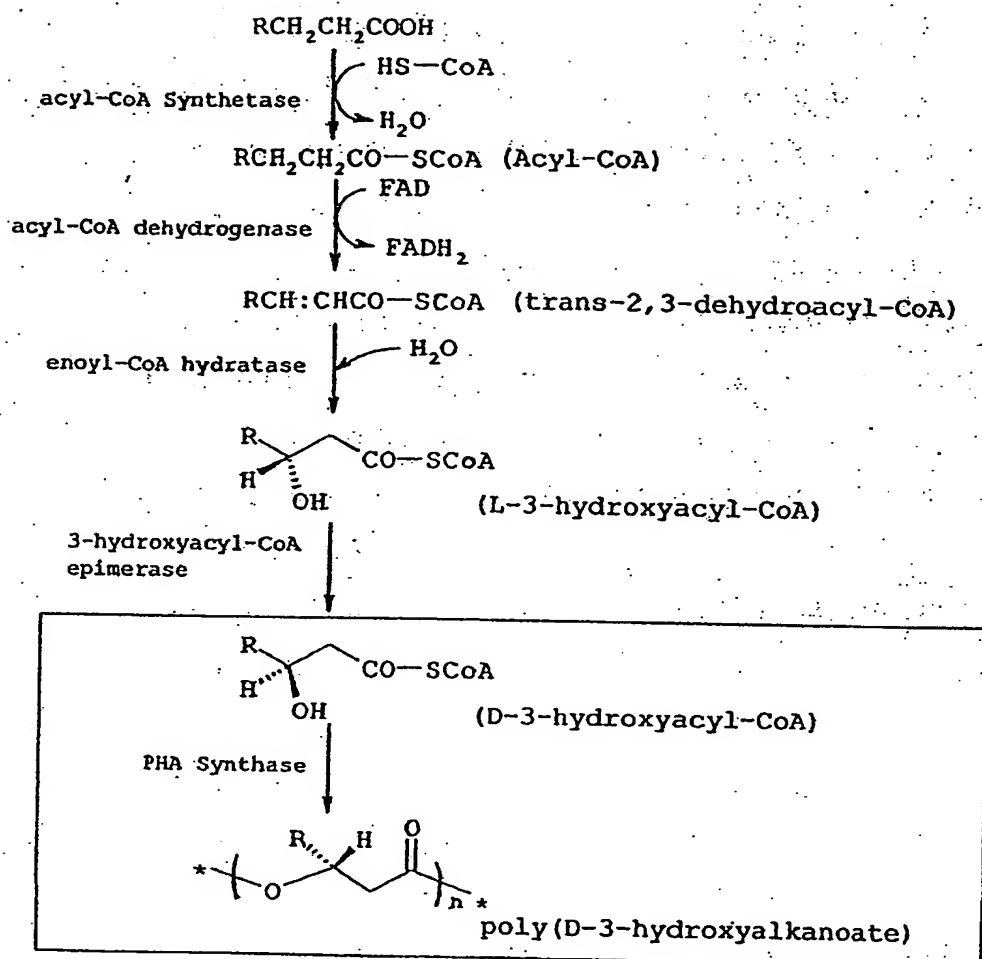
20 As for unusual-PHA having a phenoxy group, Macromol. Chem. Phys., 195, 1665-1672 (1994) reports that *Pseudomonas oleovorans* produces PHA having a 3-hydroxy-5-phenoxyvaleric acid unit and a 3-hydroxy-9-phoxynonanoic acid unit from 11-phenoxyundecanoic acid. Also, Macromolecules, 29, 3432-3435 (1996) reports that *Pseudomonas oleovorans* produces a PHA having a 3-hydroxy-4-phenoxybutyric acid unit and a

3-hydroxy-6-phenoxyhexanoic acid unit from 6-phenoxyhexanoic acid, a PHA having a 3-hydroxy-4-phenoxybutyric acid unit, a 3-hydroxy-6-phenoxyhexanoic acid unit, and a 3-hydroxy-8-phenoxyoctanoic acid unit from 8-phenoxyoctanoic acid, and a PHA having a 3-hydroxy-5-phenoxyvaleric acid unit and a 3-hydroxy-7-phenoxyheptanoic acid unit from 11-phenoxyundecanoic acid. Further, Can. J. Microbiol., 41, 32-43 (1995) reports that *Pseudomonas* oleovorans ATCC 29347 and *Pseudomonas putida* KT 2442 produce PHA having a 3-hydroxy-p-cyanophenoxyhexanoic acid unit and PHA having a 3-hydroxy-p-nitrophenoxyhexanoic acid unit from p-cyanophenoxyhexanoic acid and p-nitrophenoxyhexanoic acid respectively. Japanese Patent No. 2989175 describes a homopolymer comprised of a 3-hydroxy-5-(monofluorophenoxy)valeric acid unit or a 3-hydroxy-5-(difluorophenoxy)valeric acid unit and a copolymer containing at least a 3-hydroxy-5-(monofluorophenoxy)pentanoate unit or a 3-hydroxy-5-(difluorophenoxy)pentanoate unit and a method for producing such homopolymer or copolymer, reciting that such homopolymer and copolymer can provide water-repellency and stereoregularity maintaining high melting point and good workability.

As an example of unusual-PHA having a cyclohexyl group, Macromolecules, 30, 1611-1615 (1997) reports

that *Pseudomonas oleovorans* produces such PHA from cyclohexylbutyric acid or cyclohexylvaleric acid.

These mcl-PHA and unusual-PHA are synthesized through an enzymatic polymerization reaction using (R)-3-hydroxyacyl CoA as a substrate. (R)-3-hydroxyacyl CoA is generated through metabolic pathways such as β -oxidation pathway. The enzyme that catalyzes this polymerization reaction is called, in the present invention, a PHA synthetase. In the present invention, the aforescribed scl-PHA synthetase and the mcl-synthetase are collectively called PHA synthetase, but the mcl-PHA synthetase is often called PHA synthetase (also called PHA polymerase or PHA synthase). The following is the reaction route from alkanoic acid to PHA via the β -oxidation pathway and polymerization reaction by PHA synthetase.



On the other hand, when the production is performed through the fatty acid synthesis pathway, it is considered that (R)-3-hydroxyacyl-ACP (ACP means acyl carrier protein) generated in this pathway is converted to (R)-3-hydroxyacyl CoA from which PHA is synthesized by PHA synthetase.

<In vitro PHA synthesis utilizing enzyme>

10 Recently, attempts have been made to synthesize

PHA *in vitro* using PHA synthetase (scl-PHA synthetase or mcl-PHA synthetase) isolated from cells.

For example, Proc. Natl. Acad. Sci. USA, 92, 6279-6283 (1995) describes that PHB comprised of 3-hydroxy-n-butyric acid units is successfully synthesized from 3-hydroxybutyryl CoA by the action of scl-PHA synthetase derived from *Alcaligenes eutrophus*. In addition, Int. J. Biol. Macromol., 25, 55-60 (1999) describes that PHA comprised of a 3-hydroxy-n-butyric acid unit or a 3-hydroxy-n-valeric acid unit is successfully synthesized from 3-hydroxybutyryl CoA or 3-hydroxyvaleryl CoA using scl-PHA synthetase derived from *Alcaligenes eutrophus*. Further, this report mentions that, when racemic 3-hydroxybutyryl CoA is reacted, PHB comprised of only (R)-3-hydroxy-n-butyric acid units is successfully synthesized due to the stereoselectivity of the enzyme. Macromol. Rapid Commun., 21, 77-84 (2000) reports *in vitro* PHB synthesis using scl-PHA synthetase derived from *Alcaligenes eutrophus*.

FEMS Microbiol. Lett., 168, 319-324 (1998) describes that PHB comprised of a 3-hydroxy-n-butyric acid unit is successfully synthesized by reacting PHB synthetase derived from *Chromatium vinosum* on 3-hydroxybutyryl CoA.

In Appl. Microbiol. Biotechnol., 54, 37-43 (2000), PHA comprised of 3-hydroxydecanoic acid is

synthesized by reacting PHA synthetase derived from *Pseudomonas aeruginosa* on 3-hydroxydecanoyl CoA.

In addition to the foregoing, the present invention cites descriptions of Japanese Patent No. 2989175, Japanese Patent Application Laid-Open Nos. 2001-78753 and 2001-69968, Eur. J. Biochem., 250, 432-439 (1997), J. Biol. Chem., 218, 97-106 (1956), J. Amer. Chem. Soc., 78, 2278 (1956), Appl. Environ. Microbiol., 44, 238-241 (1982), Molecular Cloning, vol.1, 572, 1989 (Cold Spring Harbor Laboratory), J. Bacteriol., 182, 2753-2760 (2000), and Int. J. Biol. Macromol., 12, 85-91 (1990).

Meanwhile, for separation/recovery, detection and screening of a target component contained in a specimen, particularly a target component effective for medical treatment or diagnosis or useful industrially, there are being developed and utilized various methods of separation/recovery, detection and screening, utilizing fine particles of a size of micrometer to nanometer as a carrier for a molecule having a coupling affinity to the target component, such as a probe molecule. In particular, a method utilizing fine particles having a magnetic property (hereinafter called magnetic particles) as the aforescribed carrier has an advantage that, at the separation or recovery of the carrier from the specimen, the magnetic particles can be easily

separated or recovered by a magnetic force. For this reason, many developments have been made in the method utilizing the magnetic particles.

The magnetic particles, utilized as the carrier
5 for immobilizing the aforescribed probe molecule or the like, are mostly used in a state surfacially coated with an organic polymer, for stability improvement and control of magnetic property.

As an application of the magnetic particles
10 coated with the organic polymer as such a carrier, immunoassay utilizing an antigen-antibody reaction has been developed.

As an example, Japanese Patent Application Laid-Open No. H07-151755 discloses an immunoassay
15 utilizing magnetic material-containing polystyrene latex of an average particle size of 0.7 μm , manufactured by Rhone Poulenc.

Also Japanese Patent Application Laid-Open No. H10-221341 discloses an immunological measuring
20 method utilizing tosylated magnetic particles (Dynabead M-280, average particle size 2.8 μm), manufactured by Nippon Dynal Co. Also Japanese Patent Application Laid-Open No. H09-229936 discloses an immunological assay method and an apparatus,
25 utilizing magnetic particles Dynabeads M-450 uncoated, manufactured by Dynal Inc., of a particles size of 4.5 μm , 3% (w/v).

Another application area of the magnetic particles coated with the organic polymer as a carrier is an inspection/diagnosis method for nucleic acid molecules such as DNA.

5 Japanese Patent Application Laid-Open No. H05-281230 discloses an antigen-antibody reaction and an inspection/diagnosis method for nucleic acid molecules such as DNA, utilizing, as magnetic carrier particles, XP-600 manufactured by Dino Industrier
10 A.S., Norway.

In addition, as still another application area of such magnetic particles, there are developed methods for separating and recovering a target component.

15 Japanese Patent Application Laid-Open No. H09-304385 discloses a method and an apparatus for separation and recovery of basophilic cells, utilizing Dynabeads M-450 uncoated (Dynal Inc., particle size of magnetic particles: 45 μm).

20 Japanese Patent Application Laid-Open No. H10-068731 discloses a method for magnetically separating an object component in liquid, utilizing magnetic particles manufactured by Rhone-Poulenc, as magnetic particles to which an immunologically active
25 substance or nucleic acid is covalently bonded.

Also U.S. Patent Nos. 4,230,685, 3,970,518, 5,508,164, 5,567,326 and 4,018,886 discloses methods

for using magnetic particles to bind a target component thereto and separating the target component bonded with the magnetic particles.

U.S. Patent No. 5,900,481 discloses a method of binding DNA using coated magnetic particles to treat the DNA.

U.S. Patent No. 5,834,197 discloses a method of capturing a certain bacterial strain from liquid, utilizing coated magnetic particles, where a labeled antibody having a selective affinity to an antibody is attached to the beads thereby coupling the detectable label to the magnetic particles, in order to achieve easy detection and recovery of the antigen reacted with the labeled antibody.

In addition to the aforescribed patent references, there are references relating to manipulation of various molecules to be bonded to the magnetic particles, such as Analytical Chemistry, 68(13), 2121-6 (1996) and Nucleic Acids Research 23(16), 3126-31 (1995).

Also in addition to the magnetic particles described in the foregoing, there are magnetic particles already commercialized for use in methods of detecting, recovering or screening a target component, such as Ferromagnetic Particles from Spherotech Inc., Cera-Mag from Seradyn Inc. and Esteapor from Bangs Laboratory Inc.

DISCLOSURE OF THE INVENTION

As explained above, an application of a bioengineering method to the synthesis of a polymer compound is expected to enable synthesis of a novel polymer compound or endowment of novel function or structure that have been difficult to realize in the conventional organosynthetic methods. Also a biological process may often be one step process where conventional organosynthetic methods require multi-steps, and there are expected process simplification, cost reduction, time reduction etc. It is also rendered possible to reduce amounts of organic solvent, acid, alkali, surfactant etc., to employ milder reaction conditions and to achieve synthesis from a non-petroleum raw material or a low-purity raw material, thereby realizing a synthesizing process of a lower environmental burden and a resource recycling type. With respect to the synthesis from a low-purity raw material, the bioengineering synthetic process can carry out the desired reaction even with a raw material of a low purity because the enzyme, functioning as a catalyst, generally has a high substrate specificity, so that the utilization of a wasted material or a recycled raw material can also be expected.

On the other hand, as explained in the foregoing, the present inventors have investigated a construct

in which a magnetic material is coated with a polymer compound, as an elementary technology for providing a polymer compound of a high added value. Such coating of a specified magnetic material with a polymer compound can provide a composite construct having an extremely useful functionality. Various attempts have been made to produce such a construct through organosynthetic methods, but such methods have certain limitations.

10 If such a construct can be prepared by a bioengineering method, it is expected to realize utilization of a novel polymer compound and endowment of novel functions and structures, which has not been realized in the conventional organosynthetic methods, and also to realize a manufacturing process of a lower environmental burden and a resource recycling type with a lower cost. For example, based on extremely strict molecular recognition and stereospecificity inherent to the biological catalytic action, it is possible to produce a polymer compound of a novel functionality that has been difficult to realize in the conventional organosynthetic methods, or a capsule construct or a multi-layered construct coated with a polymer compound of an extremely high chirality, by an extremely simple process of a low environmental burden..

Therefore, an object of the present invention is to provide a polymer compound construct of a high functionality produceable by a bioengineering process.

The invention also provides an efficient method for producing a construct, formed by coating a magnetic material with a polymer compound and usable in various fields as a functional composite construct.

In particular, the invention provides a construct of a coated magnetic material without oleophilic treatment on a metal or a metal compound having magneticity and with an excellent uniformity in dispersion, and a manufacturing method therefor.

As explained in the foregoing, the magnetic material-containing capsule construct obtained by conventional synthetic methods has a drawback of elution of metal ions to the exterior and is currently usable only in the fields and applications where the metal ion elution does not matter. The invention provides a magnetic material-containing capsule construct that is excellent in dispersibility and magnetic response, and hardly causes elution of the metal ions to the outside, thereby being widely applicable in various fields and application, and a producing method therefor.

Also in the method for separation/recovery, detection or screening of a target component, since such a target component often has a physiological

activity or a medical/diagnostic efficacy, it is desirable to conduct the operations in an environment as close as possible to in vivo conditions, so as not to deteriorate the physiological properties. However, in the aforedescribed methods employed in the conventional carrier including the magnetic particles, since the coating layer on the surface of the magnetic particles is made of a synthetic polymer such as styrenic, acrylic or vinylic, there still remain drawbacks of a non-specific adsorption and a loss in the functionality resulting from a leakage of a monomer component remaining in the synthetic polymer. Also, not only the target component but also a target-binding molecule, to be borne and immobilized on the carrier including the magnetic particles are often substances obtained or derived from a living organism, and the coupling ability of such a target-binding molecule with the target component may be detrimentally affected by the non-natural polymer surface employed in the coating layer of the carrier containing the magnetic particles.

The present invention is to resolve these drawbacks, and provides, in executing separation/recovery, detection or screening of a target component utilizing a construct formed by bearing and immobilizing a target-binding molecule on a carrier, a method of executing separation/recovery,

detection or screening of the target component while maintaining the target component and the target-binding molecule borne and immobilized on the carrier in a condition close to in vivo condition.

5 Also the invention relates to a construct in which at least a part of magnetic material is coated with polyhydroxyalkanoate.

Also the invention relates to a method for producing a construct formed by a magnetic material
10 of which at least a part is coated with polyhydroxyalkanoate, the method being characterized in including a step of immobilizing a polyhydroxyalkanoate synthetase on a surface of the magnetic material and a step of polymerizing 3-
15 hydroxyacyl co-enzyme A by such an enzyme thereby synthesizing polyhydroxyalkanoate, whereby such a synthesizing step coats at least a part of the magnetic material with polyhydroxyalkanoate.

Furthermore, the invention relates to a method
20 for separating a target component contained in a specimen, characterized in including a step of preparing a carrier in which a molecule having a coupling affinity to the target component is immobilized on a surface, a step of mixing the
25 carrier and the specimen, a step of coupling the target component contained in the specimen, to be mixed in the mixing step, with the molecule

immobilized on the carrier surface and having the coupling affinity, and a step of separating the target component, immobilized to the carrier in the coupling step through the coupling with the molecule having the coupling affinity, together with the carrier from the specimen, wherein the carrier is at least partly coated with polyhydroxyalkanoate.

Furthermore, the invention relates to a method for detecting a target component contained in a specimen, characterized in including a step of preparing a carrier in which a molecule having a coupling affinity to the target component is immobilized on a surface, a step of mixing the carrier and the specimen, a step of coupling the target component contained in the specimen, to be mixed in the mixing step, with the molecule immobilized on the carrier surface and having the coupling affinity, and a step of selectively detecting the target component, immobilized to the carrier in the coupling step through the coupling with the molecule having the coupling affinity, wherein the carrier is at least partly coated with polyhydroxyalkanoate.

Furthermore, the invention relates to a method for screening a target component contained in a medium, for a mixed specimen containing a mixture including the target component in the medium,

characterized in including a step of preparing a carrier in which a molecule having a coupling affinity to the target component is immobilized on a surface, a step of mixing the carrier and the specimen, a step of coupling the target component contained in the specimen, to be mixed in the mixing step, with the molecule immobilized on the carrier surface and having the coupling affinity, and a step of separating the target component, immobilized to the carrier in the coupling step through the coupling with the molecule having the coupling affinity, together with the carrier from the mixed specimen, wherein the carrier is at least partly coated with polyhydroxyalkanoate.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 is a gas chromatography-mass spectroscopy chart of a methyl esterified substance derived from a 3-hydroxyoctanoic acid unit of PHA, identified in Example 11;

Fig. 2 is a gas chromatography-mass spectroscopy chart of a methyl esterified substance derived from a 3-hydroxy-5-phenylvaleric acid unit of PHA, identified in Example 14;

Fig. 3 is a gas chromatography-mass spectroscopy chart of a methyl esterified substance derived from a 3-hydroxy-5-(4-fluorophenyl)valeric acid unit of PHA,

identified in Example 15;

Fig. 4 is a gas chromatography-mass spectroscopy chart of a methyl esterified substance derived from a 3-hydroxyoctanoic acid unit contained in PHA and

5 identified in Example 26;

Fig. 5 is a schematic view showing a configuration of a magnetic capsule construct bearing a target-binding molecule on a PHA being a surface coating of a carrier (magnetic material);

10 Fig. 6 is a view schematically showing a selective binding-forming process between the target component and the target-binding molecule on the magnetic capsule construct bearing the target-binding molecule; and

15 Fig. 7 is a view schematically showing a magnetic separation process of the invention for the magnetic capsule construct bearing the target-binding molecule and forming a selective binding with the target component, based on magneticity of the
20 construct.

BEST MODE FOR CARRYING OUT THE INVENTION

<<Construct>>

The construct of the present invention has a
25 configuration in which a magnetic material is coated with PHA including a monomer unit of various structures having a substituent in a side chain, and

is extremely useful as a carrier for separating, purifying or screening microorganisms, cells, nucleic acid, protein or other biological substances, a carrier for a medical diagnostic agent enabling displacement control in a living organism, a carrier for drug delivery for carrying a drug to a diseased part of a patient, a carrier for immobilizing an enzyme, or a functional carrier for a magnetic toner, a magnetic ink, a magnetic paint or a magnetic recording medium. Now the present invention will be detailedly explained in the following.

<PHA>

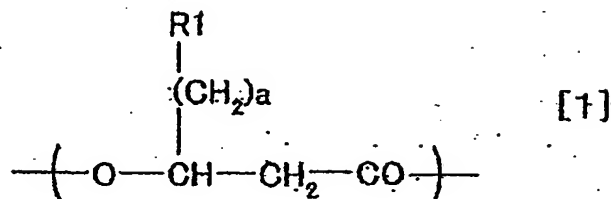
Polyhydroxyalkanoate usable in the present invention includes a short-chain-length polyhydroxyalkanoate (short-chain-length PHA, hereinafter also represented as scl-PHA), of which monomer units are constituted of a 3-hydroxyalkanoic acid unit with 4 or 5 carbon atoms; and a medium-chain-length polyhydroxyalkanoate (hereinafter also represented as mcl-PHA), of which monomer units include not only a polyhydroxybutyrate (PHB) with 4 carbon atoms or polyhydroxyvalerate (PHV) with 5 carbon atoms, but also a 3-hydroxyalkanoic acid unit with about 6 to 12 carbon atoms. In addition to the aforescribed polyhydroxyalkanoate constituted of a monomer unit having an alkyl group in the side chain (hereinafter also represented as usual-PHA), the

present invention can utilize a polyhydroxyalkanoate including a monomer unit in which various substituents (such as a phenyl group, an unsaturated hydrocarbon group, an ester group, an aryl group, a cyano group, a halogenated hydrocarbon group, or an epoxy (-O-) group) other than an alkyl group are introduced in the side chain in consideration of applications in wider fields such as a functional polymer (such PHA being hereinafter represented also as unusual-PHA), or a copolymer including these monomer units in an arbitrary unit ratio.

PHA to be employed in the method of the invention is not particularly restricted as long as it is synthesizable by PHA synthetase (for example, mcl-PHA and unusual-PHA described above). As explained in the foregoing, the PHA synthetase is an enzyme catalyzing a final step in a PHA synthesizing reaction system in vivo, and any PHA known to be synthesized in vivo is synthesized by the catalytic action of such an enzyme. In the invention, it is therefore possible, by reacting a 3-hydroxyacyl CoA corresponding to a desired PHA with the PHA synthetase immobilized to the magnetic material, to prepare a construct in which the magnetic material coated with any PHA known to be synthesized in vivo.

Specific examples of such PHA include PHA including at least monomer units represented by

following chemical formulae [1] to [10] and [A] to [D].



5 (wherein the monomer unit is at least one selected from a group of monomer units having following combinations of R1 and a:

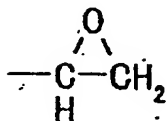
a monomer unit in which R1 is a hydrogen atom (H) and a is any of integers from 3 to 10;

10 a monomer unit in which R1 is a halogen atom and a is any of integers from 1 to 10;

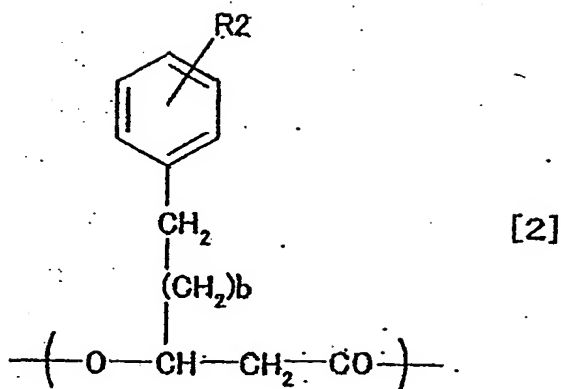
a monomer unit in which R1 is a chromophore and a is any of integers from 1 to 10;

15 a monomer unit in which R1 is a carboxyl group or a salt thereof and a is any of integers from 1 to 10; and

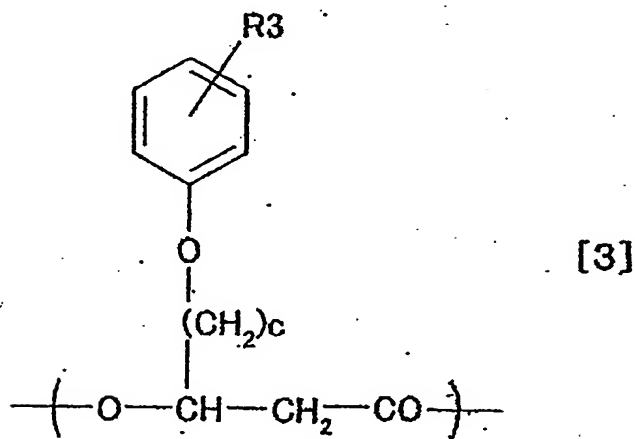
a monomer unit in which R1 is



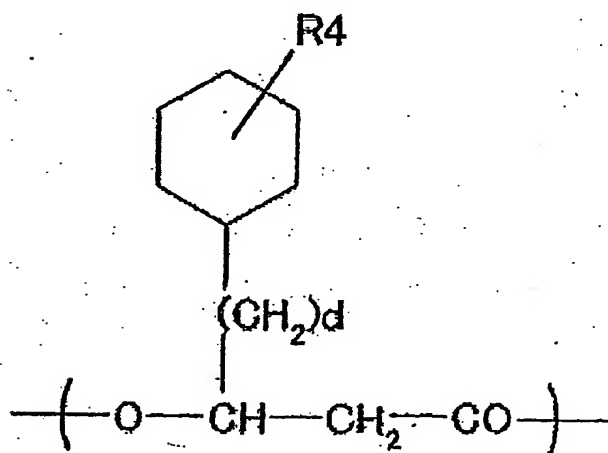
20 and a is any of integers from 1 to 7.)



(wherein b represents any of integers from 0 to 7; R2 represents any one selected from a group of a
 5 hydrogen atom (H), a halogen atom, -CN, -NO₂, -CF₃,
 -C₂F₅, -C₃F₇, a CH₃ group, a C₂H₅ group, a C₃H₇ group, a
 vinyl group, an epoxy group and COOR₂₁ (R₂₁ representing an H atom, an Na atom or a K atom); and,
 in the presence of plural units, the foregoing stands
 10 independently for each unit.)

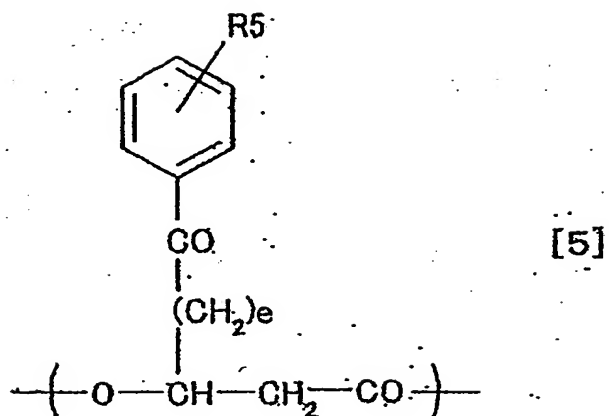


(wherein c represents any of integers from 1 to 8
 and R3 represents any one selected from a group of a
 hydrogen atom (H), a halogen atom, -CN, -NO₂, -CF₃,
 -C₂F₅, -C₃F₇, a CH₃ group, a C₂H₅ group, a C₃H₇ group
 5 and a SCH₃ group; and, in the presence of plural units,
 the foregoing stands independently for each unit.)

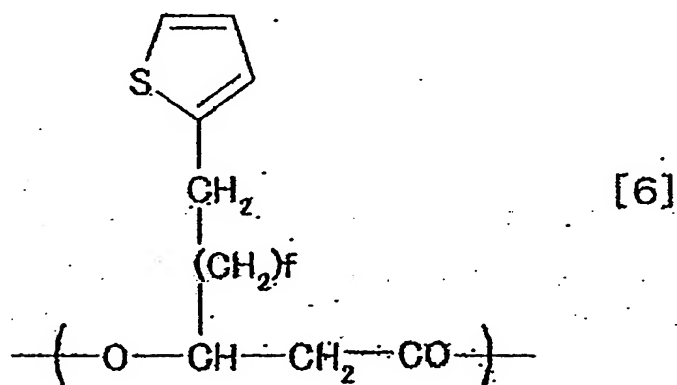


[4]

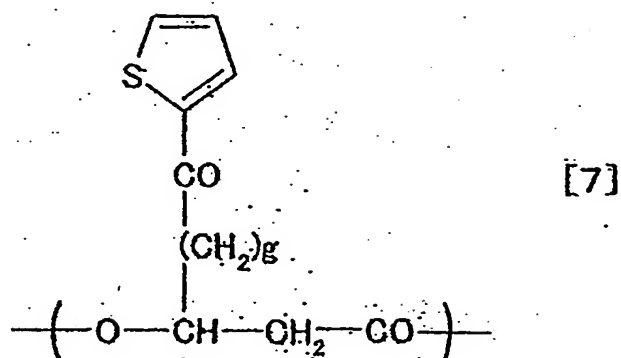
(wherein d represents any of integers from 0 to 8; R4
 10 represents any one selected from a group of an H atom,
 a CN group, a NO₂ group, a halogen atom, a CH₃ group,
 a C₂H₅ group, a C₃H₇ group, a CF₃ group, a C₂F₅ group,
 and a C₃F₇ group; and, in the presence of plural units,
 the foregoing stands independently for each unit.)



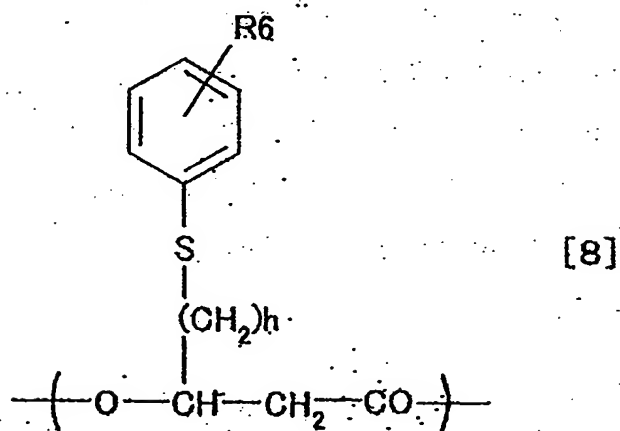
(wherein e represents any of integers from 1 to 8 and R5 represents any one selected from a group of a hydrogen atom (H), a halogen atom, -CN, -NO₂, -CF₃, -C₂F₅, -C₃F₇, -CH₃, -C₂H₅, and -C₃H₇.)



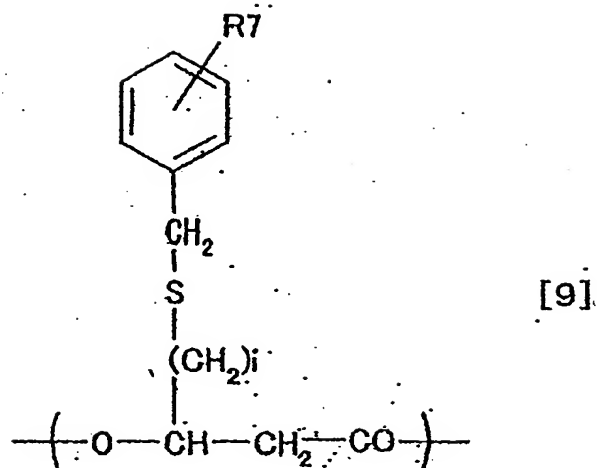
(wherein f represents any of integers from 0 to 7.)



(wherein g represents any of integers from 1 to 8).

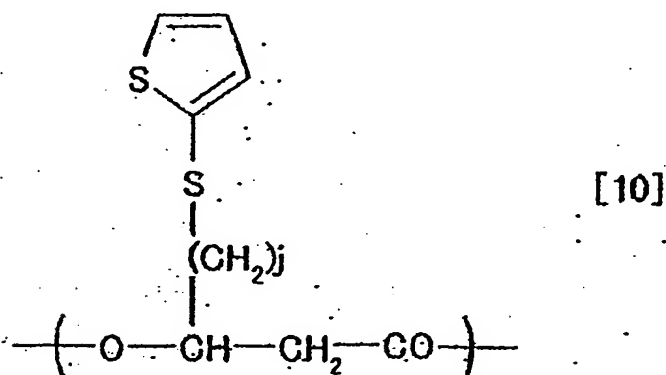


(wherein h represents any of integers from 1 to 7; R6 represents any one selected from a group of a hydrogen atom (H), a halogen atom, -CN, -NO₂, -COOR', -SO₂R'', -CH₃, -C₂H₅, -C₃H₇, -CH(CH₃)₂, and -C(CH₃)₃; R' represents a hydrogen atom (H), Na, K, -CH₃, or -C₂H₅; and R'' represents -OH, -ONa, -OK, a halogen atom, -OCH₃, or -OC₂H₅).

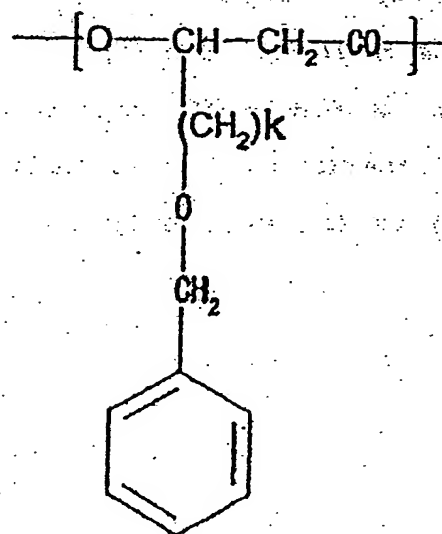


(wherein i represents any of integers from 1 to 7; R7 represents any one selected from a group of a

5 hydrogen atom (H), a halogen atom, -CN, -NO₂, -COOR', and -SO₂R''; R' represents a hydrogen atom (H), Na, K, -CH₃, or -C₂H₅; and R'' represents -OH, -ONa, -OK, a halogen atom, -OCH₃, or -OC₂H₅).

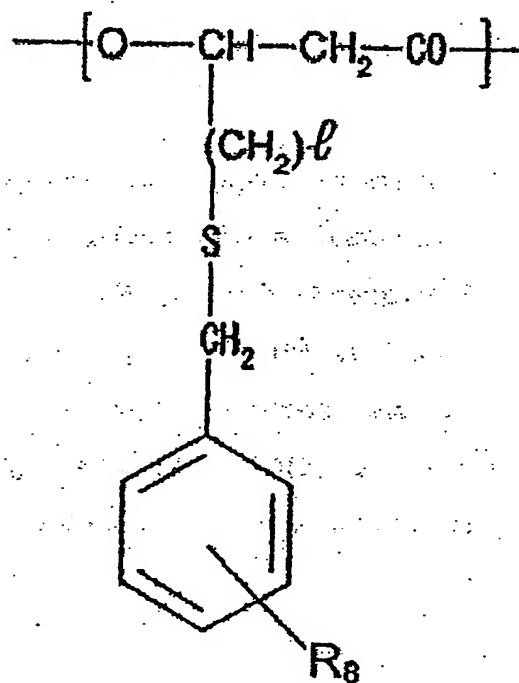


10 (wherein j represents any of integers from 1 to 9).



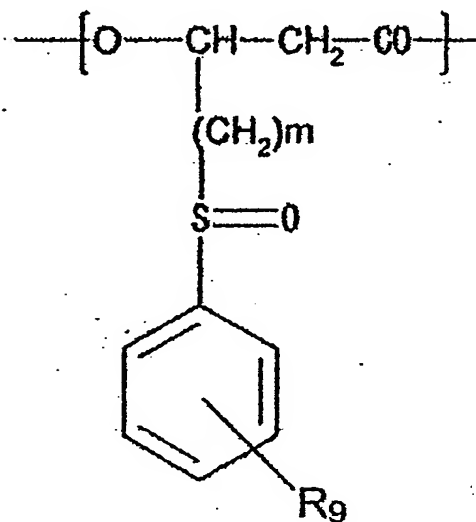
[A]

(wherein k represents any of integers from 1 to 8).



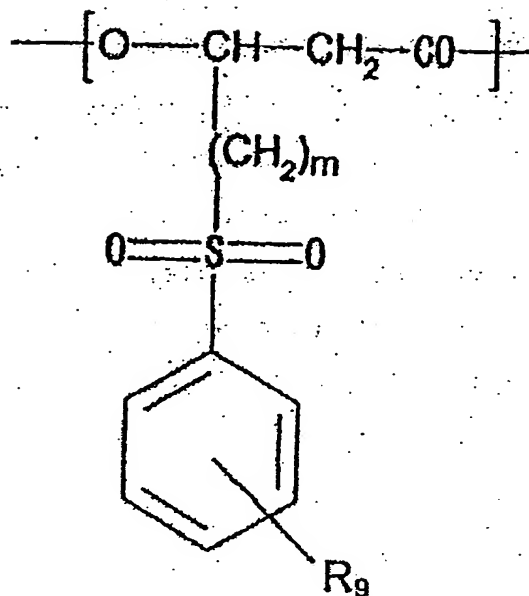
[B]

(wherein l represents any of integers from 1 to 8; R_8 represents any one selected from a group of a CH_3 group, a C_2H_5 group, a C_3H_7 group, a $(CH_3)_2-CH$ group and a $(CH_3)_3-C$ group; and, in the presence of plural units, the foregoing stands independently for each unit.)



[C]

(wherein m represents any of integers from 1 to 8; R_9 represents an H atom, a halogen atom, a CN group, a NO_2 group, $COOR_{91}$, SO_2R_{92} (R_{91} representing H, Na, K, CH_3 or C_2H_5 , and R_{92} representing OH, ONa, OK, a halogen atom, OCH_3 or OC_2H_5), a CH_3 group, a C_2H_5 group, a C_3H_7 group, a $(CH_3)_2-CH$ group or a $(CH_3)_3-C$ group; and, in the presence of plural units, the foregoing stands independently for each unit.)



[D]

(wherein m represents any of integers from 1 to 8; R_9 represents an H atom, a halogen atom, a CN group, a
 5 NO_2 group, COOR_{91} , SO_2R_{92} (R_{91} representing H, Na, K, CH_3 or C_2H_5 , and R_{92} representing OH, ONa, OK, a halogen atom, OCH_3 or OC_2H_5), a CH_3 group, a C_2H_5 group, a C_3H_7 group, a $(\text{CH}_3)_2\text{---CH}$ group or a $(\text{CH}_3)_3\text{---C}$ group; and, in the presence of plural units, the foregoing stands
 10 independently for each unit.)

Specific examples of the aforescribed halogen atom include fluorine, chlorine, and bromine. The chromophore is not particularly limited as long as the 3-hydroxyacyl CoA having the chromophore is
 15 catalyzed by PHA synthetase, but it is preferable that a methylene chain of 1 to 5 carbon atoms is

present between the chromophore and the carboxyl group to which CoA is bonded, in view of steric hindrance at the time of polymer synthesis. When the light absorption wavelength of the chromophore is within a visible range, a colored construct can be obtained and when the light absorption wavelength is outside the visible range, the construct may be used as various electronic materials. Examples of such a chromophore include nitroso, nitro, azo, diarylmethane, triarylmethane, xanthene, acridine, quinoline, methine, thiazole, indamine, indophenol, lactone, aminoketone, hydroxyketone, stilbene, azine, oxazine, thiazine, anthraquinone, phthalocyanine, and indigoid.

As PHA which is used for the present invention, a random copolymer or a block copolymer comprised of a plurality of the above described monomer units can be used. Therefore, it becomes possible to control physical properties of PHA and add some functions to the PHA by utilizing properties of each monomer unit or functional groups included therein, and also possible to express new functions due to interaction between functional groups.

In the present invention, it is also possible to utilize a homopolymer constituted of a 3-hydroxypropionic acid unit, a 3-hydroxy-n-butyric acid unit, a 3-hydroxy-n-valeric acid unit, a 4-

hydroxy-n-butyric acid unit, or a hydroxyalkanoic acid unit with 6 to 14 carbon atoms, or a copolymer constituted of plural kinds of these units. Also, if necessary, chemical modification or the like may be applied after or in the course of enzymatic synthesis of PHA.

Thus, for example, in case of forming a construct coated with a PHA of a low affinity to the magnetic material, it is possible to at first coat the magnetic material with a PHA of a high affinity thereto, and then to change the monomer unit composition of such PHA of a high affinity to the magnetic material to a monomer unit composition of a desired PHA in a direction from the interior to the exterior or in a vertical direction thereby forming a multi-layered structure or a gradient structure, whereby a PHA coating firmly bound to the magnetic material can be obtained.

In the present invention, it is also possible to utilize a PHA in which monomer units constituting a scl-PHA such as a 3-hydroxypropionic acid unit, a 3-hydroxy-n-butyric acid unit, a 3-hydroxy-n-valeric acid unit and a 4-hydroxy-n-butyric acid unit, and monomer units of mcl-PHA or unusual-PHA described above are mixedly present. Also, if necessary, chemical modification may be applied after or in the course of synthesis of PHA. The PHA preferably has a

number-averaged molecular weight from about 1,000 to 10,000,000.

In the monomer unit of the chemical formula [2], a monomer unit having a carboxyl group (COOR_{21}) as R_2 can be produced from a monomer unit represented by the chemical formula [2] and having a vinyl group as R_2 , namely having a vinylphenyl group at the end of the side chain, by a selective oxidation cleaving of the double bond of the vinyl group into a carboxyl group, whereby a PHA including a monomer unit represented by the chemical formula [2] and having a carboxyphenyl group at the end of the side chain thereof.

The aforescribed conversion from vinyl group to carboxyl group, namely for obtaining a carboxylic acid by oxidation cleaving of a carbon-carbon double bond with an oxidant is known in various methods such as a method of utilizing a permanganate (J. Chem. Soc., Perkin, Trans. 1, 806 (1973)), a method of utilizing a bichromate (Org. Synth., 4, 698 (1963)), a method of utilizing a periodide (J. Org. Chem., 46, 19 (1981)), a method of utilizing nitric acid (Japanese Patent Application Laid-Open No. S59-190945), a method of utilizing ozone (J. Am. Chem. Soc., 81, 4273 (1959)) etc., and, in relation to PHA, Macromolecular Chemistry, 4, 289-293 (2001) described before reports a method of obtaining a carboxylic

acid from a carbon-carbon double bond at the end of the side chain of PHA by a reaction under an acidic condition with potassium permanganate as the oxidant.

In executing a reaction of converting the

5 aforedescribed vinyl group into a carboxyl group in the invention, reference may be made to such an oxidation cleaving method.

In the oxidation cleaving reaction employing potassium permanganate as the oxidant, various

10 inorganic acids such as sulfuric acid, hydrochloric acid, acetic acid or nitric acid, or organic acids are employed for realizing an acidic condition in the reaction system. However an acid such as sulfuric acid, nitric acid or hydrochloric acid may cause a
15 cleaving of an ester bond in the main chain of PHA, thus resulting in a decrease in the molecular weight.

For this reason, an acidic condition is preferably realized with acetic acid. An amount of the acid added to the reaction system is usually selected

20 within a range of 0.2 to 200 mol. equivalent with respect to 1 mole of the monomer unit represented by the chemical formula [2] and having a vinyl group as R₂, preferably 0.4 to 100 mol. equivalent. An amount of acid added to the reaction system less than 0.2

25 mol. equivalent results in a low yield of the oxidation cleaving reaction, while an amount exceeding 200 mol. equivalent results in a generation

of by-products from the added acid, so that either case is undesirable. Also a crown ether may be employed for the purpose of accelerating the oxidation cleaving reaction. In such a case, the crown ether and a permanganate salt form a complex to increase the reaction activity. Examples of the crown ether employable for the aforescribed purpose include dibenzo-18-crown-6-ether, dicyclo-18-crown-6-ether, and 18-crown-6-ether. An amount of addition of the crown ether to the reaction system is selected within a range of 1.0 to 2.0 mol.equivalent with respect to 1 mole of permanganate salt, preferably 1.0 to 1.5 mol. equivalent.

In executing the aforescribed oxidation reaction in the invention, a construct coated with PHA including a unit represented by the chemical formula [2] and having a vinyl group as R₂, a permanganate salt and an acid may be collectively charged and reacted in the reaction system from the beginning, or individually charged continuously or intermittently into the reaction system. It is also possible to dissolve or suspend the permanganate salt only in the reaction system, and to add the construct covered with PHA and the acid into the reaction system either continuously or intermittently, or to suspend only the construct covered with PHA in the reaction system and to add the permanganate salt and

the acid into the reaction system either continuously or intermittently. It is furthermore possible to at first charge the construct covered with PHA and the acid and to add the permanganate salt either continuously or intermittently, or to at first charge the permanganate salt and the acid and to add the the construct covered with PHA either continuously or intermittently, or to at first charge the construct covered with PHA and the permanganate salt and to add the acid either continuously or intermittently.

In the oxidation cleaving reaction to be carried out on the construct covered with a PHA, including a unit represented by the chemical formula [2] and having a vinyl group as R₂, utilizing potassium permanganate as an oxidant, a reaction temperature is usually selected within a range of -20 to 40°C, preferably 0 to 30°C. A reaction speed depends on a stoichiometric ratio of the unit represented by the chemical formula [2] and having a vinyl group as R₂, and the permanganate salt, and on the reaction temperature, and a reaction time is selected according to a target ratio of conversion of the vinyl group into the carboxyl group, and is usually selected within a range of 2 to 48 hours in case such a target ratio is selected at about 100%.

It is also possible, in a unit represented by the chemical formula [1] and having a vinyl group as

R1, to convert a vinyl group at an end of the side chain into a carboxyl group, by applying a similar oxidation cleaving reaction.

Also PHA including a monomer unit represented by the chemical formula [C] or a monomer unit represented by the chemical formula [D] can be prepared by at first producing PHA including a monomer unit represented by the chemical formula [8] and then selectively oxidizing a sulfanyl group (-S-) thereof into a sulfinyl group (-SO-) or a sulfonyl group (-SO₂-). Such a selective oxidation of the sulfanyl group (-S-) can be attained, for example, by an oxidation with a peroxide, and, in such operation there can be employed any peroxide that can contribute to the oxidation of the sulfanyl group (-S-). In consideration of oxidation efficiency, influence on the skeleton of the main chain of PHA and on other monomer units contained therein, and simplicity of process, it is particularly preferable to employ a peroxide selected from a group of hydrogen peroxide, sodium percarbonate, m-chloroperbenzoic acid, performic acid and peracetic acid.

For example, as a peroxide for the oxidation of sulfanyl group (-S-), m-chloroperbenzoic acid (MCPBA) allows a stoichiometric oxidation of the sulfanyl group (-S-) present in the monomer unit represented

by the chemical formula [8], thereby facilitating control on the ratio of the monomer unit represented by the chemical formula [C] or that represented by the chemical formula [D]. Also because of mild
5 reaction conditions, there hardly occurs an unnecessary side-reaction such as a cleavage of the main chain skeleton of PHA or a crosslinking reaction of active sites.

As a general reaction condition for oxidizing a
10 sulfanyl group (-S-) to a sulfinyl group (-SO-), MCPBA is employed in a somewhat excess amount, specifically in an amount of 1.1 to 1.4 moles with respect to 1 mole of the monomer unit represented by the chemical formula [8] and having a sulfanyl group
15 (-S-) in PHA, and a reaction is carried out in chloroform at a temperature of 0 to 30°C. Within the aforescribed reaction conditions, the oxidation to sulfinyl group (-SO-) proceeds by about 90% of the theoretical value at a reaction time of about 10
20 hours, and by about 100% of the theoretical value at a reaction time of about 20 hours. Also for oxidizing all the sulfanyl group (-S-) into a sulfonyl group (-SO₂-), MCPBA is employed in an amount somewhat excess of 2 moles, specifically in an amount of 2.1
25 to 2.4 moles with respect to 1 mole of the monomer unit represented by the chemical formula [8] and having a sulfanyl group (-S-) in PHA, and a reaction

is carried out by selecting a solvent, a temperature and a time similarly as explained above. In such an oxidation process employing MCPBA as the oxidant, a molecule of MCPBA acts on the sulfanyl group (-S-) to convert it into a sulfinyl group (-SO-), and another molecule of MCPBA acts on the sulfinyl group (-SO-) to convert it into a sulfonyl group (-SO₂-), but the conversion from sulfanyl (-S-) to sulfinyl (-SO-) has a higher reaction activity than in the conversion from sulfinyl (-SO-) to sulfonyl (-SO₂-).

Also a monomer unit represented by the chemical formula [2] and having an epoxy group as R₂ can be produced from a monomer unit represented by the chemical formula [2] and having a vinyl group as R₂, by a selective oxidation cleaving of the double bond of the vinyl group in a vinylphenyl group at the end of the side chain, thereby introducing an epoxy group. Thus, a PHA including a monomer unit represented by the chemical formula [2] and having a vinyl group as R₂ is subjected to a selective oxidation on the vinyl group to provide a PHA represented by the chemical formula [2] and having a 1,2-epoxyethyl group as R₂.

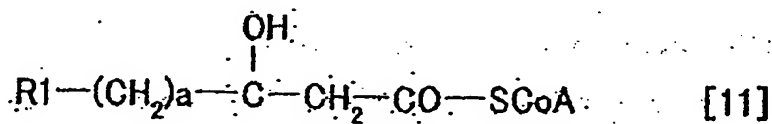
Also in such an oxidation process of epoxylation from the vinyl group to the epoxy group, a peroxide can be utilized, and there can be utilized any peroxide that can contribute to a selective partial oxidation of the vinyl group. In consideration of

oxidation efficiency, influence on the skeleton of the main chain of PHA and on other monomer units contained therein, and simplicity of process, it is particularly preferable to employ a peroxide selected from a group of hydrogen peroxide, sodium percarbonate, m-chloroperbenzoic acid, performic acid and peracetic acid. In case of employing a peroxide in the epoxylating oxidation from the vinyl group to the epoxy group, reaction conditions can refer to those in the aforescribed selective partial oxidation of the sulfanyl group with the peroxide.

The PHA employed in the construct of the present invention, synthesized by PHA synthetase, is usually an isotactic polymer comprised of R bodies alone.

<3-hydroxyacyl CoA>

3-hydroxyacyl CoA usable as the substrate for PHA synthetase in the present invention can be, as the substrate for scl-PHA synthetase, 3-hydroxypropionyl CoA, 3-hydroxybutyryl CoA or 3-hydroxyvaleryl CoA, and, as the substrate for mcl-PHA synthetase, 3-hydroxyacyl CoAs represented by following chemical formulas [11] to [20] and [A'] to [D'].



(wherein -SCoA represents a coenzyme A bonded to
alkanoic acid, in which a combination of R1 and a is
at least one selected from a following group,
corresponding to the combination of R1 and a in the
5 monomer unit represented by the chemical formula [1]:

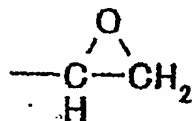
a monomer unit in which R1 is a hydrogen atom
(H) and a is any of integers from 3 to 10;

a monomer unit in which R1 is a halogen atom and
a is any of integers from 1 to 10;

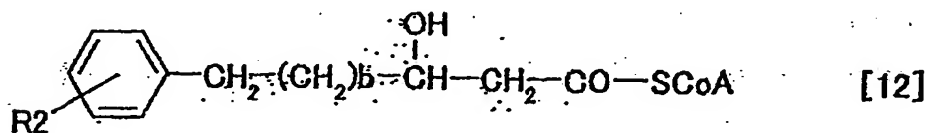
10 a monomer unit in which R1 is a chromophore and
a is any of integers from 1 to 10;

a monomer unit in which R1 is a carboxyl group
or a salt thereof and a is any of integers from 1 to
10; and

15 a monomer unit in which R1 is



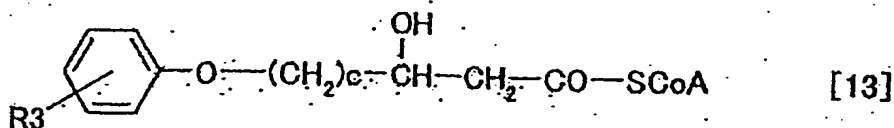
and a is any of integers from 1 to 7.)



20

(wherein -SCoA represents a coenzyme A bonded to
alkanoic acid; b corresponds to b in the monomer unit
represented by the aforescribed chemical formula

[2] and represents any of integers from 0 to 7; and
 R2 corresponds to R2 in the monomer unit represented
 by the chemical formula [2] and represents a hydrogen
 atom (H), a halogen atom, -CN, -NO₂, -CF₃, -C₂F₅, -C₃F₇,
 5 a CH₃ group, a C₂H₅ group, a C₃H₇ group, a vinyl group,
 an epoxy group or COOR₂₁ (R₂₁ representing an H atom,
 an Na atom or a K atom)).



10

(wherein -SCoA represents a coenzyme A bonded to
 alkanolic acid; c corresponds to c in the monomer unit
 represented by the aforescribed chemical formula

[3] and represents any of integers from 1 to 8; and

15

R3 corresponds to R3 in the monomer unit represented
 by the aforescribed chemical formula [3] and

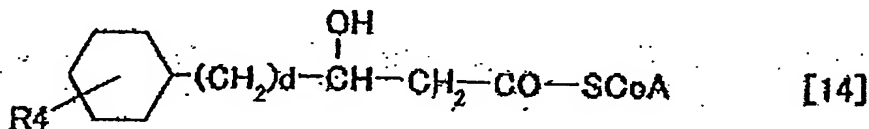
represents any one selected from a group of a

hydrogen atom (H), a halogen atom, -CN, -NO₂, -CF₃,

-C₂F₅, -C₃F₇, a CH₃ group, a C₂H₅ group, a C₃H₇ group

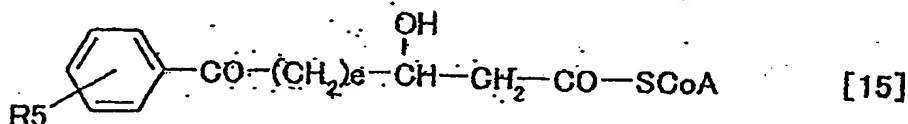
20

and a SCH₃ group.)



(wherein -SCoA represents a coenzyme A bonded to

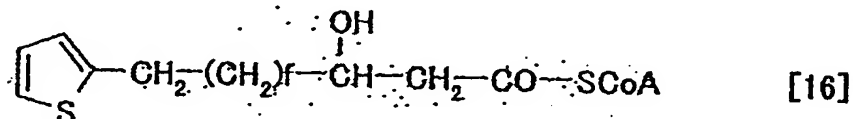
alkanoic acid; d corresponds to d in the monomer unit represented by the aforescribed chemical formula [4] and represents any of integers from 0 to 8; and R4 represents any one selected from a group of an H atom, a CN group, a NO₂ group, a halogen atom, a CH₃ group, a C₂H₅ group, a C₃H₇ group, a CF₃ group, a C₂F₅ group, and a C₃F₇ group.)



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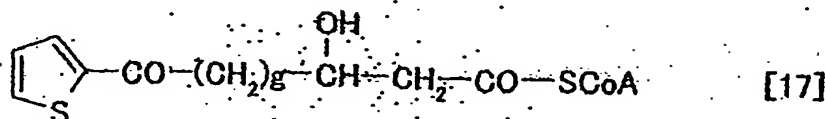
(wherein -SCoA represents a coenzyme A bonded to alkanoic acid; e corresponds to e in the monomer unit represented by the aforescribed chemical formula [5] and represents any of integers from 1 to 8; and R5 corresponds to R5 in the monomer unit represented by the aforescribed chemical formula [5] and represents any one selected from a group of a hydrogen atom (H), a halogen atom, -CN, -NO₂, -CF₃, -C₂F₅, -C₃F₇, -CH₃, -C₂H₅, and -C₃H₇.)

20

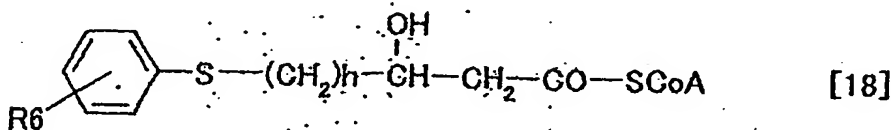


(wherein -SCoA represents a coenzyme A bonded to alkanoic acid; and f corresponds to f in the monomer

unit represented by the aforescribed chemical formula [6] and represents any one of integers from 0 to 7.)

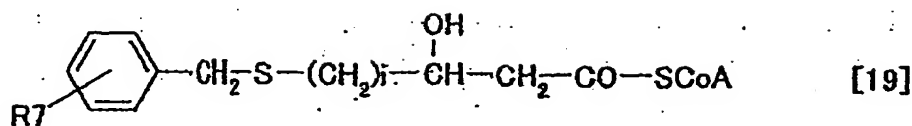


(wherein, -SCoA represents a coenzyme A bonded to alkanolic acid; and g corresponds to g in the monomer unit represented by the aforescribed chemical formula [7] and represents any one of integers from 1 to 8.)

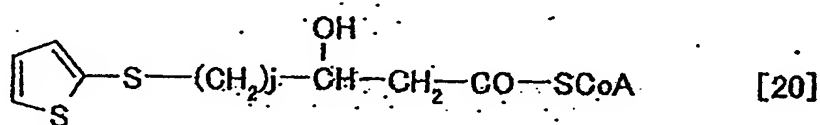


(wherein, -SCoA represents a coenzyme A bonded to alkanolic acid; h corresponds to h in the monomer unit represented by the aforescribed chemical formula [8] and represents any of integers from 1 to 7; R₆ corresponds to R₆ in the monomer unit represented by the aforescribed chemical formula [8] and represents any one selected from a group of a hydrogen atom (H), a halogen atom, -CN, -NO₂, -COOR', -SO₂R'', -CH₃, -C₂H₅, -C₃H₇, -CH(CH₃)₂, and -C(CH₃)₃; R' represents a hydrogen atom (H), Na, K, -CH₃, or -C₂H₅; and R'' represents -OH, -ONa, -OK, a halogen atom,

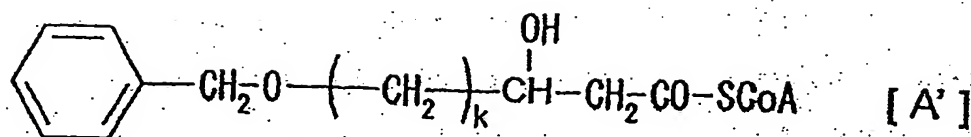
-OCH₃, or -OC₂H₅).



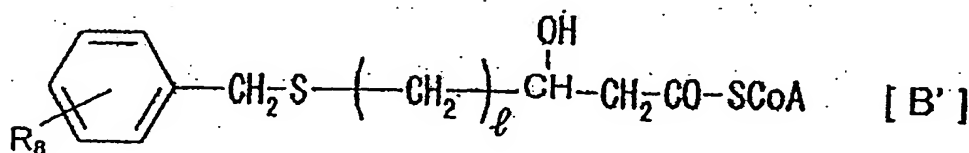
(wherein, -SCoA represents a coenzyme A bonded to
 5 alkanoic acid, i corresponds to i in the monomer unit
 represented by the aforescribed chemical formula
 [9] and represents any of integers from 1 to 7; R7
 corresponds to R7 in the monomer unit represented by
 the aforescribed chemical formula [9] and
 10 represents any one selected from a group of a
 hydrogen atom (H), a halogen atom, -CN, -NO₂, -COOR',
 and -SO₂R''; R' represents a hydrogen atom (H), Na, K,
 -CH₃, or -C₂H₅; and R'' represents -OH, -ONa, -OK, a
 15 halogen atom, -OCH₃, or -OC₂H₅).



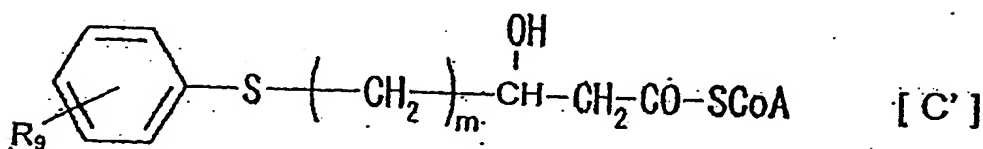
(wherein, -SCoA represents a coenzyme A bonded to
 alkanoic acid, and j corresponds to j in the monomer
 20 unit represented by the aforescribed chemical
 formula [10] and represents any of integers from 1 to
 9).



(wherein, -SCoA represents a coenzyme A bonded to alkanolic acid, and k corresponds to k in the monomer unit represented by the aforescribed chemical formula [A] and represents any of integers from 1 to 8).

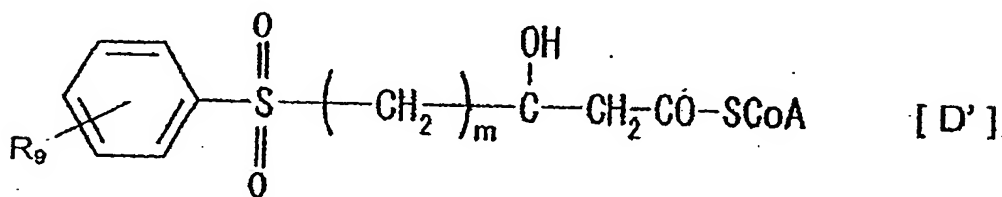


(wherein -SCoA represents a coenzyme A bonded to alkanolic acid; l corresponds to l in the monomer unit represented by the aforescribed chemical formula [B] and represents any of integers from 1 to 8; and R₈ corresponds to R₈ in the monomer unit represented by the aforescribed chemical formula [B] and represents any one selected from a group of a CH₃ group, a C₂H₅ group, a C₃H₇ group, a CF₃ group, a (CH₃)₂-CH group and a (CH₃)₃-C group.)



(wherein -SCoA represents a coenzyme A bonded to alkanolic acid; m corresponds to m in the monomer unit represented by the aforescribed chemical formula

5 [C] and represents any of integers from 1 to 8; and R9 corresponds to R9 in the monomer unit represented by the aforescribed chemical formulas [C] and [D] and represents an H atom, a halogen atom, a CN group, a NO₂ group, COOR₉₁, SO₂R₉₂ (R₉₁ representing H, Na, K, CH₃ or C₂H₅, and R₉₂ representing OH, ONa, OK, a
10 halogen atom, OCH₃ or OC₂H₅), a CH₃ group, a C₂H₅ group, a C₃H₇ group, a (CH₃)₂-CH group or a (CH₃)₃-C group.)



15

(wherein -SCoA represents a coenzyme A bonded to alkanolic acid; m corresponds to m in the monomer unit represented by the aforescribed chemical formula

[D] and m represents any of integers from 1 to 8; R9
20 represents an H atom, a halogen atom, a CN group, a NO₂ group, COOR₉₁, SO₂R₉₂ (R₉₁ representing H, Na, K, CH₃ or C₂H₅, and R₉₂ representing OH, ONa, OK, a halogen atom, OCH₃ or OC₂H₅), a CH₃ group, a C₂H₅ group, a C₃H₇ group, a (CH₃)₂-CH group or a (CH₃)₃-C group; and,
25 in the presence of plural units, the foregoing stands

independently for each unit.)

These 3-hydroxyacyl CoAs can be synthesized by a suitable method selected from, for example, *in vitro* synthesis using an enzyme, *in vivo* synthesis using living organisms such as microorganisms and plants, and chemical synthesis. Enzymatic synthesis, especially, is commonly used to synthesize these substrates. For example, it is known a method to use a commercially available acyl CoA synthetase (acyl CoA ligase, E.C.6.2.1.3) to catalyze the following reaction:

acyl CoA synthetase

3-hydroxyalkanoic acid + CoA

→ 3-hydroxyacyl CoA

(Eur. J. Biochem., 250, 432-439 (1997), Appl. Microbiol. Biotechnol., 54, 37-43 (2000) etc.). The synthesis process using enzyme or organism may be a batch process or a continuous process using immobilized enzyme or cells.

<PHA synthetase and producing microorganisms therefor>

The PHA synthetase used in the present invention can be produced by using a microorganism selected from the microorganisms known to produce PHA synthetase, or by using a transformant to which the PHA synthetase gene of such a microorganism has been introduced.

Biosynthesis of scl-PHA is an enzymatic polymerization reaction using as a substrate at least one of (R)-3-hydroxypropionyl CoA, (R)-3-hydroxybutyryl CoA and (R)-3-hydroxyvaleryl CoA that is synthesized from various carbon sources through various metabolic pathways in a living organism. In the invention, the enzyme catalyzing the scl-PHA polymerization reaction is called scl-PHA synthetase. Among such scl-PHA synthetases, a PHA synthetase executing a biosynthesis of PHB is usually called PHB synthetase (also called PHB polymerase or PHB syntase).

The scl-PHA synthetase used in the present invention can be produced by using a microorganism selected from the microorganisms known to produce such a synthetase, or by using a transformant to which the scl-PHA synthetase gene of such a microorganism has been introduced.

For producing scl-PHA synthetase, there can be utilized microorganisms known as PHB or PHB/V producing bacteria. Such a microorganism includes not only those of *Aeromonas* sp., *Alcaligenes* sp., *Chromatium* sp., *Comamonas* sp., *Methylobacterium* sp., *Paracoccus* sp. and *Pseudomonas* sp. but also *Burkholderia cepacia* KK01, *Ralstonia eutropha* TB64 and *Alcaligenes* sp. strain TL2 separated by the present inventors. The strains KK01, TB64 and TL2

have been deposited, under the Budapest Treaty on the International Recognition of the Deposit of Microorganism for the Purpose of Patent Procedure, under the respective accession numbers: FERM BP-4235, FERM BP-6933 and FERM BP-6913 in International Patent Organism Depository of Institute of Advanced Industrial Science and Technology (former National Institute of Bioscience and Human Technology, Agency of Industrial Science and Technology).

In addition to such wild type strains, it is also possible to use a transformant for producing scl-PHA synthetase. Cloning of the scl-PHA synthetase gene, construction of expression vectors and transformants can be done according to the conventional methods. As to the cloning of scl-PHA synthetase gene, the scl-PHA synthetase gene (phbC) of *Ralstonia eutropha* was cloned. Also the present inventors have cloned phbC of *Burkholderia cepacia* KK01, and that of *Ralstonia eutropha* TB64. The transformant can be prepared by introducing a vector including such phbC into a host. The vector including phbC can be obtained by introducing phbC, for example, into a plasmid vector or a phage vector. As the host, for example, *Escherichia coli* is often utilized.

Biosynthesis of mcl-PHA and unusual-PHA is also an enzymatic polymerization reaction using as a substrate (R)-3-hydroxyacyl CoA, that is synthesized

from various alkanolic acids through various metabolic pathways in vivo (such as β oxidation pathway or fatty acid synthesis pathway). As the microorganism for producing synthetase of mcl-PHA or unusual-PHA.

5 (mcl-PHA synthetase), there can be utilized microorganisms known as mcl-PHA or unusual-PHA producing bacteria. Such microorganisms include, in addition to the above described *Pseudomonas oleovorans*, *Pseudomonas resinovorans*, *Pseudomonas* strain 61-3, *Pseudomonas putida* KT 2442, and *Pseudomonas aeruginosa*, strains of *Pseudomonas* sp. such as *Pseudomonas putida* P91, *Pseudomonas cichorii* H45, *Pseudomonas cichorii* YN2, and *Pseudomonas jessenii* P161 all of which were isolated by the present inventors, strains belonging to *Burkholderia* sp. such as *Burkholderia* sp. OK3, FERM P-17370 described in Japanese Patent Application Laid-Open No. 2001-78753 and *Burkholderia* sp. OK4, FERM P-17371 described in Japanese Patent Application Laid-Open No. 2001-69968. In addition to the above-described microorganisms, it is possible to use microorganisms of genus *Aeromonas* and *Comamonas* that can produce mcl-PHA and unusual-PHA.

Strains P91, H45, YN2 and P161 have been deposited with respective accession numbers: FERM BP-7373, FERM BP-7374, FERM BP-7375, and FERM BP-7376 in International Patent Organism Depositary of Institute

of Advanced Industrial Science and Technology (former National Institute of Bioscience and Human Technology, Agency of Industrial Science and Technology).

Microbiological properties of the above described P91, H45, YN2 and P161 are as follows. As for the strain P161, the base sequence of 16S rRNA is shown as SEQ ID NO: 5.

(Bacteriological properties of *Pseudomonas putida* P91)

10 (1) Morphology

Form and size of the cell: rod, 0.6 μm x 1.5 μm

Polymorphism of the cell: -

Mobility: +

Spore formation: -

15 Gram stain: negative

Colony shape: circular, smooth edge, low convex, smooth surface, lustrous, cream color

(2) Physiological properties

Catalase: positive

20 Oxidase: positive

O/F test: oxidizing type

Reduction of nitrate: negative

Production of indole: negative

Acidification of glucose: negative

25 Arginine dihydrolase: positive

Urease: negative

Esculin hydrolysis: negative

Gelatin hydrolysis: negative

β -galactosidase: negative

Fluorescent dye production on King's B agar: positive

(3) Substrate assimilation

5 Glucose: positive

L-arabinose: negative

D-mannose: negative

D-mannitol: negative

N-acetyl-D-glucosamine: negative

10 Maltose: negative

Potassium gluconate: positive

n-capric acid: positive

Adipic acid: negative

dl-malic acid: positive

15 Sodium citrate: positive

Phenyl acetate: positive

(Bacteriological properties of *Pseudomonas cichorii* H45)

(1) Morphology

20 Form and size of the cell: rod, 0.8 μm \times 1.0 to 1.2 μm

Polymorphism of the cell: -

Mobility: +

Spore formation: -

25 Gram stain: negative

Colony shape: circular, smooth edge, low convex,
smooth surface, lustrous, cream color

(2) Physiological properties

Catalase: positive

Oxidase: positive

O/F test: oxidizing type

5 Reduction of nitrate: negative

Production of indole: negative

Acidification of glucose: negative

Arginine dihydrolase: negative

Urease: negative

10 Esculin hydrolysis: negative

Gelatin hydrolysis: negative

 β -galactosidase: negative

Fluorescent dye production on King's B agar: positive

Growth in 4% NaCl: negative

15 Accumulation of poly- β -hydroxybutyric acid: negative

(3) Substrate assimilation

Glucose: positive

L-arabinose: negative

D-mannose: positive

20 D-mannitol: positive

N-acetyl-D-glucosamine: positive

Maltose: negative

Potassium gluconate: positive

n-capric acid: positive

25 Adipic acid: negative

dl-malic acid: positive

Sodium citrate: positive

Phenyl acetate: positive

(Bacteriological properties of *Pseudomonas cichorii* YN2)

(1) Morphology

5 Form and size of the cell: rod, 0.8 μm \times 1.5 to 2.0 μm

Polymorphism of the cell: -

Mobility: +

Spore formation: -

10 Gram stain: negative

Colony shape: circular, smooth edge, low convex, smooth surface, lustrous, translucent

(2) Physiological properties

Catalase: positive

15 Oxidase: positive

O/F test: oxidizing type

Reduction of nitrate: negative

Production of indole: positive

Acidification of glucose: negative

20 Arginine dihydrolase: negative

Gelatin hydrolysis: negative

β -galactosidase: negative

Fluorescent dye production on King's B agar: positive

Growth in 4% NaCl: positive (weakly growth)

25 Accumulation of poly- β -hydroxybutyric acid: negative

Hydrolysis of Tween 80: positive

(3) Substrate assimilation

Glucose: positive

L-arabinose: positive

D-mannose: negative

D-mannitol: negative

5 N-acetyl-D-glucosamine: negative

Maltose: negative

Potassium gluconate: positive

n-capric acid: positive

Adipic acid: negative

10 dl-malic acid: positive

Sodium citrate: positive

Phenyl acetate: positive

(Bacteriological properties of *Pseudomonas jessenii* P161)

15 (1) Morphology

Form and size of the cell: spherical: ϕ 0.6 μ m, rod:

0.6 μ m \times 1.5 to 2.0 μ m

Polymorphism of the cell: + (elongation)

Mobility: +

20 Spore formation: -

Gram stain: negative

Colony shape: circular, smooth edge, low convex,

smooth surface, pale yellow

(2) Physiological properties

25 Catalase: positive

Oxidase: positive

O/F test: oxidizing type

- Reduction of nitrate: positive
Production of indole: negative
Arginine dihydrolase: positive
Urease: negative
- 5 Esculin hydrolysis: negative
Gelatin hydrolysis: negative
 β -galactosidase: negative
Fluorescent dye production on King's B agar: positive
(3) Substrate assimilation
- 10 Glucose: positive
L-arabinose: positive
D-mannose: positive
D-mannitol: positive
N-acetyl-D-glucosamine: positive
- 15 Maltose: negative
Potassium gluconate: positive
n-capric acid: positive
Adipic acid: negative
dl-malic acid: positive
- 20 Sodium citrate: positive
Phenyl acetate: positive

The PHA producing bacteria as described above may be employed singly or in a combination of two or more kinds if necessary.

- 25 For routine culture of the PHA synthetase-producing microorganisms, for example, to prepare cell stocks, to obtain sufficient cells for enzyme

production or to maintain active state of the cells, one can select a suitable culture medium containing ingredients necessary for the growth of the microorganism. Any culture medium can be used as long
5 as it does not interfere microbial growth or vitality, including common natural media such as nutrient broth and yeast extracts, and synthetic media supplemented with nutrients.

The culture can be carried out by any culture
10 method such as liquid culture or solid culture, in which the employed microorganisms can proliferate. Also there may be employed any of batch culture, fed batch culture, semi-continuous culture or continuous culture. For example, for liquid batch culture, there
15 can be employed oxygen supply method by shaking in a shaking flask or by agitated aeration in a jar fermenter. Also there may be employed a multi-step process in which a plurality of these steps are connected.

20 To produce PHA synthetase by using the above described PHA-producing microorganism, in case of scl-PHA synthetase, there can be employed a method of growing the microorganism in an inorganic medium containing, for example, yeast extract, then
25 harvesting then the cells in the logarithmic to early stationary growth phase by centrifugation or the like and extracting the enzyme from the cells. Also in

case of mcl-PHA synthetase, there can be employed a method of growing the microorganism in an inorganic medium containing alkanoic acid such as octanoic acid and nonanoic acid, and harvesting the cells in the logarithmic to early stationary growth phase by centrifugation or the like to extract the enzyme from the cells. When cells are cultured as above, scl-PHA derived from the yeast extract etc., or mcl-PHA derived from the added alkaonoic acid is synthesized in the cells. In this case, it has been considered that PHA synthetase exists in a bound form to the fine particles of PHA synthesized in the cell. However, the inventors have found that substantial enzyme activity is present in the supernatant when the cultured cells were disrupted and centrifuged. Presumably, a certain amount of free PHA synthetase is present because this enzyme is actively synthesized during this relatively early growth phase of the logarithmic to early stationary phase.

Any inorganic culture medium can be used for the above described culture process as long as the medium contains ingredients such as phosphorus source (phosphate etc.), and nitrogen source (ammonium salt, nitrate etc.) to support microbial growth. Therefore, MSB medium, E medium (J. Biol. Chem., 218, 97-106 (1956)), or M9 medium can be used as the inorganic salt medium, for example. Composition of M9 medium

which is used in Examples is shown in the following.

[M9 culture medium]

Na₂HPO₄: 6.2 g
KH₂PO₄: 3.0 g
5 NaCl: 0.5 g
NH₄Cl: 1.0 g
(per liter, pH 7.0)

In addition, it is preferable to add, to the
aforedescribed inorganic culture medium, a stock
10 solution of trace ingredients of the following
composition to about 0.3% (v/v), for good
proliferation and production of PHA synthetase.

(Trace ingredient solution)

Nitrilotriacetic acid: 1.5 g
15 MgSO₄: 3.0 g
MnSO₄: 0.5 g
NaCl: 1.0 g
FeSO₄: 0.1 g
CaCl₂: 0.1 g
20 CoCl₂: 0.1 g
ZnSO₄: 0.1 g
CuSO₄: 0.1 g
ALK(SO₄)₂: 0.1 g
H₃BO₃: 0.1 g
25 Na₂MoO₄: 0.1 g
NiCl₂: 0.1 g

(per liter)

Culture temperature is chosen to be favorable for proliferation of the above strains, so that, for example, in the range of 14 to 40°C, preferably about 20 to 35°C.

5 It is also possible to produce PHA synthetase by using a transformant to which the PHA synthetase gene of the above PHA producing strain has been introduced. Cloning of the PHA synthetase gene, construction of expression vectors and transformants can be done
10 according to the conventional methods. To culture a transformant obtained using a bacterial host such as *Escherichia coli*, a natural medium such as LB medium or a synthetic medium such as M9 medium can be used. Cells are cultured with aeration for 8 to 27 hours at
15 25 to 37°C. After culture, cells are collected to recover PHA synthetase accumulated in the cells. Antibiotics such as kanamycin, ampicillin, tetracycline, chloramphenicol, and streptomycin may be added to the culture as required. In addition, if
20 an inducible promoter is used in the expression vector, expression may be promoted by adding a corresponding inducer to the culture medium when the transformant is cultured. Such an inducer may be isopropyl- β -D-thiogalactopyranoside (IPTG),
25 tetracycline, or indoleacrylic acid (IAA).

 The PHA synthetase may be a cell lysate, a crude enzyme such as protein components precipitated with

ammonium sulfate, or a purified enzyme purified by various methods. The enzyme preparation may be added with a stabilizer or activator such as metal salts, glycerin, dithiothreitol, EDTA, and bovine serum albumin (BSA) as required.

PHA synthetase may be isolated and purified by any method as long as the enzyme activity is maintained. For example, the enzyme can be purified as follows: a crude enzyme or ammonium sulfate precipitate thereof, prepared by disrupting microbial cells by using a French press or ultrasonic homogenizer, lysozyme, or various surfactants and by centrifuging the cell lysate, is purified by affinity chromatography, cation or anion exchange chromatography, gel filtration, or a certain combination thereof. Recombinant proteins, those expressed as a fusion protein having a tag such as histidine residue at N-terminus or C terminus, can be purified easily by binding through this tag to the affinity resin. The protein of interest may be isolated from the affinity resin binding the fusion protein by treating with protease such as thrombin and blood coagulation factor Xa, by lowering pH, or by adding high concentration imidazole as a binding competitive agent. Alternatively, when pTYB1 (made by New England Biolabs Inc.) was used as an expression vector, and the tag contains inteins, the bonding may

be broken under reducing conditions by using dithiothreitol. In addition to the histidine tag, glutathione S-transferase (GST), chitin binding domain (CBD), maltose binding protein (MBP), and
5 thioredoxin are known to allow affinity purification of fusion proteins. The GST fusion protein can be purified by a GST affinity resin.

Enzyme activity of PHA synthetase can be measured by various known methods. For example, the
10 following method measures CoA released from 3-hydroxyacyl CoA during PHA polymerization reaction catalyzed by PHA synthetase utilizing color development with 5,5'-dithiobis-(2-nitrobenzoic acid):

15 Reagent 1: a 3.0 mg/ml solution of bovine serum albumin (Sigma) dissolved in 0.1 M Tris-HCl buffer (pH 8.0), Reagent 2: a 3.0 mM solution of 3-hydroxyoctanoyl CoA in 0.1 M Tris-HCl buffer (pH
20 8.0); Reagent 3: a 10 mg/ml solution of trichloroacetic acid in 0.1 M Tris-HCl buffer (pH 8.0), Reagent 4: a 2.0 mM solution of 5,5'-dithiobis-(2-nitrobenzoic acid) in 0.1 M Tris-HCl buffer (pH 8.0). First reaction (PHA synthesizing reaction):
25 100 µl of Reagent 1 is added to and mixed with 100 µl of the sample (enzyme) solution, then the mixture is pre-incubated for one minute at 30°C, to which 100 µl of Reagent 2 is added and mixed. The resultant

mixture is incubated for 1 to 30 minutes at 30°C and the reaction is stopped by adding Reagent 3. Second reaction (color development of free CoA): the resulting first reaction solution is centrifuged
5 (15,000 × g, for 10 minutes). To 500 µl of the supernatant, 500 µl of Reagent 4 is added and incubated for 10 minutes at 30°C. Then the absorbance at 412 nm is measured. Calculation of enzyme activity: the amount of enzyme that releases 1 µmol
10 of CoA within one minute is defined as one unit (U).

Also in case of scl-PHA, a similar measurement is possible by employing s-hydroxybutyryl CoA instead of 3-hydroxyoctanoyl CoA described above.

Generally, PHA synthesized by the above
15 described enzyme is an isotactic polymer made with R bodies alone.

<Magnetic material>

In the present invention, any magnetic material capable of immobilizing the PHA synthetase can be
20 suitably selected and used. Also the kind and the structure of the magnetic material can be suitably selected according to the immobilizing method for the PHA synthetase and the form of application of the prepared construct.

25 A magnetic material constituting the construct of the invention can be, for example, a metal or a metal compound having magneticity, more specifically,

magnetite (Fe_3O_4), γ -hematite ($\gamma\text{-Fe}_2\text{O}_3$), a ferrite such as MnZn ferrite, NiZn ferrite, YFe garnet, GaFe garnet, Ba ferrite or Sr ferrite, a metal such as iron, manganese, cobalt, nickel or chromium, or an alloy such as of iron, manganese, cobalt or nickel, and such examples are not restrictive. In case of immobilizing a biological substance or administering a magnetic construct into a living organism, there can be advantageously employed, in addition to magnetite (Fe_3O_4) having satisfactory biocompatibility, a ferrite composition formed by substituting part of the metal element of magnetite with at least another metal element. Such a magnetic material has a shape that varies depending on the generation conditions, e.g., polyhedral, octahedral, hexahedral, spherical, rod-shaped or flake-shaped, but a shape with low anisotropy is preferable for stable expression of functionality. A primary particle size of the magnetic material constituting the construct of the invention can be suitably selected according to the application thereof, but it is, for example, preferably within a range of 0.001 to 10 μm .

As the magnetic material of the invention, a material showing super paramagnetism can also be employed advantageously. For example, a ferrite having a particle size as small as about 20 nm or less shows super paramagnetism under thermal

perturbation to lose retentive magnetization or coercive force. A material with a super paramagnetism can be magnetically manipulated by applying magnetic field, and is free from magnetic coagulation in the
5 absence of the magnetic field due to absence of retentive magnetization or coercive force.

The magnetic material can also be a composite material such as a matrix including a metal or a metal compound, and such a matrix can be constituted
10 of various organic or inorganic materials. Also a material formed by coating a surface of an organic polymer with a magnetic material, for example, by ferrite plating, or a material formed by dispersing a magnetic material in an organic polymer can also be
15 utilized in case the magnetic material is exposed in a part of the surface.

Furthermore, a magnetic material subjected to a hydrophobic treatment by various methods, such as a method of coating the particle surface with a fatty
20 acid or a treatment with various coupling agents, represented by a silane coupling agent, can also be employed as the magnetic material of the invention.

<Preparation of construct>

A method for producing the construct of the
25 invention includes a step of immobilizing the PHA synthetase to the magnetic material, and a step of reacting 3-hydroxyacyl CoA with thus immobilized PHA

synthetase to synthesize PHA.

For immobilizing the PHA synthetase to the magnetic material, an ordinary enzyme immobilizing method may be arbitrarily selected as long as it can ensure the activity of the enzyme and is applicable to the desired magnetic material. Examples of such a method include a covalent bonding method, an ionic adsorption method, a hydrophobic adsorption method, a physical adsorption method, an affinity adsorption method, a crosslinking method, and a lattice inclusion method, but the immobilizing method utilizing ionic adsorption or hydrophobic adsorption is particularly simple and convenient.

An enzyme protein such as of PHA synthetase is a polypeptide formed by a plurality of amino acids, and shows the property of an ionic adsorbing material owing to an amino acid having an ionic group on the side chain such as lysine, histidine, arginine, aspartic acid, or glutamic acid, and also shows the property of a hydrophobic adsorbing material owing to an amino acid having a hydrophobic side chain such as alanine, valine, leucine, isoleucine, methionine, tryptophane, phenylalanine or proline, and owing to the organic polymer structure. Therefore, it can adsorb, though in a variable level, to a solid surface having an ionic property and/or a hydrophobic property.

Among the magnetic material in the invention, a metal oxide such as ferrite has hydroxyl groups on the surface, and there can be advantageously immobilized through hydrogen bond with carboxyl groups at the surface of the PHA synthetase.

The fixation of the PHA synthetase to a core magnetic material by an ionic adsorption method or a hydrophobic adsorption method can be achieved by mixing the enzyme and the core in a predetermined reaction liquid. In this operation, it is preferable to shake or agitate a reaction vessel with a suitable intensity, in order that the enzyme is uniformly adsorbed on the surface of the core.

As the polarity and amount of the surface charge and the hydrophobicity of the core and the PHA synthetase vary depending on the pH, salt concentration and temperature of the reaction liquid, it is desirable to regulate the solution within a permissible range, according to the properties of the core to be employed. For example, in case the core principally shows an ionic adsorption property, it is possible to increase the charge amount, which contributes to the adsorption between the core and the PHA synthetase, by reducing the salt concentration. It is also possible to increase opposite charges on both components by a change in the pH. Also in case the core principally shows a

hydrophobic adsorption property, the hydrophobicity of both components can be increased by an increase in the salt concentration. It is also possible to investigate the charge state or hydrophobicity of the core and PHA synthetase by electrophoresis or wet angle measurement in advance, and to select solution conditions suitable for the adsorption. Furthermore, such conditions can be determined by a direct measurement of the adsorbed amount of PHA synthetase by the core. The adsorption amount can be measured, for example, by adding a solution of PHA synthetase of a known concentration to a core dispersion, then, after an adsorption process, measuring the concentration of the PHA synthetase in the solution and determining the amount of the adsorbed enzyme by subtraction.

In case of a core on which the enzyme is difficult to be immobilized by the ionic adsorption or the hydrophobic adsorption, a covalent bond method may be adopted if the cumbersomeness of the operation and a possibility of enzyme inactivation can be tolerated.

It is furthermore possible to fuse PHA synthetase to a peptide including an amino acid sequence having a binding ability to the magnetic material to immobilize the PHA synthetase to the surface of such a magnetic material based on a

binding property between the peptide portion of the amino acid sequence having the binding ability to the magnetic material and the magnetic material.

An amino acid sequence having a binding ability to the magnetic material can be determined, for example, by a screening of a random peptide library. For example, there can be employed a phage display peptide library prepared by fusing a product of a random synthetic gene to the N-terminus of a surface protein (for example, gene III protein) of an M13 type phage, and, in such a case, an amino acid sequence having a binding ability to the magnetic material can be determined by the following procedure.

The phage display peptide library is brought into contact with the magnetic material or a component thereof, and then bound phages and not bound phages are separated by washing. The phages binding to the magnetic material can be eluted, for example, with an acid, then neutralized with a buffer and infected on *Escherichia coli* for amplifying the phages. In this selection, after being repeated several times, plural clones having a binding ability to the object magnetic material are concentrated. Then, in order to obtain a single clone, these are infected again on *Escherichia coli* to form colonies on a culture medium plate. After each colony is cultured in a liquid medium, the phages present in the supernatant are

precipitated and purified, for example, with polyethylene glycol, and are subjected to a base sequence analysis to determine the structure of the peptide.

5 Thus obtained amino acid sequence of the peptide having a binding ability to the magnetic material is utilized by fusing it to the PHA synthetase by an ordinary genetic engineering method. The peptide having the binding ability to the magnetic material
10 can be expressed as a fusion to the N-terminus or C-terminus of PHA synthetase. It can be expressed with an inserted suitable spacer sequence. The spacer sequence preferably includes about 3 to 400 amino acids, and may include any amino acid. A most
15 preferred spacer sequence does not hinder the function of the PHA synthetase and the binding thereof to the magnetic material.

 The enzyme-immobilized magnetic material prepared through the aforescribed method may be
20 used in such a prepared state or after a lyophilization etc.

 By defining 1 unit (U) of the amount of the PHA synthetase releasing 1 μmol /minute of CoA in the PHA synthesis reaction by polymerization of 3-hydroxyacyl
25 CoA, the amount of enzyme to be used in the reaction is selected within a range of 10 to 1,000 U per 1g of magnetic material, preferably 50 to 500 U.

The construct in which the magnetic material is covered by PHA is prepared by charging the enzyme-immobilized magnetic material into an aqueous reaction liquid containing a 3-hydroxyacyl CoA, constituting the raw material of the desired PHA, and causing the PHA synthetase on the surface of the magnetic material to synthesize PHA. The aqueous reaction liquid should be constructed as a reaction system adjusted to the conditions capable of exhibiting the activity of the PHA synthetase, and is prepared with a buffer normally within a pH range of 5.5 to 9.0, preferably 7.0 to 8.5. However the condition may be set outside the aforescribed range, depending on the optimum pH or pH stability of the PHA synthetase to be employed. Such a buffer can be selected suitably according to the desired pH range, as long as the activity of the employed PHA synthetase can be exhibited, but there can be advantageously employed ordinary buffer utilized in the biochemical reactions, such as acetic acid buffer, phosphoric acid buffer, potassium phosphate buffer, 3-(N-morpholino)propane sulfonic acid (MOPS) buffer, N-tris(hydroxymethyl)methyl-3-aminopropane sulfonic acid (TAPS) buffer, trishydrochloric acid buffer, glycine buffer, 2-(cyclohexylamino)ethane sulfonic acid (CHES) buffer etc. The concentration of the buffer is not particularly limited as long as the

activity of the employed PHA synthetase can be exhibited, but is advantageously selected within a range of 5.0 mM to 1.0 M, preferably in a range of 0.1 to 0.2 M. The reaction temperature is suitably selected according the characteristics of the PHA synthetase to be employed, but is normally selected within a range of 4 to 50°C, preferably 20 to 40°C. However the condition may be set outside the
aforedescribed range, depending on the optimum temperature or the heat resistance of the PHA synthetase to be employed. The reaction time, though dependent on the stability of the PHA synthetase to be employed, is normally within a range of 1 minute to 24 hours, more desirably 30 minutes to 3 hours.

The concentration of 3-hydroxyacyl CoA in the reaction liquid is suitably selected within a range capable of exhibiting the activity of the PHA synthetase to be employed, but is normally selected within a range of 0.1 mmol/L to 1.0 mol/L, preferably 0.2 mmol/L to 0.2 mol/L. Since pH of the reaction liquid tends to become lower when the concentration of 3-hydroxyacyl CoA in the reaction liquid is high, it is preferable to have a higher concentration in the aforedescribed buffer if a high concentration is selected for 3-hydroxyacyl CoA.

Also a compound having a hydroxyl group may be suitably added to the reaction liquid in view of

controlling the molecular weight of PHA and improving the hydrophilicity of the PHA coating film.

The compound having the hydroxyl group to be used in the method of the invention is at least one selected from an alcohol, a diol, a triol, an alkylene glycol, a polyethylene glycol, a polyethylene oxide, an alkylene glycol monoester, a polyethylene glycol monoester, and a polyethylene oxide monoester, and is selected as explained more specifically in the following. The alcohol, diol or triol has a linear or ramified structure with 3 to 14 carbon atoms. The alkylene glycol or alkylene glycol monoester has a linear or ramified carbon chain with 2 to 10 carbon atoms. The polyethylene glycol, polyethylene oxide, polyethylene glycol monoester, or polyethylene oxide monoester has a number-averaged molecular weight within a range from 100 to 20,000.

Such a compound having the hydroxyl group is not particularly restricted in the concentration as long as it does not hinder the polymerization reaction of 3-hydroxyacyl CoA by the PHA synthetase, but is preferably added in an amount of 0.01 to 10% (w/v) with respect to the reaction liquid of PHA synthetase and 3-hydroxyacyl CoA, more preferably 0.02 to 5% (W/v), and may be added either collectively in an early stage of the reaction or in several portions during the reaction time.

Also in the above-described process, by varying in time the composition such as type and concentration of the 3-hydroxyacyl CoA in the aqueous reaction liquid, the monomer unit composition of the PHA constituting the particulate construct can be varied in a direction from the inner side to the outer side, in case of a particulate construct, or in a perpendicular direction, in case of a flat construct.

In such a construct showing change in the monomer unit composition, there can be realized a configuration in which the single-layered PHA covers the magnetic material with a continuous change in the composition, thus forming a gradient composition in the direction from the inner side to the outer side or in the perpendicular direction. Such a configuration can be realized, for example, in the course of synthesis of PHA, by adding 3-hydroxyacyl CoA of another composition.

In another configuration, the PHA film has stepwise changes in the composition and the magnetic material is covered by plural layers of PHA with different compositions. Such a configuration can be realized, for example, by synthesizing PHA with a certain composition of 3-hydroxyacyl CoA, then collecting the construct under preparation from the reaction liquid, for example, by centrifuging, and

adding again reaction liquid having a different composition of 3-hydroxyacyl CoA.

The construct obtained in the aforescribed reaction is subjected, if necessary, to a washing
5 step. The washing method for the construct is not particularly limited as long as it does not provide the construct with a change undesirable for the object of preparation of the construct. In case of a capsule construct with the magnetic material as a
10 core and the PHA as an outer coating, it is possible to eliminate unnecessary components contained in the reaction liquid by precipitating the construct by centrifuging and removing the supernatant. Further washing is possible by adding a washing agent, such
15 as water, a buffer or methanol in which HPA is insoluble, and executing centrifugation. Also filtration or the like may be employed in place for the centrifuging. In case the construct is formed by coating a flat-shaped magnetic material with PHA, it
20 can be washed by immersion in the aforescribed washing agent. Further, the construct may be subjected to a drying step if necessary, or to various secondary processes or chemical modification.

For example, by applying chemical modification
25 to the PHA which covers the magnetic material, there can be obtained the construct having more useful functions and characteristics.

For example, by introducing a graft chain, there can be obtained construct in which at least a part of the magnetic material is covered with PHA having various characteristics, derived from such a graft chain. Also by crosslinking the PHA, it is possible to control mechanical strength, chemical resistance, heat resistance etc., of the construct.

The method of chemical modification is not particularly limited as long as it can attain the desired functions and structure, but there can be advantageously employed a method of synthesizing PHA having a reactive functional group in the side chain and executing chemical modification utilizing the chemical reaction of such a functional group.

The type of the aforescribed reactive functional group is not particularly limited as long as it can attain the desired functions and structure, but the aforescribed epoxy group can be cited as an example. The PHA having an epoxy group in the side chain can be subjected to a chemical conversion as in the ordinary polymer having an epoxy group. More specifically, there can be carried out a conversion into a hydroxyl group or introduction of a sulfon group. It is also possible to add a compound having thiol or amine, and, more specifically, a graft chain of the polymer can be formed by reaction under addition of a compound having an end amino group

highly reactive with the epoxy group.

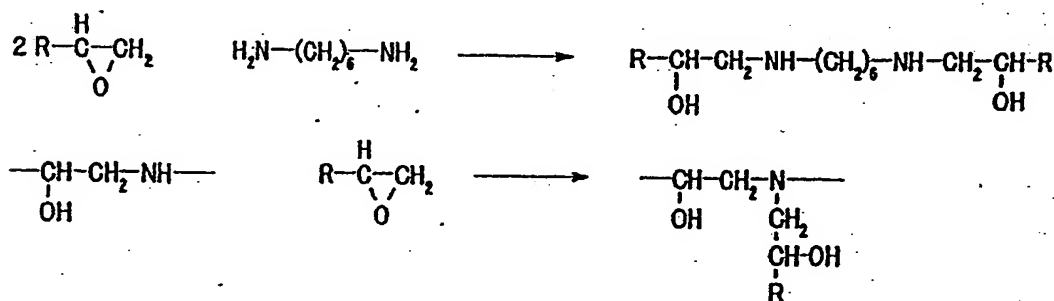
Examples of the compound having an amino group at the end include amino-modified polymers such as polyvinylamine, polyethylenimine or amino-modified polysiloxane (amino-modified silicone oil). Among these, amino-modified polysiloxane can be commercially available modified silicone oil or can be synthesized by the method described, for example, in J. Amer. Chem. Soc., 78, 2278 (1956), and is expected to provide effects by the addition of a graft chain in the polymer, such as an improvement in the heat resistance.

Also, a ligand-receptor reaction is widely utilized as a highly sensitive reaction technology. The ligand-receptor reaction includes reactions utilizing various specific binding, such as an antigen-antibody reaction, a complimentary property of nucleic acid, or a physiologically active substance and a receptor thereof such as hormone-receptor, enzyme-substrate, biotin-avidin etc. Such reaction is generally carried out by binding the ligand or the receptor with a carrier, then executing a ligand-receptor reaction, and separating the counterpart receptor or ligand from the medium. Such reaction is widely utilized in a purification method for separating and purifying an antibody, a hormone, or a nucleic acid of a specified sequence, present in

a trace amount in a medium, or a ligand-receptor assay for detecting such a substance.

The reactive functional group of PHA of the invention can be advantageously utilizing for supporting a ligand or a receptor to be used in such a ligand-receptor reaction, and useful functions and characteristics can be expressed by the grafting.

Other examples of chemical conversion of the polymer having an epoxy group include crosslinking reaction with a diamine compound such as hexamethylene diamine, succinic anhydride or 2-ethyl-4-methylimidazole, and examples of physicochemical conversion include crosslinking reaction by electron beam irradiation. Among these, the reaction between PHA having an epoxy radical in the side chain and hexamethylene diamine proceeds in the following manner to produce crosslinked polymer:



The construct of the invention includes the magnetic material by an amount of 1 to 80 wt.%, preferably 5 to 70 wt.% and further preferably 10 to 60 wt.%. An amount of the magnetic material less than

1 wt.% may result in an insufficient magnetic property, leading to a magnetic material-containing construct of an insufficient performance, and an amount of the magnetic material exceeding 80 wt.% may
5 deteriorate the function of the construct itself because of the excessive amount of the magnetic material, thus resulting in an unsatisfactory performance in practice.

A particle size of the construct of the
10 invention is suitably selected according to the application etc., but is usually 0.02 to 100 μm , preferably 0.05 to 20 μm .

Also a thickness of the coating film of the layered construct of the invention is suitably
15 selected according to the application etc., but is usually 0.02 to 100 μm , preferably 0.05 to 20 μm .

In case the construct of the invention is used for immobilizing a bio-related substance or for
administration into a living organism, there is more
20 preferred a configuration in which the magnetic material is completely covered with PHA, in order to minimize elution of the magnetic material and inhibition of the interaction of the bio-related substance.

25 In the obtained construct, the coating of the magnetic material by PHA can be confirmed by a method of combining a composition analysis, for example, by

gas chromatography and a morphological observation under an electron microscope, or a method of judging the structure by employing time of flight secondary ion mass spectrometer (TOF-SIMS) and ion sputtering thereby judging the structure from the mass spectrum of each constituent layer. However, for a more direct and simpler confirmation, there can be employed a method of combining dyeing with Nile blue A and observation under a fluorescence microscope, developed newly by the present inventors. As a result of intensive investigation for simple judgment of the PHA synthesis in a cellless (in vitro) system, the present inventors have found that Nile blue A, which generates fluorescence by specific bonding with PHA and is reported, in Appl. Environ. Microbiol., 44, 238-241 (1982), as usable for simple judgment of in vivo PHA production, can also be used for judging PHA synthesis in the cellless system by selecting suitable method and conditions of use, and have reached the above-described method. In this method, the PHA synthesis in the cellless system can be easily judged by mixing Nile blue A solution of a predetermined concentration, after filtration, with the reaction liquid containing PHA, irradiating excitation light of a predetermined wavelength and effecting observation under a fluorescence microscope of the fluorescence generated from the synthesized

PHA alone. This method, applied to the preparation of the construct of the present invention, allows to directly observe and evaluate the PHA coating the surface of the hydrophobic solution, unless the used
5 magnetic material generates fluorescence under the
aforedescribed conditions.

Also the distribution of composition of the PHA which covers the magnetic material, in the direction from the inner side to the outer side or in the
10 perpendicular direction can be evaluated by combining an ion sputtering technology and a time of flight secondary ion mass spectroscopy (TOF-SIMS).

<Utilization of construct>

One of the features of the invention is to
15 enable manufacture of a construct that has been difficult to prepare with the ordinary organosynthetic chemical methods, and it is rendered possible to obtain a construct of excellent characteristics not realizable with a capsule
20 construct or a layered construct prepared with the conventional organosynthetic chemical methods. For example, it is made possible to utilize a novel polymer compound or to provide novel function and structure, which have been difficult to realize in
25 the conventional organosynthetic methods. More specifically, utilizing extremely strict molecule identifying ability or stereoselectivity specific to

the biological catalytic action, a capsule construct
or a layered construct coated with a functional
polymer compound or a polymer compound of an
extremely high chirality, difficult to realize in the
5 conventional organosynthetic methods, can be produced
with an extremely simple process.

In particular, it is possible to prepare a
construct, coating a magnetic material of excellent
dispersion uniformity, in an extremely simple process
10 without applying an oleophilic treatment to a metal
or a metal compound having magneticity.

It is further possible to prepare, in an
extremely simple process, a capsule construct which
covers a magnetic material and which shows excellent
15 dispersibility for the magnetic material, excellent
magnetic response, and low elution of metal ions to
the exterior, thus applicable to various applications
and fields.

The construct of the invention is practically
20 free from the influence by the elution of the
magnetic material at use, since the magnetic material
is substantially absent or present only in an
extremely small amount on the surface and/or the
vicinity thereof of the particle. Therefore the
25 construct of the invention can be used in a same
manner as the conventional non-magnetic particles
even in biochemical applications in which a metal

component is often to be avoided, and can be employed, for example, for supporting antigens, antibodies, proteins, nucleic acids etc., of a wide range as a carrier for ordinary diagnostic drugs and a carrier
5 for drug delivery with a low side effect. Also it can be used as a carrier for a diagnostic drug in an enzymatic immunoassay, suppressing a non-specific color development of the enzyme resulting from elution of the magnetic material, thus being usable
10 in various detecting methods and showing an extremely high practical performance. Furthermore, the construct of the invention can also be used as a nucleic acid capturing member by supporting a specific nucleic acid or a protein probe for
15 capturing a specified nucleic acid on the surface of the particles. For such applications, the conventional magnetic material-containing polymer particle cannot be used in the PCR method since a metal component, particularly iron, hinders the PCR
20 reaction. On the other hand, the construct of the invention, in which the surfacially exposed magnetic material is practically absent, shows no hindrance to the PCR reaction and can therefore be used for the PCR method in a state supporting the captured nucleic
25 acid. Therefore, the construct of the invention can be used extremely advantageously in wide technical fields including inspection, diagnostic and

therapeutic fields utilizing nucleic acid and industrial fields utilizing nucleic acid.

However, the construct of the invention, the methods of utilizing thereof and the producing method therefor are not limited to those explained in the foregoing.

<<Methods for separation/recovery, detection and screening of target component>>

First, features of the present invention will be explained with reference to Figs. 5 to 7.

In the method for separation/recovery, detection and screening of a target component of the invention, a molecule having a binding affinity to the target component is utilized as means for selectively obtaining the target component, in a state of a construct in which the molecule 4 having a binding affinity to the target component (target component-binding molecule) is borne and immobilized in advance on the surface of a carrier. More specifically, the carrier to be employed has a structure including a coating layer of an organic polymer material on the surface of a base material of the carrier. For example, a base material constituted of a magnetic material is employed as the carrier base material 1, and at least a part of the surface of the carrier base material 1 is covered by polyhydroxyalkanoate 2 which is a polymer material of a high biological

affinity, as the coating layer of the organic polymer material, provided on the surface of the base material. In case the carrier base material contains a magnetic material, a magnetic carrier having a coating of polyhydroxyalkanoate (also represented as PHA) is also represented as "PHA magnetic construct." For immobilizing the target component-binding molecule 4 on a carrier having a coating with polyhydroxyalkanoate 2 such as the PHA magnetic construct, there is utilized a site 3, present in the coating of polyhydroxyalkanoate 2 that can selectively hold the target component-binding molecule 4. By employing a base material of a magnetic material as the carrier base material 1, a PHA magnetic construct carrying the target component-binding molecule can be prepared by the step of binding the target component-binding molecule 4 on such a PHA magnetic construct.

Then there is carried out a step of mixing the PHA coated carrier carrying the target component-binding molecule with a liquid containing a target component 5 dissolved or dispersed therein, namely, a specimen or a mixed sample, and, after a step of contacting the target component 5 contained in the specimen or the mixed sample with the target component-binding molecule 4 carried on the carrier, the target component-binding molecule 4 binds the

target component 5, whereupon the target component 5 is immobilized on the carrier. On the other hand, other components 6, 7 contained in the specimen or the mixed sample do not bind the target component binding component 4 and show little non-specific attachment to the PHA which covers the carrier surface. Even if such components 6, 7 other than the target component cause non-selective binding, they can be easily removed by a simple washing operation.

After the fixation of the target component 5 on the carrier, the carrier is recovered and separated for example by solid-liquid separating means, whereby the carrier on which the target component 5 is immobilized is recovered and separated. Also in case of a carrier utilizing a magnetic material as the carrier base material 1, through a magnetic field applied by a magneticity-generating structured member 8 (such as a permanent magnet or an electromagnet), the carrier is collected on the structured member 8 by a magnetic attractive force between the base material of the magnetic material and the structure member 8 generating the magnetic force.

In the method for separation/recovery, detection and screening of target component of the invention, as explained in the foregoing, the target component-binding molecule is held by polyhydroxyalkanoate which is a polymer material of a high biological

affinity, and the binding of the target component and the target component-binding molecule also takes place on the surface portion coated with polyhydroxyalkanoate, so that the separation/recovery, detection and screening of target component can be carried out in a condition close to that of a living organism.

The method for separation/recovery, detection and screening of target component of the invention may be applied to a specimen where components other than the target component are not present. In case the target component is present alone in the specimen but with a low concentration in a large volume, such a target component is immobilized to the carrier and is recovered with the carrier whereby the target component is present with a higher proportion in thus separated and recovered carrier and is thus concentrated, thereby facilitating the detection.

The polyhydroxyalkanoate 1 is already explained in detail in <PHA>, and the magnetic material 2 in <Magnetic material>, the molecule 4 having a binding affinity to the target component and the target component 5 will be explained in a section <target component and molecule having binding affinity to target component>; and magnetic member 8 in a section <magnetic separation and washing of target component binding magnetic construct>.

Also a compound having a hydroxyl group may be suitably added to the reaction liquid in view of controlling the molecular weight of PHA and improving the hydrophilicity of the PHA coating film.

5 The compound having the hydroxyl group to be added to the reaction liquid in the method of the invention is at least one selected from an alcohol, a diol, a triol, an alkylene glycol, a polyethylene glycol, a polyethylene oxide, an alkylene glycol
10 monoester, a polyethylene glycol monoester, and a polyethylene oxide monoster, and is selected as explained more specifically in the following. The preferred alcohol, diol or triol has a linear or ramified structure with 3 to 14 carbon atoms. The
15 preferred alkylene glycol or alkylene glycol monoester has a linear or ramified carbon chain with 2 to 10 carbon atoms. The preferred polyethylene glycol, polyethylene oxide, polyethylene glycol monoester, or polyethylene oxide monoster has a
20 number-averaged molecular weight within a range from 100 to 20,000.

Such a compound having the hydroxyl group is not particularly restricted in the concentration as long as it does not hinder the polymerization reaction of
25 3-hydroxyacyl CoA by the PHA synthetase, but is preferably added in an amount of 0.01 to 10% (w/v) with respect to the reaction liquid containing PHA

synthetase and 3-hydroxyacyl CoA, more preferably 0.02 to 5% (W/v), and may be added either collectively in an early stage of the reaction or in several portions during the reaction time.

5 The construct including a magnetic material as a core, to be used in the method of the invention, includes the magnetic material by an amount of 1 to 80 wt.%, preferably 5 to 70 wt.% and further preferably 10 to 60 wt.%. An amount of the magnetic
10 material less than 1 wt.% in the construct may result in an insufficient magnetic property, leading to a magnetic material-containing construct of an insufficient performance, and an amount of the magnetic material exceeding 80 wt.% may deteriorate
15 the function of the construct itself achieved by a sufficient PHA coating layer on the surface, because of a relative decrease of the content of the PHA coating the surface of the magnetic material constituting the core, thus resulting in an
20 unsatisfactory performance in practice.

 A particle size of the particulate construct to be used in the method of the invention is suitably selected according to the individual application etc., but is usually selected within a range of 0.02 to 100
25 μm, preferably 0.05 to 20 μm.

 <Target component, and molecule having a binding affinity to target component>

A "target component" being an object of the method of the invention is physiologically active, and is often present in a specimen, as a mixture with other substance, or, singly without other substances at a low concentration in a large volume. It is therefore desired to obtain a method of separating/recovering, detecting or screening only the "target component" in the specimen.

The method of the invention utilizes, for the
aforedescribed objective, a "molecule having a binding affinity to the target component (hereinafter also represented as "target component-binding molecule")" advantageously utilizable for capturing the "target component" only.

Specific examples of the "target component" and the "molecule having binding affinity to the target component," considered in the method of the present invention, include a nucleic acid, a protein, a peptide, a sugar chain, a lipid, a low-molecular compound, a composite thereof, and a substance containing such a substance as a portion.

"Nucleic acid" includes a deoxyribonucleic acid, a ribonucleic acid, an oligonucleotide, a polynucleotide, an aptamer, and a ribozyme.

"Protein" includes natural and artificial irregular molecules such as a glycoprotein, a lipoprotein, a membrane protein, a labeling protein,

or a low-molecular peptide.

In case such a protein is immunoreactive, there are included, for example, an antibody, an antigen, a haptene, and a complex thereof, and, particularly in case it is an antibody, there are included a monoclonal antibody, a polyclonal antibody, a recombinant protein antibody, a natural antibody, a chimeric antibody, a hybrid antibody mixture (single or plural), a single chain antibody expressing phage antibody (including the entire phages expressing single-chain antibodies), and an antibody-protein fusion, and a hybrid mixture thereof. In case the protein is a catalytic reactive member, there are included a natural enzyme, a modified enzyme prepared by genic engineering, a semi-artificial enzyme formed by complexing with a synthetic molecule such as polyethylene glycol, and a semi-artificial enzyme in which a non-natural amino acid is introduced.

The "antibody" is usually exemplified by IgG (immunoglobulin G), but there can also be employed substances of a lower molecular weight such as F(ab')₂, Fab', Fab or Fv obtained by treating with a proteolytic enzyme such as pepsin and papain, or a reducing agent such as dithiothreitol. In addition to IgG, there can also be employed IgM or a fragment of a lower molecular weight of IgM obtained by a process similar to that for IgG. Furthermore, a monoclonal

antibody or a polyclonal antibody can be used as the "target component-binding molecule" in the invention. In case of employing the monoclonal antibody as the "target component-binding molecule" of the invention, 5 for a protein having a repetitive structure such as a surface antigen of hepatitis B virus or for an antigen containing plural epitopes within the molecule as in a CA19-9 antigen, there may be employed plural monoclonal antibodies, reactive to 10 the respective epitopes, in combination. It is also possible to utilize two or more different identifying epitopes in a combination.

On the other hand, "antigen" includes various substances such as a protein, a polypeptide, a 15 steroid, a polysaccharide, a lipid, a pollen, a recombinant protein produced by a genetic engineering method, a drug etc. Thus the "antigen" considered in the invention includes, among all the substances capable of inducing an antibody production in human 20 or in an animal; single or plural substances selected under a particular object such as diagnosis, and a mixture containing the same.

"Peptide" in the invention indicates a fragment of a protein regardless of the molecular weight 25 thereof.

"Low-molecular compound" is a molecule, preferably an organic molecule, of a low molecular

weight, recognizable by a receptor. The low-molecular compound is usually a compound capable of a specific binding with a protein, is often a physiologically active substance or a drug candidate, and, in particular, a low-molecular compound of an antigen property may also be called "haptens."

"Sugar chain" includes an oligomer or a polymer of linear or branched structure, formed by a chain of a plurality (several to several tens) of a monosaccharide unit selected from glucose, mannose, N-acetylglucosamine, fucose, galactose, glucuronic acid, N-acetylglucosamine, and sialic acid. Also in the method of the invention, there is included a glycoprotein which is a composite of such a sugar chain and a protein, a glycolipid which is a composite with a lipid, or, in case such a sugar chain is expressed at the surface of a living cell, a cell itself expressing the sugar chain at the surface of the cell membrane or a fragment of the cell membrane.

"Lipid" includes a composite lipid, a natural lipid (acylglycerol), a lipoprotein which is a composite with a protein, a phospholipid such as lecithin, locating in a tissue boundary lipid membrane such as a cell organelle membrane such as a neural tissue, a plasma membrane, mitochondria, a microsome, or a cell nucleus, and a liposome as a double-membrane lipid

capsule.

The core material to be employed in the invention may have a particulate shape, a flat shape or a film shape, but preferably is in a particulate shape in consideration of a specific separating method by a magnetic operation, and is more preferably constituted of fine particles of a particle size of 0.001 to 10 μm in consideration of dispersion into liquid.

In case of employing a method of immobilizing the PHA synthetase principally by the ion adsorption method, there can be employed a core having an ionic functional group on the surface, for example, a clay mineral such as caolinite, bentonite, talc or mica, a metal oxide such as alumina or titanium dioxide, or an insoluble inorganic salt such as silica gel, hydroxy apatite, or calcium phosphate gel. Also a polymer having an ionic functional group, such as an inorganic pigment, an ion exchange resin, a chitosan, or a polyaminopolystyrene including these materials as a principal component, can also be utilized as an ion adsorbing core.

In case the base material is constituted of a magnetic material, for example, a metal oxide such as ferrite, a hydroxyl group is present on the surface thereof and can be advantageously utilized for fixation by a hydrogen bonding with the carboxy group

on the surface of the PHA synthetase.

On the other hand, in case of immobilizing the PHA synthetase principally by hydrophobic adsorption, there can be utilized a core with a non-polar surface, for example various polymers lacking an ionic functional group on the surface or showing a hydrophobic group on the surface, such as a styrenic polymer, an acrylic polymer, a methacrylic polymer, a vinyl ester or a vinylic polymer. More specifically, an organic pigment such as an azo pigment having plural aromatic rings, a phthalocyanine pigment or an anthraquinone pigment of a condensed polycyclic structure, or carbon black has a hydrophobic adsorbing property. The hydrophobic adsorption method is also applicable to a magnetic material subjected to an oleophilic treatment.

The fixation of the PHA synthetase to a core by an ionic adsorption method or a hydrophobic adsorption method can be achieved by mixing the PHA synthetase and the core in a predetermined reaction liquid. In this operation, it is preferable to shake or agitate a reaction vessel with a suitable intensity, in order that the PHA synthetase is uniformly adsorbed on the surface of the core.

As the polarity and amount of the surface charge and the hydrophobicity on the core and the PHA synthetase vary depending on a pH, a salt

concentration and a temperature of the reaction liquid, it is desirable to regulate the solution within a permissible range, according to the nature of the core to be employed. For example, in case the core principally shows an ionic adsorption property, it is possible to increase the charge amount, contributing to the adsorption between the core and the PHA synthetase, by reducing the salt concentration. It is also possible to increase opposite charges on both components by a change in the pH. Also in case the core principally shows a hydrophobic adsorption property, the hydrophobicity of both components can be increased by an increase in the salt concentration. It is also possible to investigate the charge state or the hydrophobicity of the core and the PHA synthetase by an electrophoretic measurement or a wetting angle measurement in advance, and to select a solution condition suitable for the adsorption. Furthermore, such a condition can be determined by a direct measurement of the adsorbed amount of the core and the PHA synthetase. The adsorption amount can be measured, for example, by a method of adding a solution of the PHA synthetase of a known concentration to a core dispersion, then, after an adsorption process, measuring the concentration of the PHA synthetase in the solution and determining

the amount of the adsorbed enzyme by a subtraction.

In case of a core on which the enzyme is difficult to fix by the ionic adsorption or the hydrophobic adsorption, a covalent bonding method may be adopted if the cumbersomeness of the operation and a possibility of enzyme deactivation can be tolerated. There can be employed, for example, a method of executing a diazo formation on a core material (solid particle) having an aromatic amino group, and executing a diazo coupling of the enzyme thereto, a method of forming a peptide bond between the core material (solid particle) having a carboxy group or an amino group and the PHA synthetase, a method of alkylation between the core material (solid particle) having a halogen group (halogenoalkyl group) and an amino group of the PHA synthetase, a method of reacting a polysaccharide core particle activated with cyan bromide with an amino group of the PHA synthetase, a method of crosslinking an amino group of the core material (solid particle) and an amino group of the enzyme, a method of reacting the core material (solid particle) having a carboxy group or an amino group with the PHA synthetase in the presence of a compound having an aldehyde group or a ketone group and an isocyanide compound, or a method of executing an exchange reaction between the core material (solid particle) having a disulfide group

(-S-S-) and a sulfanyl group (-SH) of the PHA synthetase.

It is also possible to adsorb the PHA synthetase on the surface of the core material (solid particle) by an affinity adsorption. The affinity adsorption is a biological adsorption between a biological polymer and a specified substance called a ligand and showing a specific affinity thereto, and can take place, for example, between an enzyme and a substrate, an antibody and an antigen, a receptor and an information material such as acetylcholine, or mRNA and tRNA. In case of immobilizing an enzyme protein by affinity adsorption, there is generally employed a method of binding a substrate or a reaction product of the enzyme, a competitive inhibitor, a coenzyme or an allosteric effector as a ligand to a solid surface and realizing an affinity adsorption on such a solid surface through a binding between such a ligand and an added enzyme protein. However, in the PHA synthetase, in case the 3-hydroxyl CoA constituting the substrate therefor is employed as the ligand, an active position of the PHA synthetase, catalyzing the PHA synthesis, is blocked by the binding with the ligand, thus becoming unable to synthesize PHA. Nevertheless, the PHA synthesizing activity of the PHA synthetase can be maintained even after the fixation by affinity adsorption, by fusing

another biological polymer to the PHA synthetase in advance and executing affinity adsorption utilizing a ligand for such a biological polymer. The fusion of the PHA synthetase and the biological polymer may be
5 achieved by a genetic engineering method, or by chemically binding the biological polymer to the PHA synthetase. There may be utilized any biological polymer as long as a ligand thereto is easily available and can be easily coupled to the core, but,
10 in case a fused substance is developed by a gene recombination, the biological polymer to be fused is preferably a protein. More specifically, *Escherichia coli* in which a fusion gene, of GST gene and a gene sequence of the PHA synthetase is introduced by
15 transformation is utilized for producing a fused protein of GST and the PHA synthetase, and such a fused protein is added to sepharose coupled with glutathione serving as a ligand for GST thereby enabling affinity adsorption of the PHA synthetase of
20 fused protein type on the sepharose.

As already described, it is furthermore possible to fuse a peptide, including an amino acid sequence having a binding ability to the magnetic material, with the PHA synthetase, and to fix the PHA
25 synthetase to the surface of such a magnetic material based on a binding property between the peptide portion of the amino acid sequence having the binding

ability to the magnetic material and the magnetic material.

<Immobilization of target component-binding molecule on PHA construct>

5 For immobilizing a molecule having a binding affinity to the target component (hereinafter represented as a target component-binding molecule) on the surface of the PHA magnetic construct, there can be utilized a physical adsorption by a physical
10 affinity such as hydrophobicity, ionic property or van der Waals force between the PHA coating the surface and the target component-binding molecule, but, in consideration of reproducibility and stability, it is more desirable to form an
15 irreversible covalent bond by combining a functional group in the side chain of the PHA and a functional group present in the target component-binding molecule either directly or in the presence of a
 converting/modifying/activating reagent.

20 As a configuration of the PHA construct to be employed in the method of the invention, there can be employed a PHA construct having an epoxy group on the side chain of the PHA coating the surface. Such an epoxy group can form a covalent bond directly with an
25 amino group ($-NH_2$) or a sulfanyl group ($-SH$) provided in the target component-binding molecule. As the covalent bond can be formed without requiring an

agent, this method is useful for immobilizing an easily denatured protein such as an enzyme protein, or an antibody (Fc) receptor protein such as A protein or G protein. It is also possible to achieve an efficient recovery of a target protein with histag, by immobilizing iminodiacetic acid (IDA) on the surface coating PHA and adding a metal ion such as Ni^{2+} . This method of forming a covalent bond utilizing the reaction with the epoxy group is naturally applicable also in case the target component-binding molecule is a low molecular chemical substance such as a drug candidate, as long as such a low molecular chemical substance has an amino group ($-\text{NH}_2$) or a sulfanyl group ($-\text{SH}$).

Such an epoxy-containing PHA construct can be converted into a PHA construct having an amino group, by a reaction under an alkaline condition with ammonium hydroxide or hexamethylene diamine hydrochlorate salt of 10 to 100 molar amount with respect to the epoxy group. In case the target component-binding molecule is a protein or a peptide, such an amino group can form, by means of a crosslinking agent such as NHS (N-hydroxysuccinimide), with a terminal carboxy group in the main chain thereof or a carboxy group of a side chain of a residue of an amino group such as aspartic acid or glutamic acid present in such a protein or peptide.

In such a case, it is necessary to convert the carboxy group of the target component-binding molecule into an activated ester by an NHS treatment, and it is necessary to confirm, in advance, whether the function of the target component-binding molecule is sufficiently sustained after the NHS treatment. This immobilization method utilizing the amide bond formation is naturally applicable also in case the target component-binding molecule is a low molecular chemical substance such as a drug candidate, as long as such a low molecular chemical substance has a carboxy group.

Furthermore, in case the target component-binding molecule is a glycoprotein such as a sugar chain or lectin, a glycolipid such as lipopolysaccharide, such an amino group can form a stable bond with an aldehyde structure (formyl group: -CHO) in the sugar chain by a Schiff's base (-CH=N-) formation and a reductive amination. This covalent bond forming method utilizing a reaction between the sugar chain and the amino group is accelerated by a partial oxidation of the sugar chain portion, introducing an aldehyde structure in the sugar chain, and also is applicable for immobilizing an antibody molecule such as IgG, having a sugar chain in the Fc portion thereof. This method utilizing the amino group and the aldehyde structure (formyl group: -CHO)

is naturally applicable also in case the target component-binding molecule is a low molecular chemical substance such as a drug candidate, as long as such a low molecular chemical substance has an aldehyde structure (formyl group: -CHO) or can introduce an aldehyde structure (formyl group: -CHO) by a partial oxidation.

Furthermore, such an amino group can be coupled with a target component-binding molecule having a sulfanyl group (-SH) in the presence of a maleimide derivative, a pyridylthio compound or an iodine/bromine acetyl compound. Basic reacting conditions of the amino group and the sulfanyl group (-SH) are, in case of employing a maleimide derivative, 2 to 4 hours at 4°C to room temperature in 0.1 M sodium phosphate (pH 6.5 to 7.5), also in case of employing a pyridylthio compound, 15 to 20 hours at the room temperature in a PBS buffer (pH 7.5), and, in case of employing an iodine/bromine acetyl compound, 1 hour at the room temperature and in the absence of light, in a 0.5 M sodium borate solution (pH 8.3), but such reacting conditions may be suitably changed according to the type of the target component-binding molecule and an application thereafter.

Another configuration of immobilizing on the PHA construct to be employed in the method of the

invention utilizes a PHA construct having a carboxy group on the side chain of the PHA. In case the target component-binding molecule is a protein or a peptide, such a carboxy group can form an amide bond, by means of a crosslinking agent such as NHS (N-hydroxysuccinimide), with a terminal amino group of the main chain thereof or an amino group on a side chain of a residue of an amino group such as lysine or arginine present in the protein or peptide. This amide forming reaction can be improved in rate and frequency by converting the carboxy group of PHA into an activated ester in advance. Also in this amide forming method utilizing the carboxy group of the PHA side chain, in case the target component-binding molecule is a DNA or an oligonucleotide, it can be carried on PHA by the aforescribed reaction by employing DNA or oligonucleotide of which terminal is converted into an amino group by an already known method. This amide forming method utilizing the carboxy group of the PHA side chain is naturally applicable also in case the target component-binding molecule is a low molecular chemical substance such as a drug candidate, as long as such a low molecular chemical substance has an amino group.

Another configuration of immobilizing on the PHA construct to be employed in the method of the invention utilizes a PHA construct having a halogen

such as a chloro group (-Cl), a bromo group (-Br) or a fluoro group (-F) on the side chain of PHA. Such a halogen, in case it is a chloro group or a bromo group, can form a sulfide bond (-S-) with the target component-binding molecule having a sulfanyl group (-SH) under a mild condition.

It is furthermore possible to bind a substance showing a specific binding to the "target component-binding molecule" (for example such a substance being protein A or protein G in case the "target component-binding molecule" is an antibody) to be immobilized on the construct surfacially coated with PHA, to such a surface utilizing the aforescribed active functional group, and then to cause a specific binding of the desired "target component-binding molecule" to such a substance showing specific binding with the "target component-binding molecule" thereby achieving immobilization thereof on the construct. Also it is further possible to immobilize the "target component-binding molecule" after a further modification. As an example of the latter, biotin is immobilized on a PHA construct of carboxy type utilizing a reagent NHS-iminobiotin (manufactured by Pierce Inc.), and a target component-binding molecule modified with avidin or streptoavidin is carried on the construct by a specific binding with biotin. Examples of usable

finity binding pair include lectin and sugar, haptene and antibody, protein A or protein G and antibody Fc, other protein pairs showing specific binding, phenylboronic acid and salicylhydroxamic acid, and other pairs of chemical portions which mutually react but do not react with a protein.

In order to prevent a non-specific adsorption on the construct, it is preferred to coat a non-carrying portion of the surface of the construct with a "blocking agent" which does not deteriorate the activity of the carried target component-binding molecule. Examples of the block agent suitable for such "blocking treatment" include collagen, gelatin (particularly cold-water fish hide gelatin), skimmed milk, a serum protein such as BSA and various compounds including a hydrophobic portion and a hydrophilic portion which do not react with protein.

<Contact and binding between PHA construct carrying target component-binding molecule and target component>

In the method of the invention, contact between a PHA construct carrying a target component-binding molecule and a target component is usually carried out in an aqueous medium, but, in case the target component has a low solubility in water as in certain drug candidates, the contact may be carried out in an emulsion system by adding a polar solvent such as an

alcohol, acetone, DMSO (dimethyl sulfoxide) or DMF (dimethylformamide) with a surfactant such as Tween, Triton or SDS, and eventually a non-polar solvent such as toluene, xylene or hexane, thereby accelerating the binding reaction. However, in case such a solvent or surfactant is employed, a concentration thereof has to be selected within such a range that does not deteriorate the affinity binding function of the carried target component-binding molecule.

In the method of the invention, in order to stimulate the contact and binding between the PHA construct carrying the target component-binding molecule and the target component, heating means or agitating means may be employed within an extent not deteriorating the affinity binding function of the target component-binding molecule, and ultrasonic means may also be employed.

When the construct to be employed in the method of the invention is a magnetic construct, a manipulation utilizing a magnetic force is also possible for stimulating contact and binding between the target component-binding molecule carried on the surface of the construct and the target component. In a manipulation by the magnetic force, it is possible to repeat application and release of the magnetic force, utilizing a structured member for generating a

magnetic force (such as a permanent magnet or an
electromagnet, which may hereinafter be collectively
called a magnet). In case of employing an
electromagnet, a current supply and a current cut-off
5 are repeated by a switching operation, thereby
repeating capture and release of the magnetic
construct. As an example of the operation utilizing a
magnet, a probe-shaped electromagnet is inserted into
a reaction vessel, and a current supply and a current
10 cut-off to the magnetic are repeated by a switching
operation or an on-off operation of a power supply,
thereby repeating capture and release of the magnetic
construct. In another example, a magnet is positioned
outside a reaction vessel, and an applied intensity
15 of the magnetic field is repeatedly varied by a
switching operation, an on-off operation of a power
supply, or a regulation of the distance of the magnet
to the reaction vessel, thereby repeating capture and
release of the magnetic construct.

20 In the method of the invention, "binding"
between the target component and the target
component-binding molecule means a specific binding
of a molecule to the other by a chemical or physical
action between the pair. Examples of such binding
25 include not only a binding between an antigen and an
antibody by a known antigen-antibody reaction, but
also binding between biotin and avidin, between a

hydrocarbon and lectin, between complementary sequences of nucleic acid and nucleotide, between molecules of an actuator and a receptor, between a co-enzyme and an enzyme, between an enzyme inhibitor and an enzyme, between a peptide sequence and an antibody specific to such a sequence or to all the proteins, between an acid and a base in polymer, between a dye and a protein binder, between a peptide and a specific protein binder (ribonuclease, S-peptide and libonuclease S-protein), between a sugar and boric acid, and between similar molecule pair having an affinity enabling a molecular association in a binding assay, but such an examples are not restrictive. Also the binding pair may be elements similar to original binding elements, such as analogs of the substance to be analyzed or binding elements, produced by recombination or molecular engineering. In case the binding elements are immuno-reactive, there can be employed an antibody, an antigen, a haptene or a complex thereof, and, in case of employing an "antibody," there can be utilized a monoclonal antibody, a polyclonal antibody, a recombinant antibody, a natural antibody, a chimera antibody, a mixture(s), a single-chain antibody-displaying phage (including the entire phage), a fragment(s) thereof expressing the single-chain antibody, or a mixture of binding elements of

antibody and protein.

Along with the recent progress in the evolution molecular engineering, there has been developed a technology of screening a nucleic acid molecule, or
5 an aptamer (also called nucleic acid antibody), having a high affinity to a target molecule such as protein from a random oligonucleotide library ("systematic evolution of ligands by exponential enrichment"; SELEX or in vitro selection). This
10 screening method based on aptamer has been applied to preparation of high affinity ligands easier and faster than antibodies (for example, Nature, 355:564(1992), International Patent Application No. WO 92/14843, Japanese Patent Application Laid-Open
15 Nos. H08-252100 and H09-216895).

Also binding between a transcription factor being a protein and a nucleic acid having a specified base sequence is expected in clarifying causes of diseases and in applications for effective diagnosis
20 and therapy.

The "binding" considered in the method of the invention naturally includes such nucleic acid-protein affinity binding.

Also the "binding" considered in the method of
25 the invention includes any and all physical or chemical adhesion, and specific/selective association, regardless whether it is permanent or temporary. In

general, it is possible to cause a physical adhesion between a ligand molecule and a receptor by an ionic interaction, a hydrogen bonding, a hydrophobic force or a van der Waals force. The interaction of

5 "binding" may be short in time, as in the case where a chemical change is induced by the binding. This generally applies to a case where the binding component is an enzyme and the "target component-binding molecule" is a substrate for the enzyme. Also
10 the chemical connection may be irreversible or reversible. The binding may also become specific under a particularly different condition.

In the method of the invention, a practical example of the contacting and binding process between
15 the target component-binding molecule carried on the PHA magnetic construct and the target component is a process of contacting a PHA magnetic construct carrying a target component-binding molecule with a biological specimen containing a natural protein such
20 as a tissue, a cell homogenate or a fluid such as serum thereby causing specific adsorption and binding between such a natural protein with the target component-binding molecule carried on the surface of the PHA magnetic construct.

25 Another example of such a process employs a construct carrying a protein constituting the target component coupling molecule, thereby causing a

selective coupling with an antibody portion displayed on the surface of a suitable bacteriophage, as in the known phage display antibody selecting method.

Also as another example, there can be conceived
5 a process of contacting with a biological specimen containing a receptor, for example, a liquid such as a hybridoma supernatant or a phage display fluid thereby causing the receptor contained in such a biological specimen to be specifically adsorbed on
10 the target component-binding molecule carried on the PHA magnetic construct.

<Magnetic separation and washing of magnetic composite binding target component>

In a method of binding the target component in a
15 magnetic composite (magnetic construct in which the target component-binding molecule is carried) and separating and washing the magnetic composite binding the target component, a magnetic separating operation is carried out by repeating exertion and release of
20 magnetic force with a structured member generating a magnetic force (such as a permanent magnet or an electromagnet). In case of employing an electromagnet, power supply and cut-off to the electromagnet are carried out by a switch operation to carry out
25 capture and release of the magnetic composite binding the target component.

In an embodiment, a probe-shaped magnet is used

to capture the magnetic composite binding the target component in a container, and remaining liquid is removed from the container. Then a washing liquid is charged into the container to wash the magnetic composite binding the target component, still captured on the probe. Otherwise, the washing operation can be carried out by taking the probe-shaped magnet, on which the magnetic composite binding the target component is captured, from a reaction liquid and moving it to a washing liquid.

In another method, a magnet is positioned outside a container to attract the magnetic composite binding the target component to an internal wall of the container at a replacement of the liquid. By removing such an external magnet, the magnetic composite binding the target component is released and is mixed with the liquid (washing liquid) whereby a washing operation can be carried out.

For such a magnetic separating operation, there can be utilized various magnetic selecting equipment commercially available for the purpose of manipulating magnetic particles. Examples of a magnetic selecting equipment include DYNALMCP manufactured by DYNAL Inc., MAIA Magnetic Separator manufactured by Serono Diagnostics Inc., Magnetight Separation Stand manufactured by Takara Shuzo Co., and BioMag Separator manufactured by Advanced.

Magnetics Inc.

<Elution and liberation of target component from
PHA construct>

In the method of the invention, it is also
5 possible, after isolation of the magnetic composite
binding the target component, to elute/liberate the
binding component from the target component-binding
molecule if necessary. In case the target component-
binding molecule and the target component are a
10 protein-protein combination, they can be liberated
under an ordinary liberating condition (pH 2, 4M
guanidine, 2M ammonium thiocyanate, 1% SDS etc.), but
the liberated target component protein is often
denatured. The liberated target component protein,
15 even if denatured, is acceptable in case it is
purified by electrophoresis or HPLC, but a restoring
process such as a dialysis may be required in case an
original steric structure etc., of the target
component protein is to be confirmed.

20 <Detection of target component>

In the method of the invention, the detection of
the target component may be carried out by any method
employable in an immunoassay or a hybridization assay
such as an ordinary colorimetry, a fluorescent method,
25 a chemiluminescence method or a radioisotope method.
Also the target component eluted/liberated by the
aforescribed method from the target component-

binding molecule can be analyzed by similar methods.

It is also possible, in case the target component is DNA, to verify the base sequence by a sequencer after an amplification by PCR or the like, and, in case the

5 target component is a protein, to carry out an enzymatic decomposition, then to separate the enzyme digested fragments by two-dimensional electrophoresis or HPLC and to carry out an electrospray/ionization and a mass spectroscopic analysis, or, to analyze the
10 target component protein either directly or after enzymatic decomposition, by a matrix assisted laser deionization-time of flight mass spectroscopy (MALDIToF-MS).

It is naturally possible also detect the target
15 component by nuclear magnetic resonance (NMR) spectroscopy, infrared absorption (IR) spectroscopy or ultraviolet absorption (UV) spectroscopy either singly or in a combination thereof.

(Examples)

20 In the following, the present invention will be explained in more details by examples thereof. These examples represent examples of the optimum embodiments of the present invention, but the present invention is by no means limited by these examples.
25 Following examples employ polyhydroxyalkanoate including a 3-hydroxyalkanoic acid unit, but any polyhydroxyalkanoate can be utilized. In the

following, "%" is based on mass unless otherwise specified, and "part" means part by mass.

At first there will be shown a method for preparing a magnetic material as a common base material.

(Reference Example 1)

<Preparation of magnetic material>

In an aqueous solution of ferrous sulfate, a solution of sodium hydroxide of 1.0 to 1.1 equivalents to iron ions was added to prepare an aqueous solution containing ferrous hydroxide. Then air was blown in while the solution was maintained at about pH 8 to carry out an oxidation reaction at 80 to 90°C to prepare a slurry for generating seed crystals.

Then, to this slurry, an aqueous solution of ferrous sulfate was added in an amount of 0.9 to 1.2 equivalents with respect to the initial alkali amount (sodium component of sodium hydroxide), and the oxidation reaction was carried out by blowing air while the slurry was maintained at about pH 8. Magnetic iron oxide particles generated after the oxidation reaction were washed, filtered and dried, and the coagulated particles were broken to obtain a magnetic material (1) of an average particle size of 0.10 μm .

In the following, there is shown an embodiment

on scl-PHA synthetase (reference examples 2 to 4, examples 1 to 10).

(Reference Example 2)

<Preparation of transformant having scl-PHA synthetase producing ability>

Strain TB64 was cultured in 100 ml of LB medium (1% polypeptone, 0.5% yeast extract, 0.5% sodium chloride, pH 7.4) overnight at 30°C, then the chromosomal DNA was isolated by the method of Marmar et al. The obtained chromosomal DNA was partially digested by a restriction enzyme Sau3AI. A vector pUC18 was also cut by a restriction enzyme BamHI. After terminal dephosphorylation (Molecular Cloning, 1, 572, (1989); Cold Spring Harbor Laboratory Press), Sau3AI partial digestion fragments of the chromosomal DNA were ligated to the cleavage site of the vector using a DNA ligation kit Ver. II (TAKARA SHUZO CO., LTD.). With these ligated chromosomal DNA fragments, *Escherichia coli* HB 101 was transformed to construct a chromosomal DNA library of strain TB64.

Next, to obtain DNA fragments covering the PHB synthetase gene of strain TB64, an expression screening was carried out. LB culture medium containing 2% glucose was used for screening, and Sudan black solution was sprayed when the colony grown on the agar medium plate to a suitable size, and a colony showing fluorescence under UV light.

irradiation was collected. The DNA fragment covering the PHB synthetase gene was obtained by recovering the plasmid from the acquired colony with the alkali method.

5 The acquired gene fragment was transformed into a vector pBBR122 (Mo Bi Tec) having a wide-host-replication range (Mo Bi Tec) not belonging to any of IncP, IncQ, or IncW incompatibility group. When *Ralstogna eutropha* TB64m1 strain (PHB synthesis
10 negative strain) was transformed with this recombinant plasmid by electroporation, the PHB synthesizing capacity of the TB64m1 strain was recovered to show complementarity.

15 The base sequence was determined by Sanger method on the fragment covering the PHB synthetase gene. As a result it was confirmed that the PHB synthetase gene indicated by SEQ ID NO.: 1 was present in the fragment.

20 Then an oligonucleotide having the base sequence in the vicinity of the starting codon of the scl-PHA synthetase gene was designed and synthesized (Amarsham Farmacia Biotech), and PCR was carried out by using the oligonucleotide as a primer to amplify the fragment including the scl-PHA synthetase gene
25 (LA-PCR kit; TAKARA SHUZO).

 The obtained PCR amplified fragment was completely digested by a restriction enzyme BamHI. A

vector pTrc99A was also cut by BamHI. After terminal dephosphorylation (Molecular Cloning, 1, 572, (1989); Cold Spring Harbor Laboratory Press), complete BamHI digestion fragments were ligated using a DNA ligation kit Ver. II (TAKARA SHUZO CO., LTD.).

With the obtained recombinant plasmid vectors, *Escherichia coli* HB 101 was transformed by the calcium chloride method (TAKARA SHUZO) to recover a recombinant plasmid pTB 64-phb from the transformant.

Then, with pTB 64-phb, *Escherichia coli* HB 101 was transformed by the calcium chloride method to obtain a pTB 64-phb transformant strain.

(Reference Example 3)

Production of scl-PHB synthetase (1)

<Construction of transformant having GST fused scl-PHA synthetase production capacity>

The pTB 64-phb transformant strain was inoculated in 200 ml of an LB medium, and incubated at 37°C, with shaking at 125 strokes/min. After a culture for 12 hours, 200 ml of the culture liquid were inoculated in 200 ml of an LB medium (total 400 ml), and incubated for 12 hours at 37°C, with shaking at 125 strokes/min. The cells were harvested by centrifugation and plasmid DNA was recovered by the normal method.

Then, an oligonucleotide (SEQ ID NO: 3) constituting an upstream primer to the pTB64-phb and

an oligonucleotide (SEQ ID NO: 4) constituting a downstream primer were designed and synthesized respectively (Amersham Pharmacia Biotech). Using these oligonucleotids as primers, PCR was carried out by using pTB 64-PHB as a template to amplify a full length of scl-PHA synthetase gene having a BamHI cleavage site upstream and an XhoI cleavage site downstream (LA-PCR kit; TAKARA SHUZO CO., LTD.).

The purified PCR amplification product was digested by BamHI and XhoI, then inserted into the corresponding restriction sites of plasmid pGEX-6P-1 (Amersham Pharmacia Biotech). An *E. coli* strain (JM109) was transformed with this vector, and consequently a strain for expression was obtained.

For confirmation, the plasmid DNA was prepared using Miniprep (Wizard Minipreps DNA Purification Systems, PROMEGA) in a large amount and digested by BamHI and XhoI, and the resulting DNA fragment was identified.

<Preparation of scl-PHA synthetase>

The obtained expression strain was pre-cultured overnight at 30°C in 100 mL of 2xYT culture medium (polypeptone 16g/L, yeast extract 10 g/L, NaCl 5 g/L, pH 7.0) added with ampicillin (100 µg/L). Then it was added to 10 liters of 2xYT culture medium added with ampicillin (100 µg/L) and culture was carried out for 3 hours at 30°C. Then isopropyl-β-D-thiogalactopyranocide (IPTG) was added to obtain a

final concentration of 1mmol/L, and the culture was continued for 3 hours at 30°C.

The recovered culture liquid was centrifuged for 10 minutes at 4°C, 78,000 m/s² (= 8,000 G), and, after the elimination of supernatant, the pellet was re-suspended in 500 mL of PBS solution at 4°C. The cell suspension was poured, 40 ml each time, in a vessel cooled to 4°C in advance, and, under pressurization of 216 MPA (= 2,200 kg/cm²) by a French press, the cell suspension was released little by little from the nozzle to disrupt the cells. The cell lysate was centrifuged for 10 minutes at 4°C, 78,000 m/s² (= 8,000 G), and the supernatant was recovered. The liquid was filtered with a filter of 0.45 µm to eliminate the solids. The presence of the desired scl-PHA synthetase fused to glutathion transferase (GST) in the supernatant was confirmed by SDS-PAGE.

Then the GST-fused PHB synthesizing enzyme was purified with glutathion sepharose 4B (Amersham Pharmacia Biotech Inc.). 6.65 ml of 75% slurry of glutathion sepharose 4B was centrifuged for 5 minutes at 4°C, 4,900 m/s² (= 500 G), and, after the elimination of supernatant, it was re-suspended in 200 ml of PBS solution at 4°C. Centrifugation was carried out again for 5 minutes at 4°C, 4,900 m/s² (= 500 G), and the supernatant was removed. Then it was re-suspended in 5 ml of PBS solution at 4°C to obtain

50% slurry of glutathion sepharose 4B.

The entire amount of the supernatants prepared before was added to 10 ml of thus obtained 50% slurry of glutathion sepharose 4B, and the mixture was
5 mildly shaken for 30 minutes at the room temperature to cause affinity adsorption of the desired fused protein in the supernatant to the glutathion sepharose 4B. The liquid was centrifuged for 5 minutes at 4°C, 4,900 m/s² (= 500 G), and, after the
10 elimination of supernatant, it was re-suspended in 5 ml of PBS solution at 4°C, and subjected to similar centrifugation again and the supernatant was removed. The glutathion sepharose 4B on which GST-fused scl-
PHA synthetase was immobilized was taken as an
15 immobilized enzyme (1).

Then, after rinsing by repeating re-suspension in PBS solution and centrifuging twice, it was finally suspended in 5 ml of Cleavage buffer (Tris-HCl 50 mmol/L, NaCl 150 mmol/L, EDTA 1 mmol/L,
20 dithiothreitol 1 mmol/L, pH 7). Then 0.5 ml of 4% solution of PreScission Protease (Amarsham Farmacia Biotech) in the cleavage buffer were added, and the mixture was mildly shaken for 4 hours at 5°C. The mixture was centrifuged for 5 minutes at 4°C, 4,900
25 m/s² (= 500 G), and the supernatant was recovered. Then 1 ml of 50% slurry of glutathion sepharose 4B prepared as explained in the foregoing was

centrifuged for 5 minutes at 4°C, 4,900 m/s² (= 500 G), to which the above recovered supernatant was added, and the mixture was mildly agitated to cause glutation sepharose 4B to adsorb PreScission Protease remaining in the supernatant. Then centrifugation was carried out for 5 minutes at 4°C, 4,900 m/s² (= 500 G), and, the supernatant was recovered. The supernatant showed a single band in SDS-PAGE, indicating the purification.

10 The activity of the contained scl-PHA synthetase was measured in the following manner. At first bovine serum albumin (Sigma Co.) was dissolved in 0.1 mol/L tris-HCl buffer (pH 8.0) in 3.0 mg/ml, and 100 µl of thus obtained solution was added to 100 µl of the
15 enzyme solution and the mixture was pre-incubated for 1 minute at 30°C. Then 100 µl of solution of 3-hydroxybutyryl CoA dissolved in 0.1 mol/L tris-HCl buffer (pH 8.0) in 3.0 mmol/L was added, then the mixture was incubated for 1 to 30 minutes at 30°C,
20 and then the reaction was terminated by adding solution of trichloroacetic acid dissolved in 0.1 mol/L tris-HCl buffer (pH 8.0) at 10 mg/ml. The solution after termination of reaction was centrifuged (147,000 m/s² (15,000 G), 10 minutes), and
25 500 µl of 2.0 mmol/L solution of 5,5'-dithiobis-(2-nitrobenzoic acid) dissolved in 0.1 mol/L tris-HCl buffer (pH 8.0) was added to 500 µL of the

supernatant and, after incubation for 10 minutes at 30°C, the optical absorbance at 412 nm was measured. The enzyme activity was calculated by taking an enzyme amount causing release of CoA of 1 μ mol in 1 minute as 1 unit (U). As a result there was obtained a specific activity of 7.5 U/ml. The obtained liquid was concentrated by ultrafiltration under the addition of lyphogel to 10 U/mL, thereby obtaining a purified enzyme liquid (1).

10 (Reference Example 4)

<Preparation of crude enzyme liquid containing scl-PHA synthetase>

The KK01 and TL2 strains were cultured for 24 hours at 30°C in 100 liters of M9 culture medium containing 0.5% of yeast extract and 0.3% of mineral solution (see following), and the recovered culture liquid was centrifuged for 10 minutes at 4°C, 78,000 m/s^2 (= 8,000 G), and, after the elimination of supernatant, the cell pellet was re-suspended in 500 ml of PBS solution at 4°C. The cell suspension was poured, 40 ml each time, in a vessel cooled to 4°C in advance, and, under pressurization of 216 MPa (= 2,200 kg/cm^2) by a French press, the cell suspension was released little by little from the nozzle to disrupt cells. The disrupted cell suspension was centrifuged for 10 minutes at 4°C, 78,000 m/s^2 (= 8,000 G), and the supernatant was recovered. The

liquid was filtered with a filter of 0.45 μm to eliminate the solids, and the activity of the contained scl-PHA synthetase was measured by the aforescribed method. As a result there were
5 obtained relative activities of 1.6 U/mL for the KK01 strain and 1.2 U/mL for the TL2 strain. The liquid was concentrated by ultrafiltration under the addition of lyphogel to 10 U/mL, thereby obtaining crude enzyme liquid (1) derived from the KK01 strain
10 and crude enzyme liquid (2) derived from the TL2 strain.

(Example 1)

The purified scl-PHA synthetase liquid was used to prepare a magnetic capsule construct (1) in the
15 following manner.

To 10 parts by mass of the purified enzyme liquid (1), 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the scl-PHA
20 synthetase to be adsorbed on the surface of the magnetic material. The mixture was centrifuged (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s^2 (= 10,000 G), 4°C, 10
25 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed enzyme-immobilized magnetic

material was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of 3-hydroxybutyryl CoA (Sigma Aldrich Japan Co.) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

A 10 μ l aliquot of the above reaction solution was put on a slide glass, to which 10 μ l of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material was observed to confirm that the magnetic material was coated with PHB on the surface.

As a control, 1 part by mass of the magnetic material (1) was added to 49 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then, after mild shaking for 2.5 hours at 30°C, similarly dyed with Nile blue A and observed under a fluorescence microscope. As a result, the surface of the magnetic material did not show fluorescence at all.

Also a part of the synthesized PHB particles was collected by centrifugation (98,000 m/s² (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to

extract PHB constituting the external coating. The extract was filtered through a membrane filter of 0.45 μm pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the
5 extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS; Shimadzu QP-5050; an EI method) to identify the methyl-esterified monomer unit. As a result, there was confirmed PHB
10 constituted of 3-hydroxybutyric acid unit.

Further, the molecular weight of PHB was evaluated by gel permeation chromatography (GPC; Tosoh HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 μm), solvent: chloroform, column temperature: 40°C,
15 converted as polystyrene) to obtain a result $M_n = 68,000$ and $M_w = 140,000$.

(Example 2)

The crude enzyme liquid containing scl-PHA synthetase was used to prepare a magnetic capsule
20 construct (2) in the following manner.

To 10 parts by mass of the crude enzyme liquid (1), 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken
for 30 minutes at 30°C to cause the scl-PHA synthetase
25 to be adsorbed on the surface of the magnetic material. The mixture was centrifuged ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), and the precipitate was

suspended in PBS solution and centrifuged again (98,000 m/s² (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed enzyme-immobilized magnetic material was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of 3-hydroxybutyryl CoA (Sigma Aldrich Japan Co.) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

A 10 µl aliquot of the above reaction solution was put on a slide glass, to which 10 µl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material was observed to confirm that the magnetic material was coated with PHB on the surface.

As a control, 1 part by mass of the magnetic material (1) was added to 49 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then, after mild shaking for 2.5 hours at 30°C, similarly dyed with Nile blue A and observed under a fluorescence microscope. As a result, the surface of the magnetic material did not show fluorescence at all.

Also a part of the synthesized PHB particles was collected by centrifugation ($98,000 \text{ m/s}^2$ (= $10,000 \text{ G}$), 4°C , 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHB constituting the external coating. The extract was filtered through a membrane filter of $0.45 \mu\text{m}$ pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified monomer unit. As a result, there was confirmed PHB constituted of 3-hydroxybutyric acid unit.

Further, the molecular weight of PHB was evaluated by gel permeation chromatography (GPC: Toso HLC-8020, column: Polymer Laboratory PLgel MIXED-C ($5 \mu\text{m}$), solvent: chloroform, column temperature: 40°C , converted as polystyrene) to obtain a result $M_n = 59,000$ and $M_w = 130,000$.

(Example 3)

The crude enzyme liquid containing scl-PHA synthetase was used to prepare a magnetic capsule construct (3) in the following manner.

To 10 parts by mass of the crude enzyme liquid (2), 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken.

for 30 minutes at 30°C to cause the scl-PHA synthetase to be adsorbed on the surface of the magnetic material. The mixture was centrifuged (98,000 m/s² (= 10,000 G), 4°C, 10 minutes), and the precipitate was
5 suspended in PBS solution and centrifuged again (98,000 m/s² (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate
10 buffer (pH 7.0), then 1 part by mass of 3-hydroxybutyryl CoA (Sigma Aldrich Japan Co.) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

15 A 10 µl aliquot of the above reaction solution was put on a slide glass, to which 10 µl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence
20 microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material was observed to confirm that the magnetic material was coated with PHB on the surface.

25 As a control, 1 part by mass of the magnetic material (1) was added to 49 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then, after mild

shaking for 2.5 hours at 30°C, similarly dyed with Nile blue A and observed under a fluorescence microscope. As a result, the surface of the magnetic material did not show fluorescence at all.

5 Also a part of the synthesized PHB particles was collected by centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHB constituting the external coating. The
10 extract was filtered through a membrane filter of 0.45 μm pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas
15 chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified monomer unit. As a result, there was confirmed PHB constituted of 3-hydroxybutyric acid unit.

Further, the molecular weight of PHB was
20 evaluated by gel permeation chromatography (GPC: Tosoh HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 μm), solvent: chloroform, column temperature: 40°C, converted as polystyrene) to obtain a result $M_n = 65,000$ and $M_w = 160,000$.

25 (Example 4)

The purified scl-PHA synthetase liquid was used to prepare a magnetic capsule construct (4) in the

following manner.

To 10 parts by mass of the purified enzyme liquid (1), 1 part by mass of nickel powder of a primary particle size of $0.02\ \mu\text{m}$ (Ni(200)UFMP, Shinku Yakin Co.) as the magnetic material (2) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the scl-PHA synthetase to be adsorbed on the surface of the magnetic material. The mixture was centrifuged ($98,000\ \text{m/s}^2$ ($= 10,000\ \text{G}$), 4°C , 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again ($98,000\ \text{m/s}^2$ ($= 10,000\ \text{G}$), 4°C , 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of $0.1\ \text{mol/L}$ phosphate buffer (pH 7.0), then 1 part by mass of 3-hydroxybutyryl CoA (Sigma Aldrich Japan Co.) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C .

A $10\ \mu\text{l}$ aliquot of the above reaction solution was put on a slide glass, to which $10\ \mu\text{l}$ of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a

result, fluorescence from the surface of the magnetic material was observed to confirm that the magnetic material was coated with PHB on the surface.

As a control, 1 part by mass of nickel powder of a primary particle size of 0.02 μm (Ni(200)UFMP, Shinku Yakin Co.) as the magnetic material (2) was added to 49 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then, after mild shaking for 2.5 hours at 30°C, similarly dyed with Nile blue A and observed under a fluorescence microscope. As a result, the surface of the magnetic material did not show fluorescence at all.

Also a part of the synthesized PHB particles was collected by centrifugation (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHB constituting the external coating. The extract was filtered through a membrane filter of 0.45 μm pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified monomer unit. As a result, there was confirmed PHB constituted of 3-hydroxybutyric acid unit.

Further, the molecular weight of PHB was

evaluated by gel permeation chromatography (GPC: Tosoh HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 μ m), solvent: chloroform, column temperature: 40°C, converted as polystyrene) to obtain a result $M_n =$
5 78,000 and $M_w = 170,000$.

(Example 5)

The purified scl-PHA synthetase liquid was used to prepare a magnetic capsule construct (5) in the following manner.

- 10 To 10 parts by mass of the purified enzyme liquid (1), 1 part by mass of γ -Fe₂O₃ fine powder of a primary particle size of 0.02 μ m (NanoTel, CI Chemical Co.) as the magnetic material (3) and 39 parts by mass of PBS were added and mildly shaken for
15 30 minutes at 30°C to cause the scl-PHA synthetase to be adsorbed on the surface of the magnetic material. The mixture was centrifuged (98,000 m/s² (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s² (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-
20 immobilized magnetic material.

- The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of 3-
25 hydroxybutyryl CoA (Sigma Aldrich Japan Co.) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2

hours at 30°C.

A 10 μ l aliquot of the above reaction solution was put on a slide glass, to which 10 μ l of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material was observed to confirm that the magnetic material was coated with PHB on the surface.

As a control, 1 part by mass of γ -Fe₂O₃ fine powder of a primary particle size of 0.02 μ m (NanoTek, CI Chemical Co.) as the magnetic material (3) was added to 49 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then, after mild shaking for 2.5 hours at 30°C, similarly dyed with Nile blue A and observed under a fluorescence microscope. As a result, the surface of the magnetic material did not show fluorescence at all.

Also a part of the synthesized PHB particles was collected by centrifugation (98,000 m/s² (= 10,000 G), 4°C, 10 minutes); dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHB constituting the external coating. The extract was filtered through a membrane filter of 0.45 μ m pore size and concentrated under a reduced

pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified monomer unit. As a result, there was confirmed PHB constituted of 3-hydroxybutyric acid unit.

Further, the molecular weight of PHB was evaluated by gel permeation chromatography (GPC: Tosoh HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 μ m), solvent: chloroform, column temperature: 40°C, converted as polystyrene) to obtain a result. M_n = 80,000 and M_w = 170,000.

(Example 6)

The purified scl-PHA synthetase liquid was used to prepare a magnetic capsule construct (6) in the following manner.

To 10 parts by mass of the purified enzyme liquid (1), 1 part by mass of magnetite fine powder of a primary particle size of 0.3 μ m (Magnetite EPT500, Toda Kogyo Co.) as the magnetic material (4) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the scl-PHA synthetase to be adsorbed on the surface of the magnetic material. The mixture was centrifuged (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and

centrifuged again ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was
5 suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of 3-hydroxybutyryl CoA (Sigma Aldrich Japan Co.) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2
10 hours at 30°C.

A 10 μl aliquot of the above reaction solution was put on a slide glass, to which 10 μl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with
15 a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material was observed to confirm that the magnetic
20 material was coated with PHB on the surface.

As a control, 1 part by mass of magnetite fine powder of a primary particle size of 0.3 μm (Magnetite EPT500, Toda Kogyo Co.) as the magnetic
material (4) was added to 49 parts by mass of 0.1
25 mol/L phosphate buffer (pH 7.0), then, after mild shaking for 2.5 hours at 30°C, similarly dyed with Nile blue A and observed under a fluorescence

microscope. As a result, the surface of the magnetic material did not show fluorescence at all.

Also a part of the synthesized PHB particles was collected by centrifugation ($98,000 \text{ m/s}^2$ ($= 10,000 \text{ G}$), 4°C , 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHB constituting the external coating. The extract was filtered through a membrane filter of $0.45 \mu\text{m}$ pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS; Shimadzu QP-5050, an EI method) to identify the methyl-esterified monomer unit. As a result, there was confirmed PHB constituted of 3-hydroxybutyric acid unit.

Further, the molecular weight of PHB was evaluated by gel permeation chromatography (GPC: Tosoh HLC-8020, column: Polymer Laboratory PLgel MIXED-C ($5 \mu\text{m}$), solvent: chloroform, column temperature: 40°C , converted as polystyrene) to obtain a result $M_n = 79,000$ and $M_w = 180,000$.

(Example 7)

The purified scl-PHA synthetase liquid was used to prepare a magnetic capsule construct (7) in the following manner.

To 10 parts by mass of the purified enzyme

liquid (1), 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the scl-PHA synthetase to be adsorbed on the surface of the magnetic material. The mixture was centrifuged (98,000 m/s² (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s² (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of 3-hydroxybutyryl CoA (Sigma Aldrich Japan Co.), 1 part by mass of polyethylene glycol 200 (PEG200, Kishida Kagaku Co., average molecular weight 190 to 210) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

A 10 µl aliquot of the above reaction solution was put on a slide glass, to which 10 µl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic

material was observed to confirm that the magnetic material was coated with PHB on the surface.

As a control, 1 part by mass of the magnetic material (1) was added to 49 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then, after mild shaking for 2.5 hours at 30°C, similarly dyed with Nile blue A and observed under a fluorescence microscope. As a result, the surface of the magnetic material did not show fluorescence at all.

Also a part of the synthesized PHB particles was collected by centrifugation (98,000 m/s² (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHB constituting the external coating. The extract was filtered through a membrane filter of 0.45 µm pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified monomer unit. As a result, there was confirmed PHB constituted of 3-hydroxybutyric acid unit.

Also a detailed structure analysis by ¹H-NMR (FT-NMR: Bruker DPX400; ¹H resonance frequency: 400 MHz; measured nuclide: ¹H; solvent: CDCl₃; reference: capillary-sealed TMS/CDCl₃; measuring temperature:

room temperature) confirmed, in addition to peaks derived from PHA constituted of 3-hydroxybutyric acid unit, peaks derived from polyethylene glycol at 3.5 to 3.8 ppm and about 4.2 ppm.

5 Further, the molecular weight of PHB was evaluated by gel permeation chromatography (GPC: Tosoh HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 μ m), solvent: chloroform, column temperature: 40°C, converted as polystyrene) to obtain a result Mn =
10 22,000 and Mw = 45,000.

(Example 8)

A ferrite sheet of 30 mm x 30 mm x 3 mm (NP-S01, Nippon Paint Co., a dispersion of ferrite particles in resin) was immersed for 1 hour in 1%
15 glutaraldehyde, then rinsed with purified water and immersed in the purified enzyme liquid (1) for 30 minutes at 30°C to fix the enzyme. The unreacted scl-PHA synthetase was removed by rinsing with PBS solution to obtain enzyme-immobilized magnetic
20 material.

The aforescribed immobilized enzyme was immersed in 0.1 mol/L phosphate buffer (pH 7.0), containing 30 mmol/L of 3-hydroxybutyryl CoA (Sigma Aldrich Japan Co.) and 0.1% of bovine serum albumin
25 (Sigma Co.) and the mixture was mildly shaken for 2 hours at 30°C. After the reaction, unreacted substance etc. were removed by rinsing with 0.1 mol/L

phosphate buffer (pH 7.0).

The ferrite sheet after the reaction was dyed with a 1% aqueous solution of Nile blue A and was observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the ferrite sheet was observed to confirm that the construct was a laminar construct in which a base material of the ferrite sheet was covered by a film of PHB.

Also the laminar construct was dried in vacuum, and immersed in chloroform under agitation for 20 hours at 60°C to extract PHB constituting the coating layer. The extract was filtered through a membrane filter of 0.45 μ m pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified monomer unit. As a result, there was confirmed PHB constituted of 3-hydroxybutyric acid unit.

(Example 9)

Evaluation of coating property of magnetic capsule construct

In order to confirm whether the magnetic particles were completely protected and covered with

the polymer, 0.1 g each of the obtained magnetic capsule constructs (1) to (7) were immersed for 2 hours in 100 ml of pure water heated to 70°C and a metal content was measured in the liquid. As a result, the metal content in the water was 3 ppm or less with all the capsule constructs. Based on these facts, these capsule constructs were judged that "metal ions did not elute."

In the following there will be shown examples relating to synthesizing enzymes for mcl-PHA and unusual-PHA (reference examples 5 to 7, examples 11 to 28),

(Reference Example 5)

<Construction of transformant having mcl-PHA synthetase production capacity>

Strain YN2 was cultured in 100 ml of LB medium (1% polypeptone (NIPPON SEIYAKU CO., LTD.), 0.5% yeast extract (Difco), 0.5% sodium chloride, pH 7.4) overnight at 30°C, then the chromosomal DNA was isolated by the method of Marmar et al. The obtained chromosomal DNA was completely digested by a restriction enzyme HindIII. A vector pUC18 was also cut by HindIII. After terminal dephosphorylation (Molecular Cloning, 1, 572, (1989); Cold Spring Harbor Laboratory Press (aforescribed), complete HindIII digestion fragments of the chromosomal DNA were ligated to the HindIII cleavage site of the

vector (a cloning site) using a DNA ligation kit Ver. II (TAKARA SHUZO CO., LTD.). With these plasmid vectors containing chromosomal DNA fragments, *Escherichia coli* HB 101 was transformed to construct a DNA library of strain YN2.

Next, to select DNA fragments covering the PHA synthetase gene of strain YN2, a probe for colony hybridization was prepared. Oligonucleotides of SEQ ID NO: 6 and SEQ ID NO: 7 were synthesized (Amersham Pharmacia Biotech), and PCR of chromosomal DNA of YN2 was carried out by using these oligonucleotides as primers. The PCR-amplified DNA fragments were used as a probe. Labeling of the probe was conducted by employing a commercially available labeling enzyme Alk Phos Direct (Amersham Pharmacia Biotech). Using the obtained labeled probe for colony hybridization, an *Escherichia coli* strain having the recombinant plasmid containing the mcl-PHA synthetase gene was selected from the chromosomal DNA library of YN2 by the colony hybridization method. The plasmid was recovered from the selected strain by the alkali process to give DNA fragment including the mcl-PHA synthetase gene.

This gene fragment was inserted into a vector pBBR122 having a wide-host-replication range (Mo Bi Tec) not belonging to any of IncP, IncQ, or IncW incompatibility group. When *Pseudomonas cichorii*

YN2ml (a PHA synthesis negative strain) was transformed with this recombinant plasmid by electroporation, the PHA synthesizing capacity of the YN2ml strain was recovered to show complementarity.

5 Consequently, it was confirmed that the selected gene fragment contained mcl-PHA synthetase gene region translatable into mcl-PHA synthetase in *Pseudomonas cichorii* YN2ml.

10 The base sequence of this DNA fragment was determined by the Sanger method. As a result, it was shown that there were base sequences represented by SEQ ID NO: 8 and SEQ ID NO: 9 each encoding a polypeptide. With respect to these mcl-PHA synthetase genes, PCR was carried out by using the chromosomal
15 DNA as a template to produce the complete mcl-PHA synthetase gene. More specifically, an upstream primer (SEQ ID NO: 10) and a downstream primer (SEQ ID NO: 11) corresponding to the mcl-PHA synthetase gene of SEQ ID NO: 8, and an upstream primer (SEQ ID
20 NO: 12) and a downstream primer (SEQ ID NO: 13) corresponding to the mcl-PHA synthetase gene of SEQ ID NO: 9 were synthesized respectively (Amersham Pharmacia Biotech).

25 Using these primers, PCR was carried out for each of the base sequences shown by SEQ ID NO: 8 and SEQ ID NO: 9, then a full length of mcl-PHA synthetase gene was amplified (LA-PCR kit; TAKARA

SHUZO CO., LTD.). Next, the obtained PCR amplified fragment and an expression vector pTrc99A were digested by the restriction enzyme HindIII and dephosphorylated (Molecular Cloning, vol.1, p.572, (1989); Cold Spring Harbor Laboratory Press, then the DNA fragment including a full length PHA synthetase gene excluding unnecessary base sequences at both terminuses was linked to a restriction site of the expression vector pTrc99A by using a DNA ligation kit Ver. II (TAKARA SHUZO CO., LTD.).

An *E. coli* strain (*Escherichia coli* HB101: TAKARA SHUZO) was transformed with each of the obtained recombinant plasmids by the calcium chloride method. The obtained recombinants were cultured and the recombinant plasmids were amplified, then the recombinant plasmids were respectively recovered. The recombinant plasmid having a DNA of SEQ ID NO: 8 was designated as pYN2-C1, and the recombinant plasmid having a DNA of SEQ ID NO: 9 was designated as pYN2-C2. An *E. coli* strain (*Escherichia coli* HB101fB fadB deletion strain) was transformed with pYN2-C1 and pYN2-C2 respectively by the calcium chloride method to obtain recombinant *E. coli* strains having respective recombinant plasmids, i.e., a pYN2-C1 recombinant strain and a pYN2-C2 recombinant strain.

(Reference Example 6)

Production of mcl-PHA synthetase 1)

<Construction of transformant having mcl-PHA synthetase production capacity>

For the pYN2-C1, an upstream primer (SEQ ID NO: 14) and a downstream primer (SEQ ID NO: 15) were designed and synthesized respectively (Amersham Pharmacia Biotech). PCR was carried out using these primers and template pYN2-C1 to synthesize a full length PHA synthetase gene having a BamHI restriction site upstream and a XhoI restriction site downstream (LA-PCR kit, TAKARA SHUZO CO., LTD.).

Similarly, for pYN2-C2, an upstream primer (SEQ ID NO: 16) and a downstream primer (SEQ ID NO: 17) were designed and synthesized respectively (Amersham Pharmacia Biotech). PCR was carried out using these primers and the template pYN2-C2 to amplify the full length PHA synthetase gene having a BamHI restriction site upstream and a XhoI restriction site downstream (LA-PCR kit, TAKARA SHUZO CO., LTD.).

Respective purified PCR amplification products were digested by BamHI and XhoI, then inserted into the corresponding restriction sites of plasmid pGEX-6P-1 (Amersham Pharmacia Biotech). An *E. coli* strain JM109 was transformed with these vectors to obtain expressing strains. For confirmation, each plasmid DNA was prepared by Miniprep (Wizard Minipreps DNA Purification Systems, PROMEGA) in a large amount and digested by BamHI and XhoI, and the resulting DNA

fragment was identified.

<Preparation of mcl-PHA synthetase>

Each obtained strain was pre-cultured in 10 ml of LB-Amp medium overnight, and then an 0.1 ml culture was transferred to 10 ml of LB-Amp medium and cultured at 37°C, 170 rpm for 3 hours under shaking. Then, IPTG was added to the culture (enc concentration 1 mmol/L), then the culture was continued for 4 to 12 hours at 37°C.

10 The *E. coli* cells induced with IPTG were collected ($78,000 \text{ m/s}^2$ (= 8,000 G), 2 minutes, 4°C) and resuspended in a 1/10 volume of phosphate buffer physiological saline (PBS; 8 g NaCl, 1.44 g Na_2HPO_4 , 0.24 g KH_2PO_4 , 0.2 g KCl, 1,000 ml purified water) at 15 4°C. The cells were disrupted by freeze and thawing and sonication, and subjected to centrifugation ($78,000 \text{ m/s}^2$ (= 8,000 G), 10 minutes, 4°C) to remove solid impurities. Confirming that the aimed expressing protein was present in the supernatant by 20 SDS-PAGE, the induced and expressed GST fusion protein was purified by using Glutathione Sepharose 4B (Amersham Pharmacia Biotech). The Glutathione Sepharose was previously treated to avoid nonspecific adsorption, that is, the Glutathione Sepharose was 25 washed with an equivalent amount of PBS for three times ($78,000 \text{ m/s}^2$ (= 8,000 G), 1 minutes, 4°C), and then an equivalent amount of 4% bovine serum albumin

PBS was added thereto and processed at 4°C for one hour. After the process, the Sepharose was washed with an equivalent amount of PBS twice, and re-suspended in an 1/2 amount of PBS. The pre-treated 40
5 μ l of Glutathione Sepharose was added to 1 ml of the above cell free extract, and gently stirred at 4°C to adsorb fusion proteins GST-YN2-C1 and GST-YN2-C2 onto Glutathione Sepharose respectively. After centrifugation ($78,000 \text{ m/s}^2$ (= 8,000 G), 1 minutes,
10 4°C) to collect the Glutathione Sepharose, it was washed with 400 μ l of PBS for three times. Thereafter, 40 μ l of 10 mmol/L of glutathione was added thereto and stirred for one hour at 4°C to elute the adsorbed fusion protein. After centrifugation ($78,000 \text{ m/s}^2$ (= 8,000 G), 2 minutes, 4°C), the supernatant was
15 recovered and dialyzed against PBS to purify the GST fusion protein. A single band was confirmed by SDS-PAGE.

Then 500 μ g of each GST fusion protein was
20 digested by PreScission protease (Amersham Pharmacia Biotech, 5U), and the protease and the GST were removed therefrom by passing through Glutathione Sepharose. The flow-through fraction was further loaded to Sephadex G200 column equilibrated with PBS,
25 then expression proteins YN2-C1 and YN2-C2 were obtained as final purified products. By SDS-PAGE, single bands (60.8 kDa and 61.5 kDa, respectively)

were confirmed.

The above described enzymes were concentrated with a bioliquid concentrating agent (Mizubutorikun AB-1100, Atto Corp.) to obtain 10 U/ml of purified enzyme solutions.

The enzyme activity was measured by the above-described method. The protein concentration of the sample was determined by using a micro BCA protein assay reagent kit (Pierce Chemical Co.). The results are shown in Table 1.

Table 1

	Specific Activity
pYN2-C1	4.1 U/mg protein
pYN2-C2	3.6 U/mg protein

(Reference Example 7)

Production of mcl-PHA synthetase. (2)

Each of strains P91, H45, YN2, and P161 was inoculated in 200 ml of an M9 medium containing 0.5% of yeast extract (Difco) and 0.1% of octanoic acid, and incubated at 30°C, under shaking at 125 strokes/min. After 24 hours, the cells were harvested by centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), then the cells were re-suspended in 200 ml of 0.1 mol/L Tris-HCl buffer (pH 8.0) and further subjected to centrifugation for washing. The cells were re-suspended in 2.0 ml of 0.1 mol/L Tris-HCl

buffer (pH 8.0) and disrupted by an ultrasonic homogenizer and then centrifuged ($118,000 \text{ m/s}^2$ (= 12,000 G), 4°C , 10 minutes) to collect the supernatant thereby obtaining the crude enzyme.

5 Each purified enzyme activity was measured by the above described method, and the result is shown in Table 2.

Table 2

	Activity
P91	0.1 U/ml
H45	0.2 U/ml
YN2	0.4 U/ml
P161	0.2 U/ml

10 (Example 11)

Preparation of magnetic capsule construct (8)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (1). The mixture was centrifuged ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C , 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C , 10 minutes) to obtain enzyme-immobilized magnetic

material.

The aforedescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of 3-hydroxyoctanoyl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

A 10 µl aliquot of the above reaction solution was put on a slide glass, to which 10 µl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (1) was observed to confirm that the magnetic material (1) was coated with PHA on the surface. This product was taken as a magnetic capsule construct (8).

As a control, 1 part by mass of the magnetic material (1) was added to 49 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then, after mild shaking for 2.5 hours at 30°C, similarly dyed with Nile blue A and observed under a fluorescence microscope. As a result, the surface of the magnetic material (1) did not show fluorescence at all.

Also a part of the synthesized PHA particles was collected by centrifugation ($98,000 \text{ m/s}^2$ ($= 10,000 \text{ G}$), 4°C , 10 minutes); dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to
5 extract PHA constituting the external coating. The extract was filtered through a membrane filter of $0.45 \mu\text{m}$ pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a
10 conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified PHA monomer unit. As a result, as shown in Fig. 1, there was confirmed PHA constituted of 3-
15 hydroxyoctanoic acid unit.

Further, the molecular weight of PHA was evaluated by gel permeation chromatography (GPC: Toso HLC-8020, column: Polymer Laboratory PLgel MIXED-C ($5 \mu\text{m}$), solvent: chloroform, column temperature: 40°C ,
20 converted as polystyrene) to obtain a result $M_n = 20,000$ and $M_w = 39,000$.

(Example 12)

Preparation of magnetic capsule construct (9)

To 10 parts by mass of the mcl-PHA synthetase
25 solution (10 U/ml) derived from pYN2-C2 recombinant strain, 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly

shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (1). The mixture was centrifuged (98,000 m/s² (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s² (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of (R)-3-hydroxyoctanoyl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997) (aforescribed) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

A 10 µl aliquot of the above reaction solution was put on a slide glass, to which 10 µl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (1) was observed to confirm that the magnetic material (1) was coated with PHA on the surface. This product was taken as a magnetic capsule

construct (9).

Also a part of the particles was collected by centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C , 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was filtered through a membrane filter of $0.45 \mu\text{m}$ pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified PHA monomer unit. As a result, as in example 11, there was confirmed PHA constituted of 3-hydroxyoctanoic acid unit.

(Example 13)

Preparation of magnetic capsule constructs (10) to (13)

To 99 parts by mass of the crude mcl-PHA synthetase derived from strain YN2, H45, P91 or P161, 1 part by mass of the magnetic material (1) was added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (1). The mixture was centrifuged ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C , 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again ($98,000 \text{ m/s}^2$ (= 10,000

G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

Each enzyme-immobilized magnetic material was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of (R)-3-hydroxyoctanoyl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

A 10 µl aliquot of the above reaction solution was put on a slide glass, to which 10 µl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, in any of the reaction liquids of the enzyme-immobilized magnetic materials, fluorescence from the surface of the magnetic material (1) was observed to confirm that the magnetic material (1) was coated with PHA on the surface. These products were taken as magnetic capsule constructs (10) to (13).

Also a part of the particles was collected by centrifugation (98,000 m/s² (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was

filtered through a membrane filter of 0.45 μm pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified PHA monomer unit. As a result, as in example 11, there was confirmed PHA constituted of 3-hydroxyoctanoic acid unit.

10 (Example 14)

Preparation of magnetic capsule construct (14)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (1). The mixture was centrifuged (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of (R)-3-hydroxy-5-phenylvaleryl CoA (prepared by hydrolyzing

3-hydroxyphenylvaleryl ester, obtained by a Reformatsky reaction, to obtain 3-hydroxy-5-phenylvaleric acid, and then following a method described in Eur. J. Biochem., 250, 432-439 (1997)) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

A 10 μ l aliquot of the above reaction solution was put on a slide glass, to which 10 μ l of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (1) was observed to confirm that the magnetic material (1) was coated with PHA on the surface. This product was taken as a magnetic capsule construct (14).

Also a part of the particles was collected by centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was filtered through a membrane filter of 0.45 μm pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was

subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified PHA monomer unit. As a result, as shown in Fig. 2, there was confirmed PHA constituted of 3-hydroxy-5-phenylvaleric acid unit.

Further, the molecular weight of PHA was evaluated by gel permeation chromatography (GPC: Toso HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 μ m), solvent: chloroform, column temperature: 40°C, converted as polystyrene) to obtain a result $M_n = 17,000$ and $M_w = 35,000$.

(Example 15)

Preparation of magnetic capsule construct (15)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (1). The mixture was centrifuged (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was

suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of (R)-3-hydroxy-5-(4-fluorophenyl)valeryl CoA (prepared by hydrolyzing 3-hydroxyphenylvaleryl ester, obtained by a Reformatsky reaction, to obtain 3-hydroxy-5-(4-fluorophenyl)valeric acid, and then following a method described in Eur. J. Biochem., 250, 432-439 (1997)) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

A 10 µl aliquot of the above reaction solution was put on a slide glass, to which 10 µl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (1) was observed to confirm that the magnetic material (1) was coated with PHA on the surface. This product was taken as a magnetic capsule construct (15).

Also a part of the particles was collected by centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was

filtered through a membrane filter of 0.45 μm pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified PHA monomer unit. As a result, as shown in Fig. 3, there was confirmed PHA constituted of 3-hydroxy-5-(4-fluorophenyl)valeric acid unit.

(Example 16)

Preparation of magnetic capsule construct (16)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (1). The mixture was centrifuged (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of (R)-3-

hydroxyoctanoyl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

5 A 10 μ l aliquot of the above reaction solution was put on a slide glass, to which 10 μ l of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence
10 microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (1) was observed to confirm that the magnetic material (1) was coated with PHA on the
15 surface. This product was taken as a magnetic capsule construct (16).

Also the capsule construct was recovered by a centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), and, after drying, the mass of the polymer
20 formed on the surface of the construct was measured by a time-of-flight secondary ion mass spectrometer (TOF-SIMS IV, CAMECA). The obtained mass spectrum confirmed that the surface of the capsule construct was principally comprised of a homopolymer of
25 polyhydroxy octanoate. Also the measurements of the similar TOF-SIMS mass spectrum under gradual scraping of the surface of the capsule construct by ion

sputtering confirmed a homopolymer of polyhydroxy octanoate in all the cases. These results confirmed that the capsule construct of the present example was formed by coating the hydrophilic inorganic particles directly with the hydrophobic homopolymer of polyhydroxy octanoate.

Further, the molecular weight of PHA was evaluated by gel permeation chromatography (GPC: Toso HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 μ m), solvent: chloroform, column temperature: 40°C, converted as polystyrene) to obtain a result $M_n = 25,000$ and $M_w = 47,000$.

(Example 17)

Preparation of magnetic capsule construct (17)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (1). The mixture was centrifuged (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was

suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of (R)-3-hydroxyoctanoyl CoA (prepared according to J. Bacteriol., 182, 2753-2760 (2000)) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 10 minutes at 30°C. Then, to this reaction liquid under mild shaking at 30°C, a 0.1 mol/L phosphate buffer (pH 7.0), containing 1 part by mass of (R)-3-hydroxyoctanoyl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)) and 0.1 parts by mass of bovine serum albumin (Sigma Co.), was added at a rate of 1 parts by mass per minute by a microtube pump (MP-3N, Tokyo Rikakikai Co.). After 1 hour and 30 minutes, the generated granulates were recovered by centrifugation (98000 m/s², 4°C, 10 minutes), and, after elimination of the supernatant, 25 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), containing 1 part by mass of (R)-3-hydroxyoctanoyl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)) and 0.1 parts by mass of bovine serum albumin (Sigma Co.), were added to the granulates and the mixture was mildly shaken for 20 minutes at 30°C.

After the reaction, a 10 µl aliquot of the above reaction solution was put on a slide glass, to which 10 µl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass,

covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (1) was observed to confirm that the magnetic material (1) was coated with PHA on the surface. This product was taken as a magnetic capsule construct (17).

Also the capsule construct was recovered by a centrifugation ($98,000 \text{ m/s}^2$ ($= 10,000 \text{ G}$), 4°C , 10 minutes), and, after drying, the mass of the polymer formed on the surface of the construct was measured by a time-of-flight secondary ion mass spectrometer (TOF-SIMS IV, CAMECA). The obtained mass spectrum confirmed that the surface of the capsule construct was comprised of a homopolymer of polyhydroxy octanoate. Also the measurements of the similar TOF-SIMS mass spectrum under gradual scraping of the surface of the capsule construct by ion sputtering revealed a copolymer of 3-hydroxyoctanoic acid and 3-hydroxypimelic acid (molar ratio 21:1), in which the proportion of 3-hydroxyoctanoic acid gradually decreased while that of 3-hydroxypimelic acid gradually increased toward the interior of the granulate, and which eventually changed to a homopolymer of polyhydroxy pimelate. These results confirmed that the capsule construct of the present

example was formed by coating the hydrophilic granular base material with polyhydroxy pimelate having a hydrophilic function group, then with a copolymer of 3-hydroxy pimelic acid having a hydrophilic functional group and 3-hydroxy octanoic acid having a hydrophobic functional group with a gradually increasing proportion of 3-hydroxy octanoic acid toward the outer surface, and finally coating the outermost layer with a homopolymer of polyhydroxy octanoate.

Further, the molecular weight of PHA was evaluated by gel permeation chromatography (GPC: Tosoh HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 μ m), solvent: chloroform, column temperature: 40°C, converted as polystyrene) to obtain a result $M_n = 23,000$ and $M_w = 43,000$.

(Example 18)

Preparation of magnetic capsule constructs (18) to (21)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (1). The mixture was centrifuged (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes), and the

precipitate was suspended in PBS solution and centrifuged again ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C , 10 minutes) to obtain enzyme-immobilized magnetic material.

5 The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 0.8 part by mass of (R,S)-3-hydroxy-5-phenoxyvaleryl CoA (prepared by hydrolyzing 3-hydroxy-5-phenoxyvaleryl ester, obtained by a
10 Reformatsky reaction of 3-phenoxypropanal and ethyl bromoacetate, to obtain 3-hydroxy-5-phenoxyvaleric acid and then according to a process in Eur. J. Biochem., 250, 432-439 (1997)), 0.2 parts by mass of (R, S)-3-hydroxy-7,8-epoxyoctanoyl CoA (prepared by
15 epoxylating an unsaturated part of 3-hydroxy-7-octenoic acid, prepared by a method in Int. J. Biol. Macromol., 12, 85-91 (1990)) and then according to a process in Eur. J. Biochem., 250, 432-439 (1997)), and 0.1 parts by mass of bovine serum albumin (Sigma
20 Co.) were added and the mixture was mildly shaken for 2 hours at 30°C to obtain a magnetic capsule construct (18).

As a comparative reference, a magnetic capsule construct (19) was prepared in the same manner as
25 explained above, except that (R, S)-3-hydroxy-7,8-epoxyoctanoyl CoA was replaced by 3-hydroxyoctanoyl CoA.

A 10 μ l aliquot of the above sample was put on a slide glass, to which 10 μ l of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and
5 observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, in each sample, fluorescence from the surface of the magnetic material (1) was observed to confirm that the
10 magnetic material (1) was coated with PHA on the surface.

As a control, 1 part by mass of the magnetic material (1) was added to 49 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then, after mild
15 shaking for 2.5 hours at 30°C, similarly dyed with Nile blue A and observed under a fluorescence microscope. As a result, the surface of the magnetic material (1) did not show fluorescence at all.

Also a part of the synthesized PHA particles was
20 collected by centrifugation (98,000 m/s² (= 10,000 G), 4°C, 10 minutes); dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was subjected to an analysis by ¹H-NMR
25 (equipment: FT-NMR: Bruker DPX400; measured nuclide: ¹H; solvent: deuterionized chloroform (containing TMS)). Percentages of the side chain units calculated from

such a measurement are shown in Table 3.

Table 3

Monomer Unit	Magnetic Capsule Construct (18)	Magnetic Capsule Construct (19)
3-hydroxy-5-phenoxy Valeric Acid	75%	71%
3-hydroxy-7,8-epoxy Octanoic Acid	25%	-
3-hydroxy Octanoic Acid	-	29%

5 There were repeated three times operations of
subjecting 50 parts by mass of the aforescribed
magnetic capsule construct (18) to centrifugation
(98,000 m/s² (= 10,000 G), 4°C, 10 minutes) to recover
the capsule construct and suspending the same in 50
10 parts by mass of purified water, and 0.5 parts by
mass of hexamethylene diamine were dissolved in the
suspension. After dissolution was confirmed, water
was removed by lyophilization (thus providing a
magnetic capsule construct (20)). Then the magnetic
15 capsule construct (20) was reacted for 12 hours at
70°C (thus providing a magnetic capsule construct
(21)).

Each of the magnetic capsule constructs (20) and
(21) was suspended in chloroform, and agitated for 20
20 hours at 60°C to extract PHA constituting the outer
coating, which, after elimination of chloroform by
drying under vacuum, was subjected to a differential

scanning calorimeter (DSC; Perkin Elmer Inc., Pyris 1, temperature increase rate: 10°C/min). As a result, the magnetic capsule construct (20) showed a clear exothermic peak at about 90°C, indicating that a reaction between an epoxy group in the polymer and hexamethylene diamine took place and the crosslinking between polymers was proceeding. On the other hand, the magnetic capsule construct (21) did not show a clear heat flow, indicating that the crosslinking reaction was almost completed.

Also infrared absorption was measured on similar samples (FT-IR; Perkin Elmer, 1720X). As a result, peaks for amine (around 3340 cm^{-1}) and epoxy (around 822 cm^{-1}) observed in the magnetic capsule construct (20) disappeared in the magnetic capsule construct (21).

These results indicate that a crosslinked polymer can be obtained by reacting PHA having an epoxy unit in the side chain and hexamethylene diamine.

On the other hand, a similar evaluation was conducted on the magnetic capsule construct (19) as a comparative reference, but a result clearly showing the mutual crosslinking of polymers as explained before could not be obtained.

(Example 19)

Preparation of magnetic capsule constructs (22)

to (23)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (1). The mixture was centrifuged (98,000 m/s² (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s² (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 0.8 part by mass of (R,S)-3-hydroxy-5-phenoxyvaleryl CoA (prepared by hydrolyzing 3-hydroxy-5-phenoxyvaleryl ester, obtained by a Reformatsky reaction of 3-phenoxypropanal and ethyl bromoacetate, to obtain 3-hydroxy-5-phenoxyvaleric acid and then following a process in Eur. J. Biochem., 250, 432-439 (1997)), 0.2 parts by mass of (R, S)-3-hydroxy-7,8-epoxyoctanoyl CoA (prepared by epoxylating an unsaturated part of 3-hydroxy-7-octenoic acid, prepared by a process in Int. J. Biol. Macromol., 12, 85-91 (1990)) and then following a process in Eur. J. Biochem., 250, 432-439 (1997)),

and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C to obtain a magnetic capsule construct (22).

5 10 µl of the aforescribed magnetic capsule construct (22) was put on a slide glass, to which 10 µl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a
10 fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, in each sample, fluorescence from the surface of the magnetic material (1) was observed to confirm that the magnetic material (1)
15 was coated with PHA on the surface.

As a control, 1 part by mass of the magnetic material (1) was added to 49 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then, after mild shaking for 2.5 hours at 30°C, similarly dyed with
20 Nile blue A and observed under a fluorescence microscope. As a result, the surface of the magnetic material (1) did not show fluorescence at all.

Also a part of the synthesized PHA particles was collected by centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G),
25 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The

extract was subjected to an analysis by ^1H -NMR
(equipment: FT-NMR: Bruker DPX400; measured nuclide:
 ^1H ; solvent: deuterionized chloroform (containing TMS)).

Percentages of the side chain units calculated from
such a measurement were 78% for 3-hydroxy-5-
phenoxyvaleric acid and 22% for 3-hydroxy-7,8-
epoxyoctanoic acid.

There were repeated three times operations of
subjecting 50 parts by mass of the aforescribed
magnetic capsule construct (22) to centrifugation
(98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes) to recover
the capsule construct and suspending the same in 50
parts by mass of purified water, and water was
removed by lyophilization. Then 10 parts by mass of
terminal-amino modified polysiloxane (modified
silicone oil TSF4700, GE Toshiba Silicone Co.) were
added and reacted for 2 hours at 70°C. The construct
was rinsed by repeating operations of suspending
methanol and centrifuging (98,000 m/s^2 (= 10,000 G),
4°C, 20 minutes) and dried to obtain a magnetic
capsule construct (23) having a polysiloxane graft
chain.

(Example 20)

Preparation of magnetic capsule construct (24)

To 10 parts by mass of the mcl-PHA synthetase
solution (10 U/ml) derived from pYN2-C1 recombinant
strain, 1 part by mass of the magnetic material (1)

and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (1). The mixture was centrifuged (98,000 m/s² (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s² (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 0.8 part by mass of (R,S)-3-hydroxy-5-phenoxyvaleryl CoA (prepared by hydrolyzing 3-hydroxy-5-phenoxyvaleryl ester, obtained by a Reformatsky reaction of 3-phenoxypropanal and ethyl bromoacetate, to obtain 3-hydroxy-5-phenoxyvaleric acid and then following a process in Eur. J. Biochem., 250, 432-439 (1997)), 0.2 parts by mass of (R)-3-hydroxypimelyl CoA (prepared according to J. Bacteriol., 182, 2753-2760 (2000)), and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C to obtain a magnetic capsule construct (24).

A 10 µl aliquot of the aforescribed magnetic capsule construct (24) were put on a slide glass, to which 10 µl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide

glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (1) was observed to confirm that the magnetic material (1) was coated with PHA on the surface.

Also a part of the magnetic capsule construct (24) was collected by centrifugation (98,000 m/s² (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was subjected to an analysis by ¹H-NMR (equipment: FT-NMR: Bruker DPX400; measured nuclide: ¹H; solvent: deuterionized chloroform (containing TMS)). Percentages of the side chain units calculated from such a measurement were 83% for 3-hydroxy-5-phenoxyvaleric acid and 17% for 3-hydroxypimelic acid. (Example 21).

Preparation of magnetic capsule construct (25)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (1). The mixture was centrifuged

(98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 0.8 part by mass of (R,S)-3-hydroxy-5-phenoxyvaleryl CoA (prepared by hydrolyzing 3-hydroxy-5-phenoxyvaleryl ester, obtained by a Reformatsky reaction of 3-phenoxypropanal and ethyl bromoacetate, to obtain 3-hydroxy-5-phenoxyvaleric acid and then following a process in Eur. J. Biochem., 250, 432-439 (1997)), 0.2 parts by mass of (R)-3-hydroxy-8-bromooctanoyl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)), and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C to obtain a magnetic capsule construct (25).

A 10 μ l aliquot of the aforescribed magnetic capsule construct (25) was put on a slide glass, to which 10 μ l of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of

the magnetic material (1) was observed to confirm that the magnetic material (1) was coated with PHA on the surface.

Also a part of the particles was collected by
5 centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was subjected to an analysis by ^1H -NMR (equipment: FT-NMR:
10 Bruker DPX400; measured nuclide: ^1H ; solvent: deuterionized chloroform (containing TMS)). Percentages of the side chain units calculated from such a measurement were 89% for 3-hydroxy-5-phenoxyvaleric acid and 11% for 3-hydroxy-8-
15 bromooctanoic acid.

(Example 22)

Preparation of magnetic capsule construct (26)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant
20 strain, 1 part by mass of nickel powder of a primary particle size of $0.02 \mu\text{m}$ (Ni(200)UFMP, Shinku Yakin Co.) (magnetic material (2)) as magnetic metal synthesized by a gaseous method, and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at
25 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (2). The mixture was centrifuged ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10

minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s² (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

5 The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of (R)-3-hydroxy-5-phenylvaleryl CoA (prepared by hydrolyzing 3-hydroxyphenylvaleryl ester to obtain 3-hydroxy-5-phenylvaleric acid and then following a process in J. Biochem., 250, 432-439 (1997)) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

15 A 10 µl aliquot of the above reaction solution was put on a slide glass, to which 10 µl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (2) was observed to confirm that the magnetic material (2) was coated with PHA on the surface. This product was taken as a magnetic capsule construct (26).

25 Also a part of the particles was collected by centrifugation (98,000 m/s² (= 10,000 G), 4°C, 10

minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was filtered through a membrane filter of 0.45 μm pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified PHA monomer unit. As a result, there was confirmed PHA constituted of 3-hydroxy-5-phenylvaleric acid unit.

Further, the molecular weight of PHA was evaluated by gel permeation chromatography (GPC: Toso HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 μm), solvent: chloroform, column temperature: 40°C, converted as polystyrene) to obtain a result $M_n = 18,000$ and $M_w = 36,000$.

(Example 23)

20 Preparation of magnetic capsule construct (27)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of $\gamma\text{-Fe}_2\text{O}_3$ fine powder of a primary particle size of 0.02 μm (NanoTek, CI Chemical Co.) synthesized in gaseous method (magnetic material (3)), and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the

mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (3). The mixture was centrifuged ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C , 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C , 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of (R)-3-hydroxy-5-phenylvaleryl CoA (prepared by hydrolyzing 3-hydroxyphenylvaleryl ester to obtain 3-hydroxy-5-phenylvaleric acid and then following a process in Eur. J. Biochem., 250, 432-439 (1997)) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C .

A $10 \mu\text{l}$ aliquot of the above reaction solution was put on a slide glass, to which $10 \mu\text{l}$ of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (3) was observed to confirm that the magnetic material (3) was coated with PHA on the

surface. This product was taken as a magnetic capsule construct (27).

Also a part of the particles was collected by centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was filtered through a membrane filter of 0.45 μm pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified PHA monomer unit. As a result, there was confirmed PHA constituted of 3-hydroxy-5-phenylvaleric acid unit.

Further, the molecular weight of PHA was evaluated by gel permeation chromatography (GPC: Toso HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 μm), solvent: chloroform, column temperature: 40°C, converted as polystyrene) to obtain a result $M_n = 17,000$ and $M_w = 35,000$.

(Example 24)

Preparation of magnetic capsule construct (28)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of magnetite fine powder of a

primary particle size of 0.3 μm (Magnetite EPT500, Toda Kogyo Co.) synthesized by a wet process (magnetic material (4)), and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to
5 cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (4). The mixture was centrifuged ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again ($98,000 \text{ m/s}^2$ (= 10,000
10 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of (R)-3-
15 hydroxy-5-phenylvaleryl CoA (prepared by hydrolyzing 3-hydroxyphenylvaleryl ester to obtain 3-hydroxy-5-phenylvaleric acid and then following a process in Eur. J. Biochem., 250, 432-439 (1997)), and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were
20 added and the mixture was mildly shaken for 2 hours at 30°C.

A 10 μL aliquot of the above reaction solution was put on a slide glass, to which 10 μL of a 1% solution of Nile blue A in water was added. These
25 solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm

long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (4) was observed to confirm that the magnetic material (4) was coated with PHA on the surface. This product was taken as a magnetic capsule construct (28).

Also a part of the particles was collected by centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was filtered through a membrane filter of 0.45 μm pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified PHA monomer unit. As a result, there was confirmed PHA constituted of 3-hydroxy-5-phenylvaleric acid unit.

Further, the molecular weight of PHA was evaluated by gel permeation chromatography (GPC: Toso HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 μm), solvent: chloroform, column temperature: 40°C, converted as polystyrene) to obtain a result $M_n = 15,000$ and $M_w = 34,000$.

(Example 25)

Preparation of magnetic capsule construct (29)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (1). The mixture was centrifuged (98,000 m/s² (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s² (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of (R)-3-hydroxyoctanoyl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)), 1 part by mass of polyethylene glycol 200 (PEG200, Kishida Kagaku Co., average molecular weight 190 to 210) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

A 10 µl aliquot of the above reaction solution was put on a slide glass, to which 10 µl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence

microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (1) was observed to confirm that the magnetic material (1) was coated with PHA on the surface.

As a control, 1 part by mass of the magnetic material (1) was added to 49 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then, after mild shaking for 2.5 hours at 30°C, similarly dyed with Nile blue A and observed under a fluorescence microscope. As a result, the surface of the magnetic material (1) did not show fluorescence at all.

Also a part of the particles was collected by centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was filtered through a membrane filter of 0.45 μm pore size and concentrated under a reduced pressure by using a rotary evaporator. Then the extract was subjected to methanolysis by a conventional method and analyzed by gas chromatography-mass spectroscopy (GC-MS, Shimadzu QP-5050, an EI method) to identify the methyl-esterified PHA monomer unit. As a result, there was confirmed PHA constituted of 3-hydroxyoctanoic acid unit.

Also a detailed structure analysis by ^1H -NMR
(FT-NMR: Bruker DPX400; ^1H resonance frequency: 400
MHz; measured nuclide: ^1H ; solvent: CDCl_3 ; reference:
capillary-sealed TMS/ CDCl_3 ; measuring temperature:
5 room temperature) confirmed, in addition to peaks
derived from PHA constituted of 3-hydroxyoctanoic
acid unit, peaks derived from polyethylene glycol at
3.5 to 3.8 ppm and about 4.2 ppm.

Further, the molecular weight of PHA was
10 evaluated by gel permeation chromatography (GPC: Toso
HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5
 μm), solvent: chloroform, column temperature: 40°C ,
converted as polystyrene) to obtain a result $M_n =$
7,000 and $M_w = 13,000$, and a molecular weight
15 reducing effect was obtained.

(Example 26)

Preparation of laminar construct

A ferrite sheet of 30 mm \times 30 mm \times 3 mm (NP-S01,
Nippon Paint Co., a dispersion of ferrite particles
20 in resin) was immersed for 1 hour in 1%
glutaraldehyde, then rinsed with pure water and
immersed in a solution (10 U/ml) of mcl-PHA
synthetase derived from pYN2-C1 strain for 30 minutes
at 30°C to fix the enzyme. The unreacted mcl-PHA
25 synthetase was removed by rinsing with PBS solution
to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was

immersed in 0.1 mol/L phosphate buffer (pH 7.0),
containing 30 mmol/L of 3-hydroxyoctanoyl CoA
(prepared according to Eur. J. Biochem., 250, 432-439
(1997)), and 0.1% of bovine serum albumin (Sigma Co.)
5 and the mixture was mildly shaken for 2 hours at 30°C.
After the reaction, unreacted substance etc., were
removed by rinsing with 0.1 mol/L phosphate buffer
(pH 7.0).

The ferrite sheet after the reaction was dyed
10 with a 1% aqueous solution of Nile blue A and was
observed under a fluorescence microscope (330 to 380
nm excitation filter, 420 nm long path absorption
filter, Nikon Corp.). As a result, fluorescence from
the surface of the ferrite sheet was observed to
15 confirm that the construct was a laminar construct in
which a base material of the ferrite sheet was
covered by a film of PHA.

Also the laminar construct was dried in vacuum,
and immersed in chloroform under agitation for 20
20 hours at 60°C to extract PHA constituting the coating
layer. The extract was filtered through a membrane
filter of 0.45 μ m pore size and concentrated under a
reduced pressure by using a rotary evaporator. Then
the extract was subjected to methanolysis by a
25 conventional method and analyzed by gas
chromatography-mass spectroscopy (GC-MS, Shimadzu QP-
5050, an EI method) to identify the methyl-esterified

PHA monomer unit. As a result, there was confirmed
PHA constituted of 3-hydroxyoctanoic acid unit as
shown in FIG. 4.

(Example 27)

5 Evaluation of coating property of magnetic
capsule construct

In order to confirm whether the magnetic
particles were completely protected and covered with
the polymer, 0.1 g each of the obtained magnetic
10 capsule constructs (8) to (29) were immersed for 2
hours in 100 ml of purified water heated to 70°C and a
metal content in the water was measured. As a result,
the metal content was 3 ppm or less with all the
capsule constructs. Based on these facts, these
15 capsule constructs were judged that "metal ions did
not elute."

(Example 29)

Preparation of magnetic capsule construct (30)

To 10 parts by mass of the mcl-PHA synthetase
20 solution (10 U/ml) derived from pYN2-C1 recombinant
strain, 1 part by mass of the magnetic material (1)
and 39 parts by mass of PBS were added and mildly
shaken for 30 minutes at 30°C to cause the mcl-PHA
synthetase to be adsorbed on the surface of the
25 magnetic material (1). The mixture was centrifuged
(98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes), and the
precipitate was suspended in PBS solution and

centrifuged again ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C , 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was
5 suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of (R)-3-hydroxy-5-(4-vinylphenyl)valeryl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)), and 0.1 parts by mass of bovine serum albumin (Sigma
10 Co.) were added and the mixture was mildly shaken for 2 hours at 30°C .

After the reaction, a 10 μl aliquot of the above reaction solution was put on a slide glass, to which
10 μl of a 1% solution of Nile blue A in water was
15 added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of
20 the magnetic material (1) was observed to confirm that the magnetic material (1) was coated with PHA on the surface. This product was taken as a magnetic capsule construct (30).

Also a part of the particles was collected by
25 centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C , 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA

constituting the external coating. The extract was subjected to an analysis by ^1H -NMR (equipment: FT-NMR: Bruker DPX400; measured nuclide: ^1H ; solvent: deuterionized chloroform (containing TMS)). As a result, it was confirmed that PHA was constituted of (R)-3-hydroxy-5-(4-vinylphenyl)valeric acid unit.

Further, the molecular weight of PHA was evaluated by gel permeation chromatography (GPC: Tosoh HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 μm), solvent: chloroform, column temperature: 40°C, converted as polystyrene) to obtain a result $M_n = 21,000$ and $M_w = 39,000$.

(Example 30)

Preparation of magnetic capsule construct (31)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of nickel powder of a primary particle size of 0.02 μm (Ni(200)UFMP, Shinku Yakin Co.) (magnetic material (2)) as magnetic metal synthesized by a gaseous method, and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (2). The mixture was centrifuged (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized

magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of (R)-3-hydroxy-5-(4-methylphenoxy)valeryl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)) and 0.1 parts by mass of bovine serum albumin (Sigma-Co.) were added and the mixture was mildly shaken for 10 minutes at 30°C. Then, to this reaction liquid under mild shaking at 30°C, a 0.1 mol/L phosphate buffer (pH 7.0), containing 1 part by mass of (R)-3-hydroxy-5-(4-methylphenyl)valeryl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)) and 0.1 parts by mass of bovine serum albumin (Sigma Co.), was added at a rate of 1 parts by mass per minute by a microtube pump (MP-3N; Tokyo Rikakikai Co.). After 1 hour and 30 minutes, the generated granulates were recovered by centrifugation (98,000 m/s², 4°C, 10 minutes); and, after elimination of the supernatant, 25 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), containing 1 part by mass of (R)-3-hydroxy-5-(4-methylphenyl)valeryl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)) and 0.1 parts by mass of bovine serum albumin (Sigma Co.), were added to the granulates and the mixture was mildly shaken for 20 minutes at 30°C.

After the reaction, a 10 µl aliquot of the above

reaction solution was put on a slide glass, to which
10 μ l of a 1% solution of Nile blue A in water was
added. These solutions were mixed on the slide glass,
covered with a cover glass, and observed under a
5 fluorescence microscope (330 to 380 nm excitation
filter, 420 nm long path absorption filter, Nikon
Corp.). As a result, fluorescence from the surface of
the magnetic material (2) was observed to confirm
that the magnetic material (2) was coated with PHA on
10 the surface. This product was taken as a magnetic
capsule construct (31).

Also the capsule construct was recovered by a
centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10
minutes), and, after drying, the mass of the polymer
15 formed on the surface of the construct was measured
by a time-of-flight secondary ion mass spectrometer
(TOF-SIMS IV, CAMECA). The obtained mass spectrum
confirmed that the surface of the capsule construct
was principally comprised of a homopolymer of
20 polyhydroxy (4-methylphenyl)valeric acid. Also the
measurements of the similar TOF-SIMS mass spectrum
under gradual scraping of the surface of the capsule
construct by ion sputtering showed a copolymer of 3-
hydroxy-5-(4-methylphenyl)valeric acid and 3-hydroxy-
25 5-(4-methylphenoxy)valeric acid, in which the
proportion of 3-hydroxy-5-(4-methylphenyl)valeric
acid gradually decreased while that of 3-hydroxy-5-

(4-methylphenoxy)valeric acid gradually increased toward the interior of the granulate, and which eventually changed to a homopolymer of polyhydroxy (4-methylphenoxy)valeric acid. These results confirmed that the capsule construct of the present example was formed by coating the surface of base material with polyhydroxy(4-methylphenoxy)valeric acid of a high polarity, then with a copolymer of 3-hydroxy-5-(4-methylphenoxy)valeric acid and 3-hydroxy-5-(4-methylphenyl)valeric acid with a gradually increasing proportion of 3-hydroxy-5-(4-methylphenyl)valeric acid toward the outer surface, and finally coating the outermost layer with a homopolymer of polyhydroxy(4-methylphenyl)valeric acid of a low polarity.

Further, the molecular weight of PHA was evaluated by gel permeation chromatography (GPC: Toso HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 μ m), solvent: chloroform, column temperature: 40°C, converted as polystyrene) to obtain a result $M_n = 17,000$ and $M_w = 36,000$.

(Example 31)

Preparation of magnetic capsule construct (32)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of γ -Fe₂O₃ fine powder of a primary particle size of 0.02 μ m (NanoTek, CI

Chemical Co.) synthesized in gaseous method (magnetic material (3)), and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of
5 the magnetic material (3). The mixture was centrifuged (98,000 m/s² (= 10,000 G), 4°C, 10 minutes); and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s² (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized
10 magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 1 part by mass of (R)-3-hydroxy-5-phenylsulfanylvaleryl CoA (prepared
15 according to J. Biochem., 250, 432-439 (1997))) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

After the reaction, a 10-μl aliquot of the above
20 reaction solution was put on a slide glass, to which 10 μl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation
25 filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (3) was observed to confirm

that the magnetic material (3) was coated with PHA on the surface. This product was taken as a magnetic capsule construct (32).

Also a part of the particles was collected by
5 centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was subjected to an analysis by ^1H -NMR (equipment: FT-NMR:
10 Bruker DPX400; measured nuclide: ^1H ; solvent: deuterionized chloroform (containing TMS)). As a result, it was confirmed that PHA was constituted of (R)-3-hydroxy-5-phenylsulfanylvaleric acid unit.

(Example 32)

15 Preparation of magnetic capsule constructs (33) and (34)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of magnetite fine powder of a
20 primary particle size of $0.3 \mu\text{m}$ (Magnetite EPT500, Toda Kogyo Co.) synthesized by a wet process (magnetic material (4)), and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the
25 surface of the magnetic material (4). The mixture was centrifuged ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS

solution and centrifuged again ($98,000 \text{ m/s}^2$ ($= 10,000 \text{ G}$), 4°C , 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was
5 suspended in 48 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then 0.8 parts by mass of (R)-3-hydroxy-5-phenylvaleryl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)), 0.2 parts by mass of (R)-3-hydroxy-5-(4-vinylphenyl)valeryl CoA
10 (prepared according to Eur. J. Biochem., 250, 432-439 (1997)), and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C to obtain a sample 1.

As a comparative reference, a sample 2 was
15 obtained in the same process as explained above except that (R)-3-hydroxy-5-(4-vinylphenyl)valeryl CoA was replaced by (R)-3-hydroxy-5-(4-methylphenyl)valeryl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)).

20 A 10 μl aliquot of each sample was put on a slide glass, to which 10 μl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380
25 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (4) was observed

to confirm that the magnetic material (4) was coated with PHA on the surface. These products were taken as a magnetic capsule constructs (33) and (34).

As a control, 1 part by mass of the magnetic material (4) was added to 49 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then, after mild shaking for 2.5 hours at 30°C, similarly dyed with Nile blue A and observed under a fluorescence microscope. As a result, the surface of the magnetic material (4) did not show fluorescence at all.

Also a part of the particles was collected by centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was subjected to an analysis by ^1H -NMR (equipment: FT-NMR: Bruker DPX400; measured nuclide: ^1H ; solvent: deuterionized chloroform (containing TMS)). Percentages of the side chain units calculated from the measured results are shown in Table 4.

Table 4

Composition of PHA of external coating of capsule construct ($^1\text{H-NMR}$, unit %)

Monomer Unit	Magnetic Capsule Construct 33	Magnetic Capsule Construct 34
3-hydroxy-5-valeric Acid	83%	85%
3-hydroxy-5-(4-vinylphenyl) Valeric Acid	17%	-
3-hydroxy-5-(4-methylphenyl) Valeric Acid	-	15%

5 (Example 33)

Preparation of capsule construct 35

An epoxylating reaction was conducting on the magnetic capsule construct prepared in example 32. 1 part by mass of the magnetic capsule construct 4 was charged in a four-necked flask, and was agitated with 6 parts by mass of distilled water. The interior of the flask was heated to 40°C, and 1 part by mass of a 30 % hexane solution of peracetic acid was dropwise added continuously, and was reacted for 5 hours under agitation at 40°C. The reaction proceeded without mutual coagulation of the particles of the magnetic capsule construct. After the reaction, the reaction liquid was cooled to the room temperature and filtered to recover the magnetic capsule construct.

20 The recovered magnetic capsule construct was re-suspended in distilled water, and centrifuged (29,400

m/s² (= 3,000 G), 4°C, 30 minutes). After the separation, the magnetic capsule construct was rinsed by suspending in distilled water again and centrifuging again. This rinsing operation was
5 further repeated three times. Thereafter drying under vacuum was conducted to obtain a desired magnetic capsule construct (35).

Also a part of the particles was collected by centrifugation (98,000 m/s² (= 10,000 G), 4°C, 10
10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was subjected to an analysis by ¹H-NMR (equipment: FT-NMR: Bruker DPX400; measured nuclide: ¹H; solvent:
15 deuterionized chloroform (containing TMS)). Percentages of the side chain units calculated from the measured results were 85% for 3-hydroxy-5-phenylvaleric acid, 11% for 3-hydroxy-5-(4-vinylphenyl)valeric acid and 4% for 3-hydroxy-5-(4-
20 epoxyphenyl)valeric acid.

Since this reaction system was an inhomogeneous reaction, it is estimated that the surface of the capsule construct was epoxylated while the interior of the coating layer remained unreacted.

25 (Example 34)

Crosslinking reaction of capsule construct

There were repeated three times operations of

subjecting 50 parts by mass of the magnetic capsule construct (35) prepared in the example 33 to centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C , 10 minutes) to recover the capsule construct and
5 suspending the same in 50 parts by mass of purified water, and 0.5 parts by mass of hexamethylene diamine were dissolved in the suspension. After dissolution was confirmed, water was removed by lyophilization (thus providing a magnetic capsule construct (36)).
10 Then the magnetic capsule construct (36) was reacted for 12 hours at 70°C (thus providing a magnetic capsule construct (37)).

Each of the magnetic capsule constructs (36) and (37) was suspended in chloroform, and agitated for 20
15 hours at 60°C to extract PHA constituting the outer coating, which, after elimination of chloroform by drying under vacuum, was subjected to a differential scanning calorimeter (DSC; Perkin Elmer Inc., Pyris 1, temperature increase rate: 10°C/min). As a result,
20 the magnetic capsule construct (36) showed a clear exothermic peak at about 90°C , indicating that a reaction between an epoxy group in the polymer and hexamethylene diamine took place and the crosslinking between polymers was proceeding. On the other hand,
25 the magnetic capsule construct (37) did not show a clear heat flow, indicating that the crosslinking reaction was almost completed.

Also infrared absorption was measured on similar samples (FT-IR; Perkin Elmer, 1720X). As a result, peaks for amine (around $3,340\text{ cm}^{-1}$) and epoxy (around 822 cm^{-1}) observed in the magnetic capsule construct (36) disappeared in the magnetic capsule construct (37).

These results indicate that a crosslinked polymer can be obtained by reacting PHA having an epoxy unit in the side chain and hexamethylene diamine.

On the other hand, a similar evaluation was conducted on the magnetic capsule construct (34) as a comparative reference, but a result clearly showing the mutual crosslinking of polymers as explained before was not obtained.

(Example 35)

Grafting of capsule construct

There were repeated three times operations of subjecting 50 parts by mass of the aforescribed magnetic capsule construct (35) prepared in the example 33 to centrifugation ($98,000\text{ m/s}^2$ ($= 10,000\text{ G}$), 4°C , 10 minutes) to recover the capsule construct and suspending the same in 50 parts by mass of purified water, and water was removed by lyophilization. Then 10 parts by mass of terminal-amino modified polysiloxane (modified silicone oil TSF4700, GE Toshiba Silicone Co.) were added and reacted for 2

hours at 70°C. The construct was rinsed by repeating operations of suspending methanol and centrifuging (98,000 m/s² (= 10,000 G), 4°C, 20 minutes) and dried to obtain a magnetic capsule construct (38) having a polysiloxane graft chain.

(Example 36)

Preparation of capsule construct

An oxidation cleaving reaction of the vinyl group was conducted on the magnetic capsule construct (33) prepared in the example 32. 10 parts by mass of the magnetic capsule construct were transferred to a three-necked flask, and 300 parts by mass of distilled water containing 50 ppm of hydrogen peroxide were added. Agitation was conducted for 3 hours at the room temperature, under blowing of ozone at a rate of 1 part by mass per hour. The reaction proceeded without coagulation of the particles of the magnetic capsule construct. After the reaction, the reaction liquid was filtered to recover the magnetic capsule construct. The magnetic capsule construct was suspended again in distilled water and was centrifuged (29,400 m/s² (= 3,000 G), 4°C, 30 minutes) to wash off the remaining hydrogen peroxide-containing water. This rinsing operation was further repeated twice. Thereafter drying under vacuum was conducted to obtain a magnetic capsule construct (39).

Also a part of the particles was collected by

centrifugation ($98,000 \text{ m/s}^2$ ($= 10,000 \text{ G}$), 4°C , 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was
5 subjected to an analysis by ^1H -NMR (equipment: FT-NMR: Bruker DPX400; measured nuclide: ^1H ; solvent: deuterionized chloroform (containing TMS)). Percentages of the side chain units calculated from the measured results were 84% for 3-hydroxy-5-
10 phenylvaleric acid, 11% for 3-hydroxy-5-(4-vinylphenyl)valeric acid and 5% for 3-hydroxy-5-(4-carboxyphenyl)valeric acid.

Since this reaction system was an inhomogeneous reaction, it was estimated that the surface of the
15 capsule construct was subjected to oxidation cleaving while the interior of the coating layer remained unreacted.

(Example 37)

Preparation of capsule construct (40)

20 1 part by mass of the magnetic capsule construct (32) prepared in the example 31 was added to 150 parts by mass of a commercially available aqueous solution of hydrogen peroxide (Mitsubishi Gas Chemical Co., containing 31% of hydrogen peroxide,
25 according to standard JIS K-8230) and 30 parts by mass of deionized water, and the mixture was transferred to an eggplant-shaped flask and was

reacted on an oil bath for 1 hour at 100°C. After the reaction, it was cooled to the room temperature, and the magnetic capsule construct was separated by centrifugation ($29,400 \text{ m/s}^2$ (= 3,000 G), 4°C, 10 minutes). The magnetic capsule construct was suspended again in distilled water and was centrifuged again to wash off the remaining hydrogen peroxide-containing water. This rinsing operation was further repeated twice. Thereafter drying under vacuum was conducted to obtain a magnetic capsule construct (40).

Also a part of the particles was collected by centrifugation ($98,000 \text{ m/s}^2$ (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was subjected to an analysis by ^1H -NMR (equipment: FT-NMR: Bruker DPX400; measured nuclide: ^1H ; solvent: deuterionized chloroform (containing TMS)). Percentages of the side chain units calculated from the measured results were 64% for 3-hydroxy-5-phenylsulfanylvaleric acid, 16% for 3-hydroxy-5-phenylsufinylvaleric acid and 20% for 3-hydroxy-5-phenylsulfonylvaleric acid.

Since this reaction system was an inhomogeneous reaction, it was estimated that the surface of the capsule construct was subjected to oxidation while

the interior of the coating layer remained unreacted.

(Example 38)

Preparation of capsule construct (41)

1 part by mass of the magnetic capsule construct
5 (32) prepared in the example 31 was added to 90 parts
by mass of a commercially available aqueous solution
of hydrogen peroxide (Mitsubishi Gas Chemical Co.,
containing 31% of hydrogen peroxide, according to
standard JIS K-8230) and 30 parts by mass of
10 deionized water, and the mixture was transferred to
an eggplant-shaped flask and was reacted on an oil
bath for 1 hour at 100°C. After the reaction, it was
cooled to the room temperature, and the magnetic
capsule construct was separated by centrifugation
15 (29,400 m/s² (= 3,000 G), 4°C, 10 minutes). The
magnetic capsule construct was suspended again in
distilled water and was centrifuged again to wash off
the remaining hydrogen peroxide-containing water.
This rinsing operation was further repeated twice.
20 Thereafter drying under vacuum was conducted to
obtain a magnetic capsule construct (41).

Also a part of the particles was collected by
centrifugation (98,000 m/s² (= 10,000 G), 4°C, 10
minutes), dried in vacuum, suspended in chloroform
25 and agitated for 20 hours at 60°C to extract PHA
constituting the external coating. The extract was
subjected to an analysis by ¹H-NMR (equipment: FT-NMR:

Bruker DPX400; measured nuclide: ^1H ; solvent: deuterionized chloroform (containing TMS)).

Percentages of the side chain units calculated from the measured results were 61% for 3-hydroxy-5-phenylsulfanylvaleric acid, 31% for 3-hydroxy-5-phenylsufinylvaleric acid and 8% for 3-hydroxy-5-phenylsulfonylvaleric acid.

Since this reaction system was an inhomogeneous reaction, it was estimated that the surface of the capsule construct was subjected to oxidation while the interior of the coating layer remained unreacted.

(Example 39)

Preparation of magnetic capsule construct (42)

To 10 parts by mass of the mcl-PHA synthetase solution (10 U/ml) derived from pYN2-C1 recombinant strain, 1 part by mass of the magnetic material (1) and 39 parts by mass of PBS were added and mildly shaken for 30 minutes at 30°C to cause the mcl-PHA synthetase to be adsorbed on the surface of the magnetic material (1). The mixture was centrifuged (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes), and the precipitate was suspended in PBS solution and centrifuged again (98,000 m/s^2 (= 10,000 G), 4°C, 10 minutes) to obtain enzyme-immobilized magnetic material.

The aforescribed immobilized enzyme was suspended in 48 parts by mass of 0.1 mol/L phosphate

buffer (pH 7.0), then 1 part by mass of (R)-3-hydroxy-5-(4-vinylphenyl)valeryl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)), 1 part by mass of polyethylene glycol 200 (PEG200, Kishida Kagaku Co., average molecular weight 190 to 210) and 0.1 parts by mass of bovine serum albumin (Sigma Co.) were added and the mixture was mildly shaken for 2 hours at 30°C.

A 10 µl aliquot of the above reaction solution was put on a slide glass, to which 10 µl of a 1% solution of Nile blue A in water was added. These solutions were mixed on the slide glass, covered with a cover glass, and observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the magnetic material (1) was observed to confirm that the magnetic material (1) was coated with PHA on the surface.

As a control, 1 part by mass of the magnetic material (1) was added to 49 parts by mass of 0.1 mol/L phosphate buffer (pH 7.0), then, after mild shaking for 2.5 hours at 30°C, similarly dyed with Nile blue A and observed under a fluorescence microscope. As a result, the surface of the magnetic material (1) did not show fluorescence at all.

Also a part of the particles was collected by

centrifugation (98,000 m/s² (= 10,000 G), 4°C, 10 minutes), dried in vacuum, suspended in chloroform and agitated for 20 hours at 60°C to extract PHA constituting the external coating. The extract was
5 subjected to an analysis by ¹H-NMR (equipment: FT-NMR: Bruker DPX400; measured nuclide: ¹H; solvent: deuterionized chloroform (containing TMS)). As a result, PHA was confirmed to be constituted of (R)-3-hydroxy-5-(4-vinylphenyl)valeric acid unit. Also
10 there were confirmed, in addition to peaks derived from (R)-3-hydroxy-5-(4-vinylphenyl)valeric acid unit, peaks derived from PEG at 3.5 to 3.8 ppm and at about 4.2 ppm.

Further, the molecular weight of PHA was
15 evaluated by gel permeation chromatography (GPC: Toso HLC-8020, column: Polymer Laboratory PLgel MIXED-C (5 µm), solvent: chloroform, column temperature: 40°C, converted as polystyrene) to obtain a result Mn = 7,700 and Mw = 14,000, and a molecular weight
20 reducing effect was obtained.

(Example 40)

Preparation of laminar construct

A ferrite sheet of 30 mm × 30 mm × 3 mm (NP-S01, Nippon Paint Co., a dispersion of ferrite particles
25 in resin) was immersed for 1 hour in 1% glutaraldehyde, then rinsed with purified water and immersed in a solution (10 U/ml) of PHA synthetase

derived from pYN2-C1 strain for 30 minutes at 30°C to fix the enzyme. The unreacted PHA synthetase was removed by rinsing with PBS solution to obtain enzyme-immobilized magnetic material.

5 The aforescribed immobilized enzyme was immersed in 0.1 mol/L phosphate buffer (pH 7.0), containing 30 mmol/L of (R)-3-hydroxy-5-(4-methylthio(phenoxy))valeryl CoA (prepared according to Eur. J. Biochem., 250, 432-439 (1997)), and 0.1%
10 of bovine serum albumin (Sigma Co.) and the mixture was mildly shaken for 2 hours at 30°C. After the reaction, unreacted substance etc. were removed by rinsing with 0.1 mol/L phosphate buffer (pH 7.0).

 The sheet after the reaction was dyed with a 1%
15 aqueous solution of Nile blue A and was observed under a fluorescence microscope (330 to 380 nm excitation filter, 420 nm long path absorption filter, Nikon Corp.). As a result, fluorescence from the surface of the sheet was observed to confirm that the
20 construct was a laminar construct in which a base material of the ferrite sheet was covered by a film of PHA.

 Also the laminar construct was dried in vacuum, and immersed in chloroform under agitation for 20
25 hours at 60°C to extract PHA constituting the coating layer. The extract was subjected to an analysis by ¹H-NMR (equipment: FT-NMR: Bruker DPX400; measured

nuclide: ^1H ; solvent: deuterionized chloroform (containing TMS). As a result, there was confirmed PHA constituted of 3-hydroxy-5-(4-methylthio(phenoxy)valeric acid unit.

5 (Example 41)

Evaluation of coating property of magnetic capsule construct

In order to confirm whether the magnetic particles were completely protected and covered with the polymer, 0.1 g each of the obtained magnetic capsule constructs (30) to (42) were immersed for 2 hours in 100 ml of pure water heated to 70°C and a metal content in the water was measured. As a result, the metal content was 3 ppm or less with all the capsule constructs. Based on these facts, these capsule constructs were judged that metal ions did not elute.

(Example 43)

20 α -fetoprotein (AFP) immunoassay by anti-AFP antibody carried on magnetic capsule construct (18)

1. Immobilization of antibody on epoxylated PHA magnetic capsule construct

The epoxylated magnetic capsule construct prepared in the example (18) was uniformly dispersed in a phosphate buffer (0.1 M; pH 7.4) (theoretical concentration 5 to 10×10^8 particle/mL), and an anti-AFP antibody dissolved in a similar phosphate buffer

was added in such a manner that the final ratio of the antibody to the magnetic capsule construct (18) became 5-10 $\mu\text{g}/10^7$, and the mixture was mildly agitated by pipetting.

5 After a reaction for 1 hour at 30°C by a rotary shaker, bovine serum albumin (BSA) for blocking was so added as to reach a final concentration of 0.3%, and a reaction was carried out for further 15 hours to obtain the antibody immobilized on the surface of
10 the magnetic capsule construct (18).

2. Reaction with target component (antigen: AFP)

In a 5 mL Eppendorf tube (precoated with BSA), 500 μL of the dispersion of the anti-AFP-immobilized magnetic capsule construct (18), prepared in step 1,
15 was mixed with 20 μL of an AFP solution of a concentration of 1 $\mu\text{g}/\text{mL}$ and was reacted for 30 minutes at 37°C. The tube was brought into contact with a magnet to collect the magnetic capsule construct, and the supernatant was removed by
20 decantation. Thereafter, 2 mL of a 0.04% NaCl solution were added and agitated in the tube. Then the magnetic capsule construct was collected by the magnetic force and the supernatant was removed by decantation in the same manner as explained above.
25 The similar rinsing operation was repeated three times.

3. Reaction with enzyme-labeled secondary

antibody

An alkali phosphatase-labeled AFP antibody Fab' was prepared according to J. Immunoassay, 4, 209 (1983) and Biochemistry, 11(12), 2291 (1972), and was mixed and dissolved in 0.1 M trishydrochloric acid buffer (containing 2% BSA, 1 mM $MgCl_2$ and 0.1 mM $ZnCl_2$; pH 7.5), and 500 μL of the mixture were added to the antigen AFP-binding capsule construct, prepared in step 2, and reacted for 10 minutes at 37°C. Then the AFP-binding magnetic capsule construct reacted with the enzyme-labeled secondary antibody was rinsed in a similar manner as in step 2.

4. Detection

Into the tube containing the AFP-binding magnetic capsule construct reacted with the enzyme-labeled secondary antibody, 500 μL of glycine-NaOH buffer (0.1 M; pH 10.3; containing $MgCl_2$ (1mM) and egg white albumin (250 mg/L) were added and, after a reaction for 5 minutes at 37°C, 500 μL of the aforescribed buffer containing 4-nitrophenylphosphoric acid (final concentration 5.5 mM) were added and reacted for 60 minutes at 37°C. After the reaction, 500 μL of a 1 M NaOH aqueous solution were added to terminate the reaction, and absorbances in ultraviolet and visible regions were measured. As a result, there was detected absorbance at 405 nm, resulting from a reaction product by the

labeled enzyme alkali phosphatase, thus confirming the recovery of the AFP.

(Example 44)

Recovery of Concanavalin A utilizing
5 cellopentaose-immobilized aminated magnetic capsule construct (35)

1. Amination of magnetic capsule construct (35)

The magnetic capsule construct (35) prepared in example 33 was subjected to amination by adding 2,2'-
10 (ethylenedioxy)-dimethylamine followed by a reaction for 15 hours at 30°C. After the reaction, it was rinsed five times with bis-2-methoxyethyl ether to eliminate remaining amine, and further rinsed with distilled water three times to obtain aminated
15 magnetic capsule construct (35').

2. Immobilization of cellopentaose on magnetic capsule construct (35')

The magnetic capsule construct (35') obtained in the step 1 was uniformly dispersed in a phosphate
20 buffer as in example 1 and was agitated, for 30 hours at 30°C, with a product of oxidation in advance of a non-reducing terminal of D-(+)-cellopentase (Sigma Co.) with sodium periodate to generate an aldehyde structure (-CHO: formyl group), thereby immobilizing
25 cellopentaose on the magnetic capsule construct (35'). Thereafter, butanedienic anhydride was added and reacted to block excessive amino groups remaining on

the surface of the magnetic capsule construct, and rinsing with distilled water was conducted three times to obtain a cellopentaose-immobilizing magnetic capsule construct (35').

5 3. Recovery of Concanavalin A

Concanavalin A (Sigma Co.) and BSA were dissolved in the aforescribed phosphate buffer, and the cellopentaose-immobilizing magnetic capsule construct (35') obtained in the step 2 was added and
10 reacted for 15 hours at 30°C, and the magnetic capsule construct was recovered by a magnetic force. The recovered particles were processed with sodium dodecylsulfate (SDS) and the elute was subjected to an SDS-PAGE analysis, which showed a substantially
15 single band at 104 kDa, indicating the recovery of Concanavalin A.

(Example 45)

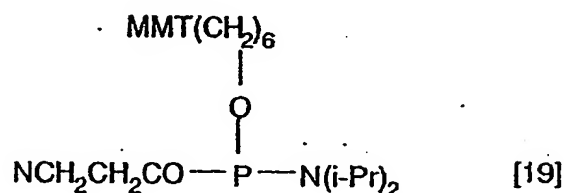
Screening of DNA fragment with magnetic capsule construct (39)

20 1. Synthesis and amination of model nucleic acid M13p18ssDNA

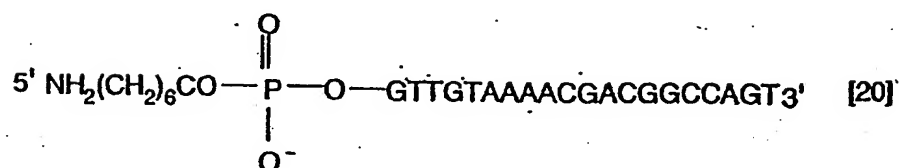
A 20-mer oligonucleotide having a complimentary base sequence to a single-chain DNA of *E. coli* M13 phage mp18 and represented by a SEQ ID NO: 18 was
25 synthesized by an auto DNA synthesizer (381A; ABI Inc.):

SEQ ID NO: 18: 5'-GTTGTAAAACGACGGCCAGT-3'

Then, an amino group ($-\text{NH}_2$) was introduced at 5' of the aforedescribed oligonucleotide 20-mer (compound "20"), utilizing a deoxyuridylate derivative monomer having an amino group (compound "19") instead of an ordinary amidide reagent. Then there were carried out, according to ordinary methods, a cleavage from the CPG support, a deprotection and a purification with a high speed performane liquid chromatography (HPLC).



10



2. Immobilization of aminated probe
oligonucleotide on magnetic capsule construct (39)

The carboxylated magnetic capsule construct (39)

obtained in the example 36 was rinsed in advance with 0.01 M sodium hydroxide solution, and the obtained particles were reacted with EDC (1-ethyl-3-(3-diethylaminopropyl)-carbodiimide hydrochlorate) and aminated oligonucleotide of a theoretical amount of about double of the carboxy group for 18 hours at 4°C, thereby obtaining a magnetic capsule construct carrying the probe oligonucleotide.

3. Synthesis and labeling of target model oligonucleotide

With respect to the oligonucleotide of the aforescribed SEQ ID NO: 18, a completely complementary 20-mer oligonucleotide (SEQ ID NO: 19), an oligonucleotide with mismatching in one base (SEQ ID NO: 20) and an oligonucleotide with mismatching in two bases (SEQ ID NO: 21) were respectively synthesized by an automatic synthesizing equipment and purified by the ordinary method:

SEQUENCE ID NO. 19: 5'-ACTGGCCGTCGTTTACAAC-3'

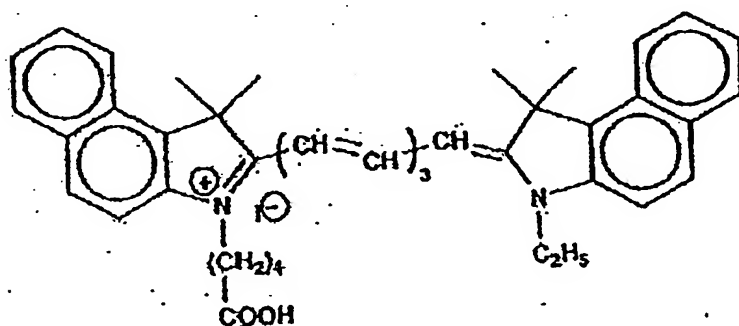
SEQUENCE ID NO. 20: 5'-ACTGGCCGTCCTTTACAAC-3'

SEQUENCE ID NO. 21: 5'-ACTGGCGGTCGTTATACAAC-3'.

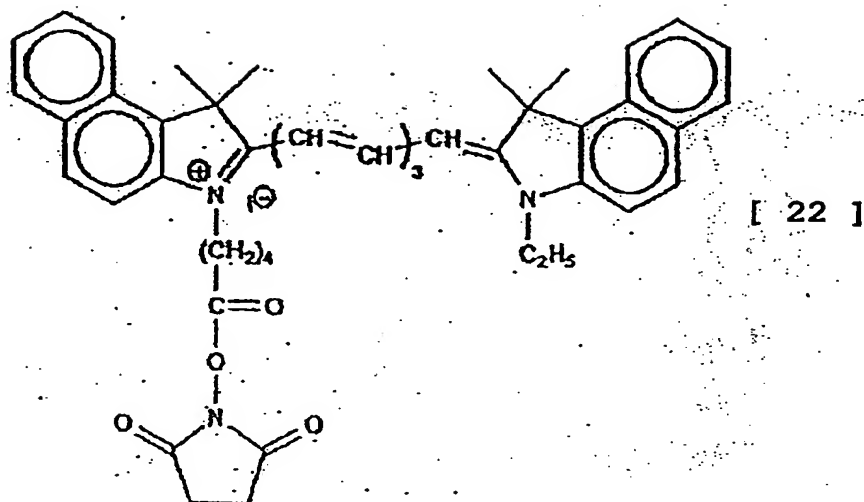
Each of the three purified model oligonucleotides was subjected to amination of the 5' end, utilizing a deoxyuridylate derivative monomer (compound "19") as in the step 1.

Separately, 170 mg of a cyanine dye (compound "21") were dissolved in 5 ml of dry DMF, and 50 µl of

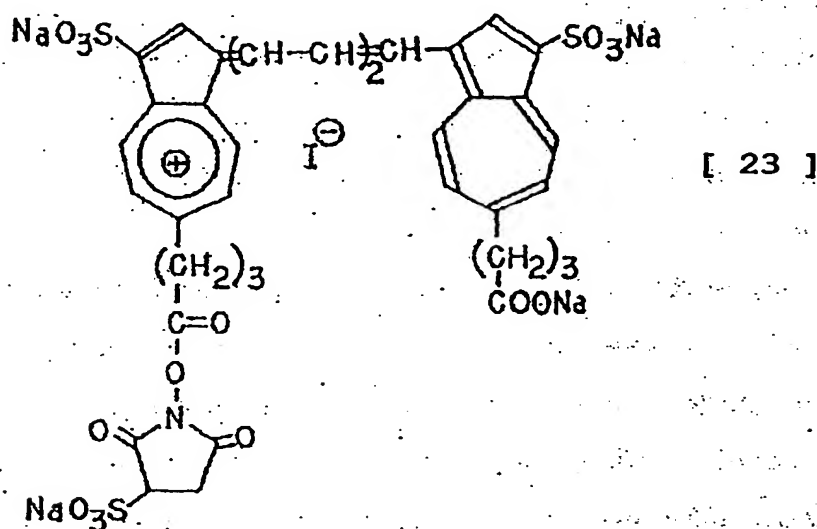
dry pyridine was added according to a method disclosed in Japanese Patent No. 03368011. Then 128 mg of DSC (disuccimidyl carbonate) were added, and agitation was conducted for 20 hours in a dark place at the room temperature. The mixture was added with 150 ml of diethyl ether, and the precipitate was collected, rinsed with diethyl ether and dried. The obtained active ester (compound "22") was used for labeling the target model oligonucleotide (completely complementary strand: SEQ ID NO: 19).

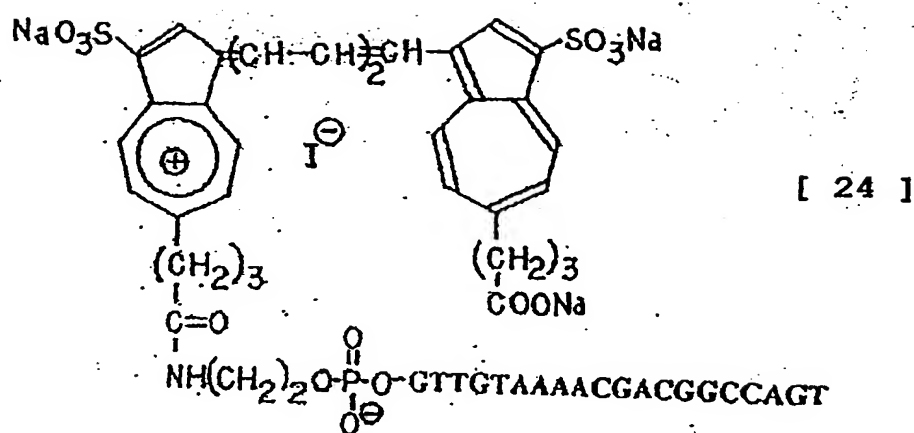


[21]



Also a compound [23], which was an active ester
 of an azulene dye, was used for labeling amino
 5 compounds of the aforescribed oligonucleotides of
 mismatching in one base (SEQ ID NO: 20) and in two
 bases (SEQ ID NO: 21) (compound "24").





4. Screening of completely complementary oligonucleotide

5 The magnetic capsule construct (39) having the probe oligonucleotide obtained in the step 2 and the labeled target oligonucleotide obtained in the step 3 were so regulated as to obtain a theoretical probe/target ratio of 1/10, then maintained for 2

10 minutes at 80°C and returned to the room temperature (hybridization condition).

 The magnetic capsule construct was separated magnetically, and was subjected to a fluorescence measurement under an excitation with a laser of 780

15 nm. As a result, fluorescence was observed with a peak at about 820 nm, resulting from the compound of

the formula [21], thus confirming that the target DNA of the SEQ ID NO: 15 was selectively recovered.

(Example 46)

Recovery of nonylphenol utilizing magnetic capsule construct (39) carrying antialkylphenol antibody

1. Immobilization of anti-alkylphenol antibody on magnetic capsule construct (39)

A carboxy group on the PHA side chain of the carboxy-type magnetic capsule construct (39) prepared in the example 36 was activated with N-ethyl-N'-(dimethylaminopropyl)carbodiimide (EDC) and N-hydroxysuccinimide (NHS) by the ordinary method, and reacted with an anti-alkylphenol antibody (Wako Pure Chemical Co.) of a theoretical twice amount, thereby obtaining a magnetic capsule construct (39) carrying the anti-alkylphenol antibody.

2. Recovery and detection of nonylphenol

According to the method of "alkylphenol ELISA kit" (Wako Pure Chemical Co.), a standard liquid of nonylphenol and the magnetic capsule construct carrying the anti-alkylphenol antibody obtained in the step 1 were mixed and the reaction was accelerated by a switching operation of an electromagnet adhered to an external side of a container. After this operation was conducted for 30 minutes at 30°C, the electromagnet was maintained

turned on to capture the magnetic capsule construct on the internal wall of the container, and the reaction liquid was removed. Then a rinsing liquid was added to the container, and the rinsing was accelerated by a switching operation of the electromagnet. After repeating the rinsing step of 5 minutes with new rinsing liquid three times, a solution of a chromophore-carrying substrate was added and an absorbance was measured. As a result, a strong absorption was observed at 450 nm, thus indicating that nonylphenol could be recovered and detected by this method.

(Example 47)

Separation and recovery of liposome utilizing magnetic capsule construct (25) carrying a long-chain alkane

1. Immobilization of dodecane on magnetic capsule construct (25), utilizing dodecanethiol

1-dodecanethiol was dissolved in hexane, then sodium iodide, potassium carbonate and diethyl amine were added, and mixed with the bromo-type magnetic capsule construct (25) prepared in the example 21 under agitation for 20 hours at the room temperature to obtain a magnetic capsule construct carrying dodecane through a sulfide bond (-S-).

2. Recovery and detection of fluorescent liposome

Liposome including FITC inside was prepared utilizing a "Liposome Kit" manufactured by Sigma Co. and according to a protocol thereof, and was mixed under agitation with the dodecane-carrying magnetic capsule construct obtained in the step 1. After mixing for 15 minutes, a probe-shaped electromagnet was charged in a container, and was turned on to capture the particles on the probe-shaped electromagnet. Then the magnetic capsule construct was transferred to a container containing a rinsing liquid, then released therein by turning off the electromagnet and was agitated for 15 minutes. After repeating this rinsing operation three times, a fluorescence measurement was conducted. As a result, a strong fluorescence was observed at 520 nm. resulting from FITC, thus indicating that nonylphenol could be recovered and detected by this method.

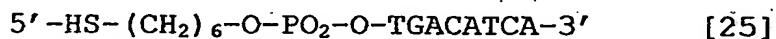
(Example 48)

Separation and recovery of transfer protein utilizing magnetic capsule construct (39) carrying consensus binding sequence gene fragment

Utilizing an affinity of a Transcription factor ATF-2 having a basic leucine zipper and a consensus binding sequence thereof, separation and recovery of ATF-2 was carried out.

1. Synthesis of fragment of terminal thiol-type ATF-2 consensus binding sequence

An automatic DNA synthesizer was employed to synthesize a single-stranded nucleic acid (TGACATCA, SEQ ID NO: 22). Also at a 5' terminal of the ssDNA of the SEQ ID NO: 18, a sulfanyl group (-SH) was introduced (compound "25") during the synthesis using the automatic DNA synthesizer with a Thio-Modifier (Glenn Research Inc.). Then DNA was recovered by an ordinary deprotection, purified by the ordinary method and used in the following experiments.



2. Immobilization of terminal thiol DNA fragment on magnetic capsule construct

A carboxy group on the PHA side chain of the carboxy-type magnetic capsule construct (39) prepared in the reference example 8 was activated with N-ethyl-N'-(dimethylaminopropyl)carbodiimide (EDC) and N-hydroxysuccinimide (NHS) by an ordinary method, then activated to thiol by 2-(2-pyridinyldithio)ethanamine (PDEA) dissolved in a borate buffer (pH 8.5; regulated with NaOH) and was subjected to an immobilization operation by adding the terminal thiol DNA fragment. After the reaction, the active group remaining on the magnetic capsule construct was deactivated with cysteine-NaCl.

3. Binding reaction and labeling

According to a protocol of Clontec Inc., the DNA

fragment-carrying magnetic capsule construct obtained
in the step 2 was introduced into a PBS solution of
AFT-2, and the binding reaction based on the affinity
of DNA fragment and AFT-2 was accelerated by a magnet
5 in a similar manner as in the example 46. After the
reaction, rinsing was conducted in a similar manner
as in the example 46, and an antigen-antibody
reaction was conducted by an ordinary method in a
solution of anti-AFT-2 antibody (polyclonal) labeled
10 with FITC.

4. Rinsing and detection

After the antigen-antibody reaction, rinsing
with a magnet and fluorescence detection were carried
out in the same manner as in the example 46. As a
15 result, there was confirmed a fluorescence having a
maximum at 520 nm due to the label FITC, thus
confirming recovery and detection based on the
affinity binding of the transfer protein and
consensus sequence DNA fragment.

20

INDUSTRIAL APPLICABILITY OF THE INVENTION

The construct of the invention, showing
excellent dispersibility of magnetic material and
magnetic response, low elution of metal ions to the
25 exterior and excellent biological compatibility, can
be widely applicable to various uses and fields.

Also according to the invention, a capsule

construct or a laminar construct, in which a magnetic material is coated, can be produced with an extremely simple process of a low environmental burden.

In particular, it is rendered possible to
5 produce a construct which coats a magnetic material with excellent dispersibility, without an oleophilic treatment on a metal or a metal oxide having magneticity, and a manufacture thereof.

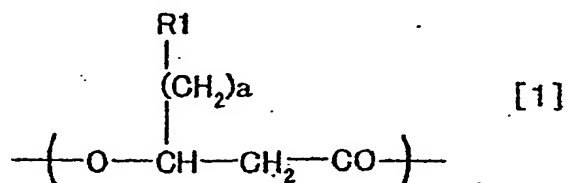
Also the capsule construct coating the magnetic
10 material, obtained by the invention, can efficiently fix a target component-binding molecule by utilizing a side chain of polyhydroxyalkanoate constituting the coating polymer, also shows an excellent dispersibility, and can achieve efficient separation,
15 recovery and detection of a target component in a specimen under a condition close to in vivo, because of the use of polyhydroxyalkanoate of a high biological affinity as the polymer material coating the surface.

CLAIMS

1. A construct characterized in that at least a part of a magnetic material is coated with polyhydroxyalkanoate.

2. A construct according to claim 1, wherein the polyhydroxyalkanoate includes a 3-hydroxypropionic acid unit or a 3-hydroxyalkanoic acid unit.

3. A construct according to claim 1, wherein the polyhydroxyalkanoate includes at least one selected from a group consisting of monomer units represented by chemical formulas [1] to [10] and [A] to [D]:



(wherein the monomer unit is at least one selected from a group of monomer units having following combinations of R1 and a:

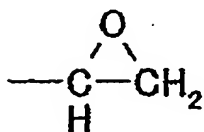
a monomer unit in which R1 is a hydrogen atom (H) and a is any of integers from 3 to 10;

a monomer unit in which R1 is a halogen atom and a is any of integers from 1 to 10;

a monomer unit in which R1 is a chromophore and a is any of integers from 1 to 10;

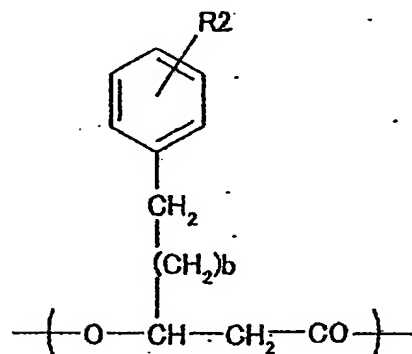
a monomer unit in which R1 is a carboxyl group or a salt thereof and a is any of integers from 1 to 10; and

a monomer unit in which R1 is



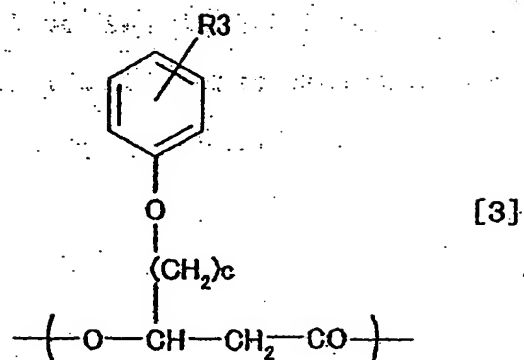
5

and a is any of integers from 1 to 7.);

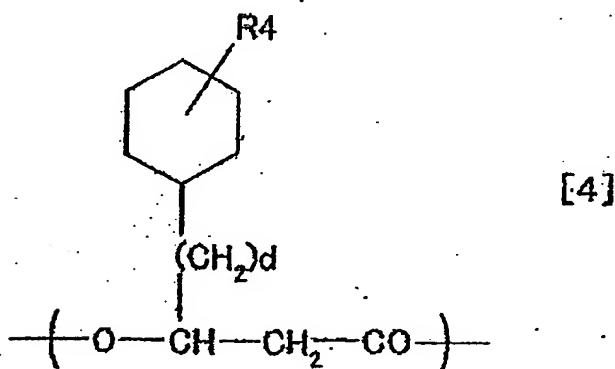


[2]

- 10 (wherein b represents any of integers from 0 to 7; R2 represents any one selected from a group of a hydrogen atom (H), a halogen atom, -CN, -NO₂, -CF₃, -C₂F₅, -C₃F₇, a CH₃ group, a C₂H₅ group, a C₃H₇ group, a vinyl group, an epoxy group and COOR₂₁ (R₂₁ representing an H atom, an Na atom or a K atom); and,
- 15 in the presence of plural units, the foregoing stands independently for each unit.);

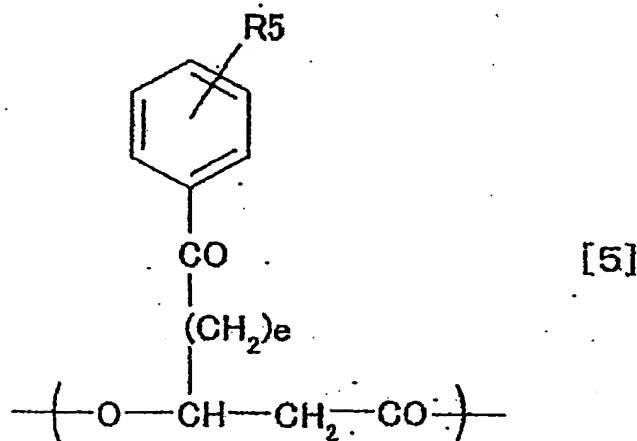


- (wherein c represents any of integers from 1 to 8 and R3 represents any one selected from a group of a hydrogen atom (H), a halogen atom, -CN, -NO₂, -CF₃, -C₂F₅, -C₃F₇, a CH₃ group, a C₂H₅ group, a C₃H₇ group and a SCH₃ group; and, in the presence of plural units, the foregoing stands independently for each unit.);

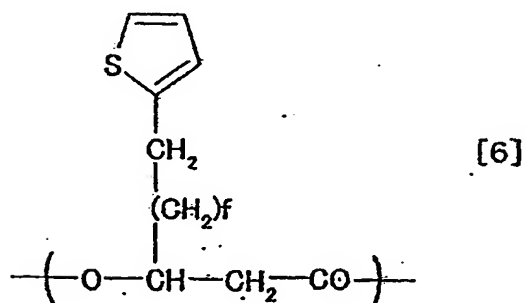


- (wherein d represents any of integers from 0 to 8; R4 represents any one selected from a group of an H atom, a CN group, a NO₂ group, a halogen atom, a CH₃ group, a C₂H₅ group, a C₃H₇ group, a CF₃ group, a C₂F₅ group,

and a C_3F_7 group; and, in the presence of plural units, the foregoing stands independently for each unit.);

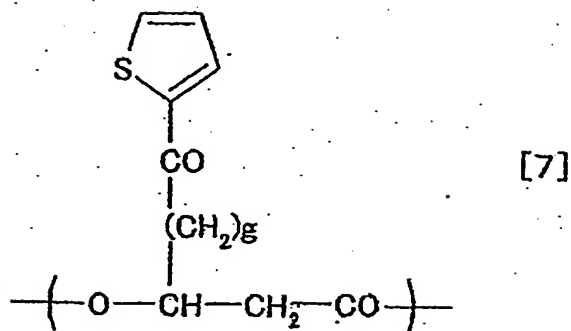


- 5 (wherein e represents any of integers from 1 to 8 and R_5 represents any one selected from a group of a hydrogen atom (H), a halogen atom, $-CN$, $-NO_2$, $-CF_3$, $-C_2F_5$, $-C_3F_7$, $-CH_3$, $-C_2H_5$, and $-C_3H_7$.);

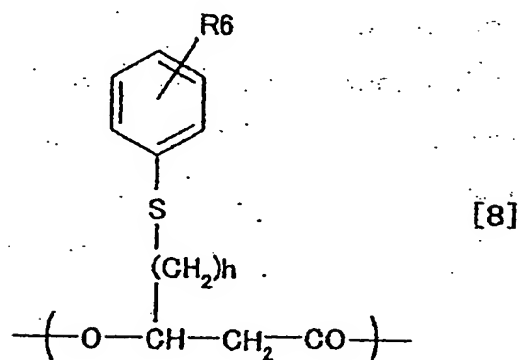


10

(wherein f represents any of integers from 0 to 7.);



(wherein g represents any of integers from 1 to 8);



5

(wherein h represents any of integers from 1 to 7; R6 represents any one selected from a group of a

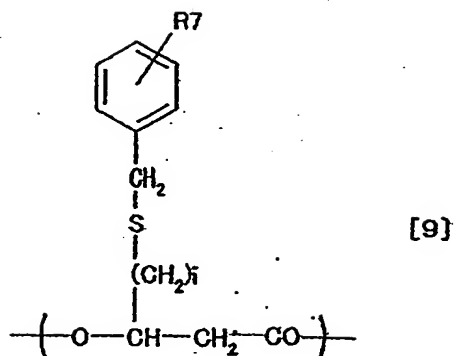
hydrogen atom (H), a halogen atom, -CN, -NO₂, -COOR',

10 -SO₂R'', -CH₃, -C₂H₅, -C₃H₇, -CH(CH₃)₂, and -C(CH₃)₃; R'

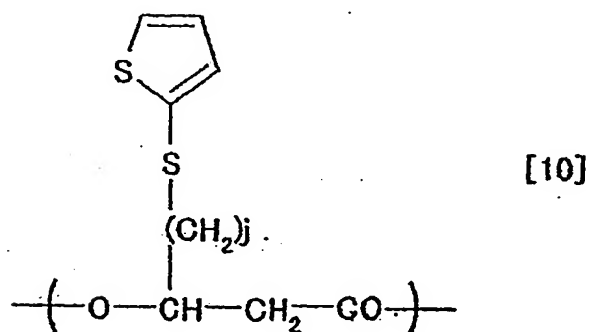
represents a hydrogen atom (H), Na, K, -CH₃, or -C₂H₅;

and R'' represents -OH, -ONa, -OK, a halogen atom,

-OCH₃, or -OC₂H₅);

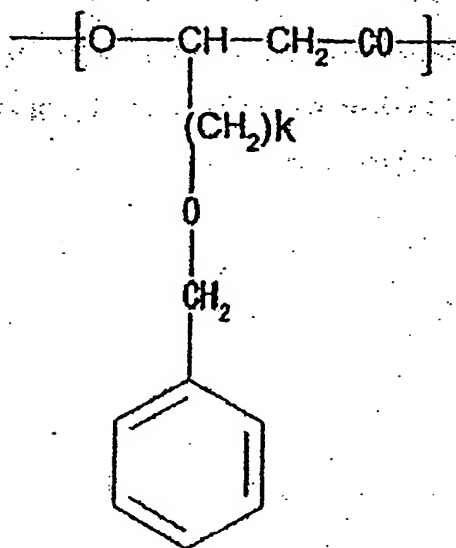


(wherein i represents any of integers from 1 to 7; R⁷ represents any one selected from a group of a
 5 hydrogen atom (H), a halogen atom, -CN, -NO₂, -COOR', and -SO₂R''; R' represents a hydrogen atom (H), Na, K, -CH₃, or -C₂H₅; and R'' represents -OH, -ONa, -OK, a halogen atom, -OCH₃, or -OC₂H₅);



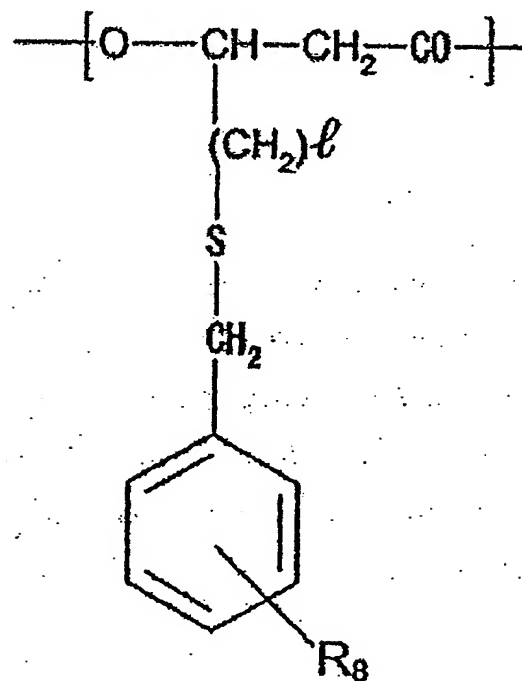
10

(wherein j represents any of integers from 1 to 9);



[A]

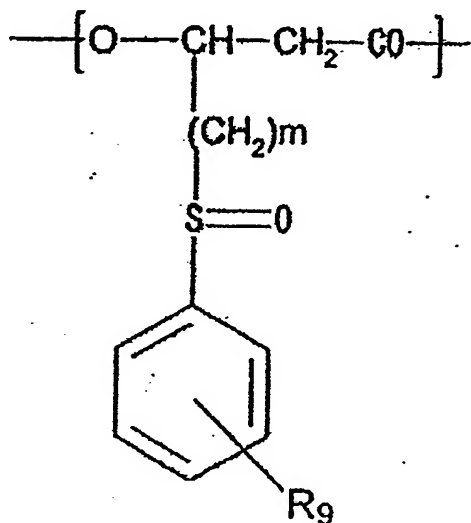
(wherein k represents any of integers from 1 to 8);



[B]

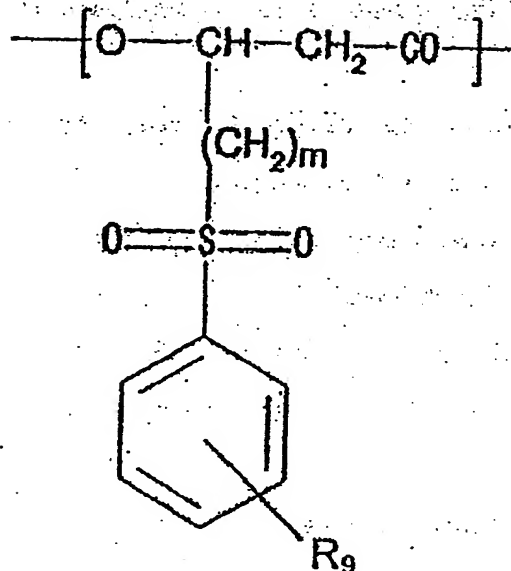
5 (wherein l represents any of integers from 1 to 8; R₈

represents any one selected from a group of a CH₃ group, a C₂H₅ group, a C₃H₇ group, a (CH₃)₂-CH group and a (CH₃)₃-C group; and, in the presence of plural units, the foregoing stands independently for each unit.);



5

(wherein m represents any of integers from 1 to 8; R₉ represents an H atom, a halogen atom, a CN group, a NO₂ group, COOR₉₁, SO₂R₉₂ (R₉₁ representing H, Na, K, CH₃ or C₂H₅, and R₉₂ representing OH, ONa, OK, a halogen atom, OCH₃ or OC₂H₅), a CH₃ group, a C₂H₅ group, a C₃H₇ group, a (CH₃)₂-CH group or a (CH₃)₃-C group; and, in the presence of plural units, the foregoing stands independently for each unit.);



[D]

(wherein n represents any of integers from 1 to 8; R₉ represents an H atom, a halogen atom, a CN group, a NO₂ group, COOR₉₁, SO₂R₉₂ (R₉₁ representing H, Na, K, CH₃ or C₂H₅, and R₉₂ representing OH, ONa, OK, a halogen atom, OCH₃ or OC₂H₅), a CH₃ group, a C₂H₅ group, a C₃H₇ group, a (CH₃)₂-CH group or a (CH₃)₃-C group; and, in the presence of plural units, the foregoing stands independently for each unit.).

4. A construct according to claim 1, wherein at least a part of the polyhydroxyalkanoate is chemically modified polyhydroxyalkanoate.

5. A construct according to claim 4, wherein the chemically modified polyhydroxyalkanoate is polyhydroxyalkanoate having at least a graft chain.

6. A construct according to claim 4, wherein at

least a part the polyhydroxyalkanoate is crosslinked polyhydroxyalkanoate.

7. A construct according to claim 1, wherein the polyhydroxyalkanoate coating the magnetic
5 material has a composition changing in a direction perpendicular to the magnetic material.

8. A construct according to claim 1, wherein the magnetic material is constituted of a metal or a metal compound.

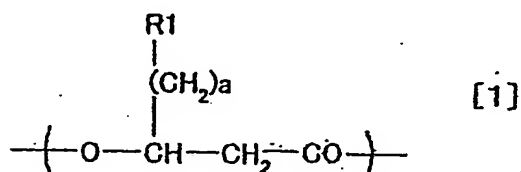
10 9. A construct according to claim 4, wherein the polyhydroxyalkanoate is immobilized to the magnetic material.

10. A method for producing a construct in which at least a part of a magnetic material is coated with
15 polyhydroxyalkanoate, comprising a step of a polyhydroxyalkanoate synthesizing enzyme on a surface of the magnetic material, and a step of causing the enzyme to synthesize polyhydroxyalkanoate from a 3-hydroxyacyl coenzyme A.

20 11. A method for producing a construct according to claim 10, further comprising a step of dispersing the magnetic material in an aqueous medium, prior to the step of immobilizing the polyhydroxyalkanoate synthesizing enzyme.

25 12. A method for producing a construct according to claim 10, wherein the polyhydroxyalkanoate includes at least one selected

from a group consisting of monomer units represented by chemical formulas [1] to [D], 3-hydroxyacyl coenzyme A corresponding respectively to the units is represented by chemical formulas [11] to [20] and [A'] to [D']:



(wherein the monomer unit is at least one selected from a group of monomer units having following combinations of R1 and a:

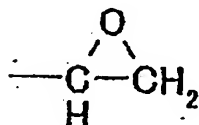
a monomer unit in which R1 is a hydrogen atom (H) and a is any of integers from 3 to 10;

a monomer unit in which R1 is a halogen atom and a is any of integers from 1 to 10;

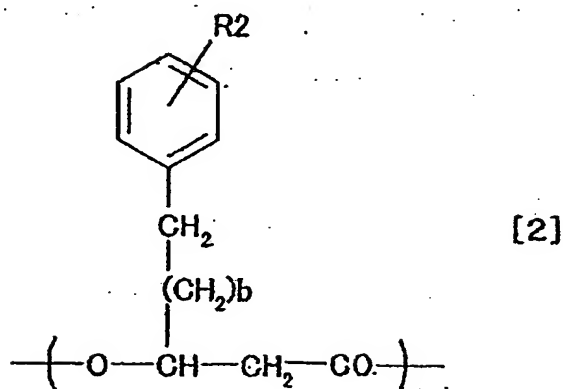
a monomer unit in which R1 is a chromophore and a is any of integers from 1 to 10;

a monomer unit in which R1 is a carboxyl group or a salt thereof and a is any of integers from 1 to 10; and

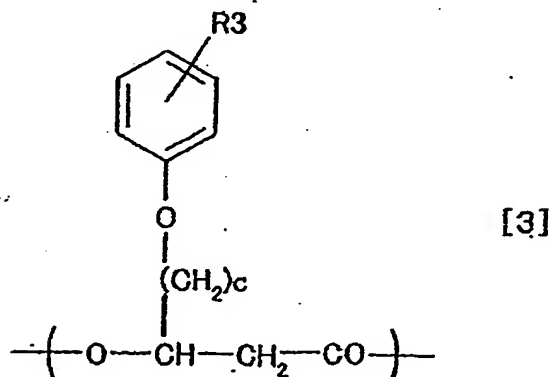
a monomer unit in which R1 is



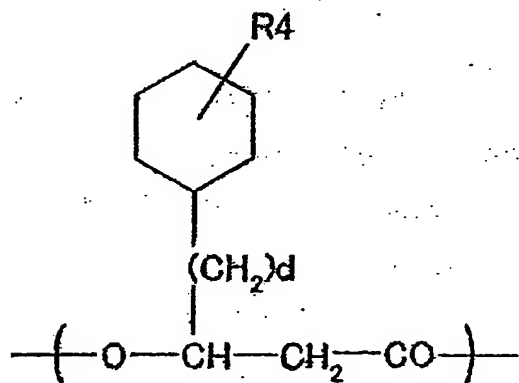
and a is any of integers from 1 to 7.);



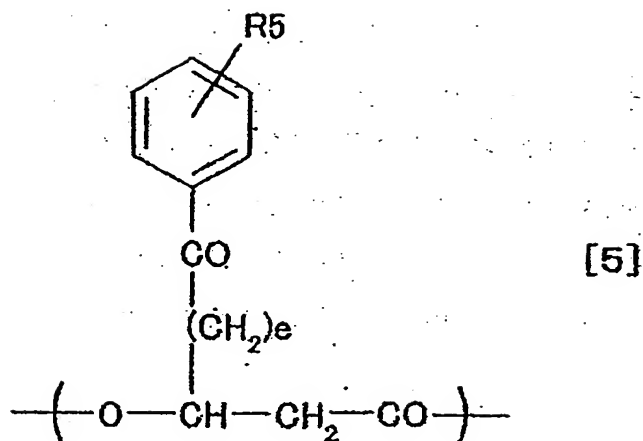
- 5 (wherein b represents any of integers from 0 to 7; R2 represents any one selected from a group of a hydrogen atom (H), a halogen atom, -CN, -NO₂, -CF₃, -C₂F₅, -C₃F₇, a CH₃ group, a C₂H₅ group, a C₃H₇ group, a vinyl group, an epoxy group and COOR₂₁ (R₂₁
- 10 representing an H atom, an Na atom or a K atom); and, in the presence of plural units, the foregoing stands independently for each unit.);



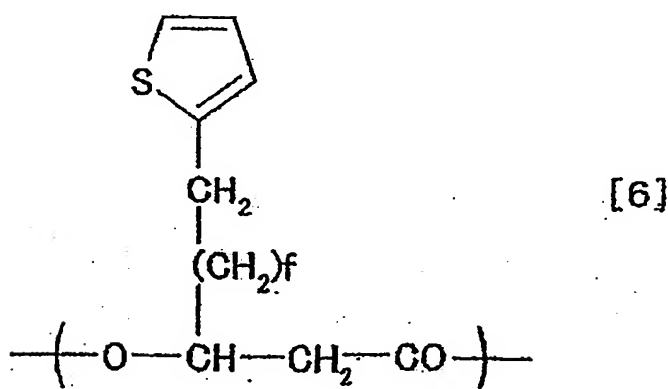
(wherein c represents any of integers from 1 to 8 and R3 represents any one selected from a group of a hydrogen atom (H), a halogen atom, -CN, -NO₂, -CF₃, -C₂F₅, -C₃F₇, a CH₃ group, a C₂H₅ group, a C₃H₇ group, and a SCH₃ group; and, in the presence of plural units, the foregoing stands independently for each unit.);



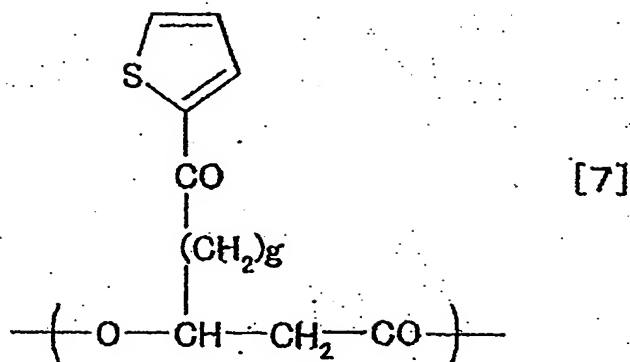
(wherein d represents any of integers from 0 to 8; R4 represents any one selected from a group of an H atom, a CN group, a NO₂ group, a halogen atom, a CH₃ group, a C₂H₅ group, a C₃H₇ group, a CF₃ group, a C₂F₅ group, and a C₃F₇ group; and, in the presence of plural units, the foregoing stands independently for each unit.);



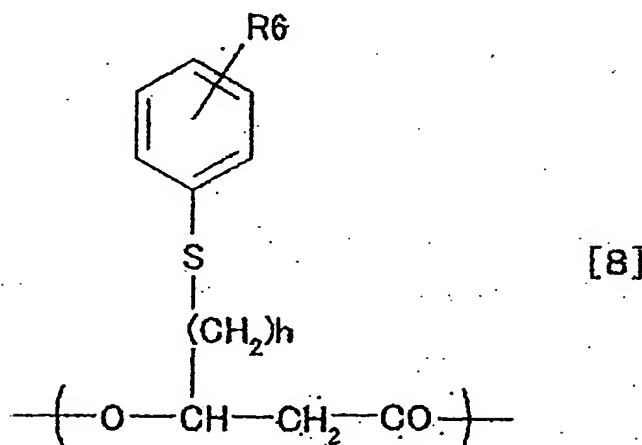
(wherein e represents any of integers from 1 to 8 and
 R5 represents any one selected from a group of a
 hydrogen atom (H), a halogen atom, -CN, -NO₂, -CF₃,
 5 -C₂F₅, -C₃F₇, -CH₃, -C₂H₅, and -C₃H₇.);



(wherein f represents any of integers from 0 to 7.);



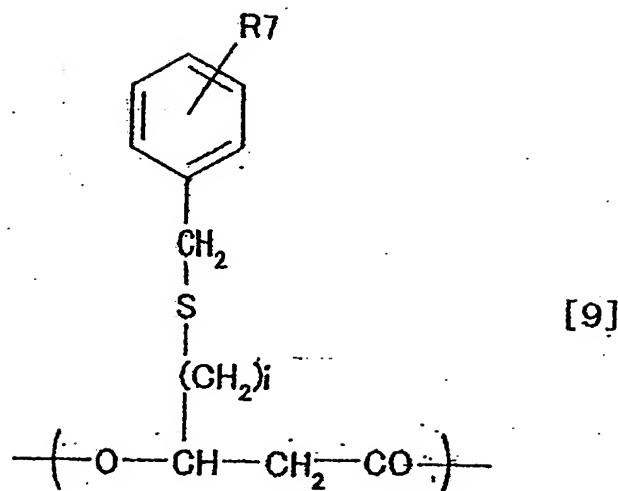
(wherein g represents any of integers from 1 to 8);



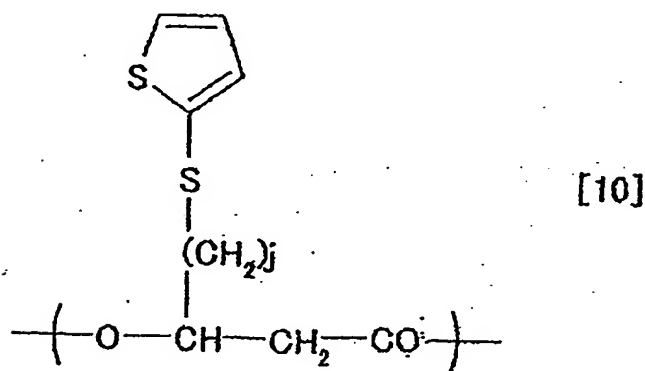
5

(wherein h represents any of integers from 1 to 7; R₆ represents any one selected from a group of a hydrogen atom (H), a halogen atom, -CN, -NO₂, -COOR', -SO₂R'', -CH₃, -C₂H₅, -C₃H₇, -CH(CH₃)₂, and -C(CH₃)₃; R' represents a hydrogen atom (H), Na, K, -CH₃, or -C₂H₅; and R'' represents -OH, -ONa, -OK, a halogen atom, -OCH₃, or -OC₂H₅);

10

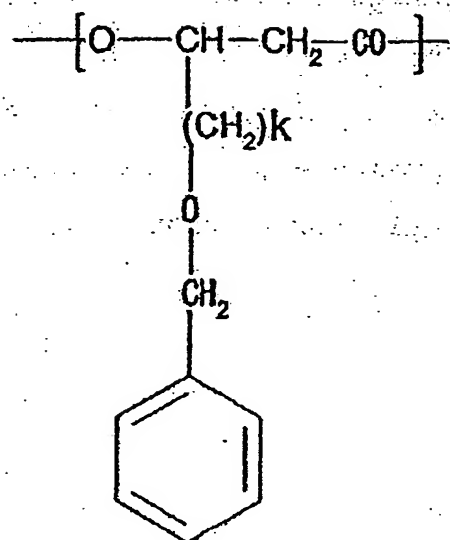


(wherein i represents any of integers from 1 to 7;
 R^7 represents any one selected from a group of a
 5 hydrogen atom (H), a halogen atom, $-\text{CN}$, $-\text{NO}_2$, $-\text{COOR}'$,
 and $-\text{SO}_2\text{R}''$; R' represents a hydrogen atom (H), Na, K,
 $-\text{CH}_3$, or $-\text{C}_2\text{H}_5$; and R'' represents $-\text{OH}$, $-\text{ONa}$, $-\text{OK}$, a
 halogen atom, $-\text{OCH}_3$, or $-\text{OC}_2\text{H}_5$);



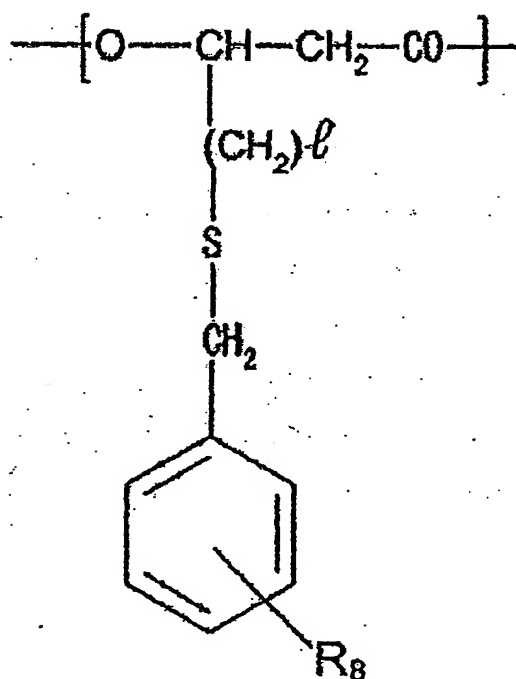
(wherein j represents any of integers from 1 to 9);

251



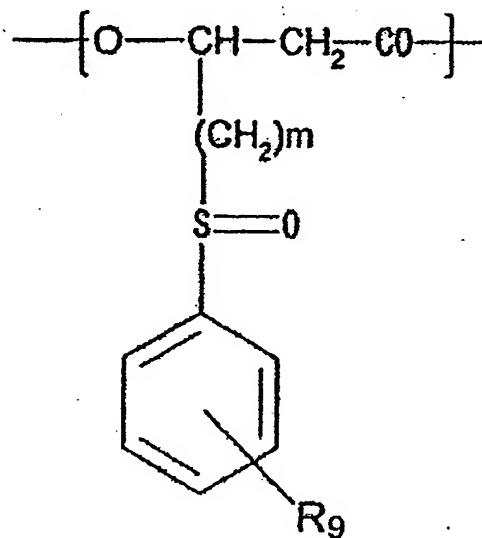
[A]

(wherein k represents any of integers from 1 to 8);

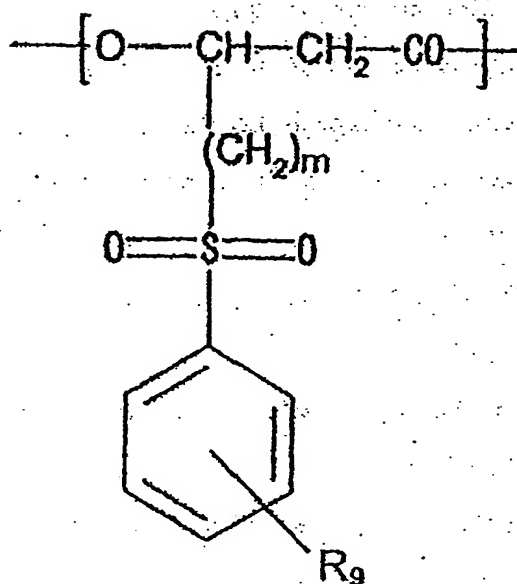


[B]

(wherein l represents any of integers from 1 to 8; R_8 represents any one selected from a group of a CH_3 group, a C_2H_5 group, a C_3H_7 group, a $(CH_3)_2-CH$ group and a $(CH_3)_3-C$ group; and, in the presence of plural units, the foregoing stands independently for each unit.);

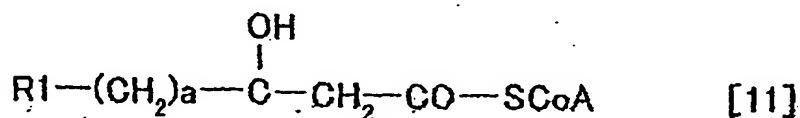


(wherein m represents any of integers from 1 to 8; R_9 represents an H atom, a halogen atom, a CN group, a NO_2 group, $COOR_{91}$, SO_2R_{92} (R_{91} representing H, Na, K, CH_3 or C_2H_5 , and R_{92} representing OH, ONa, OK, a halogen atom, OCH_3 or OC_2H_5), a CH_3 group, a C_2H_5 group, a C_3H_7 group, a $(CH_3)_2-CH$ group or a $(CH_3)_3-C$ group; and, in the presence of plural units, the foregoing stands independently for each unit.);



[D]

(wherein n represents any of integers from 1 to 8; R₉ represents an H atom, a halogen atom, a CN group, a NO₂ group, COOR₉₁, SO₂R₉₂ (R₉₁ representing H, Na, K, CH₃ or C₂H₅, and R₉₂ representing OH, --ONa, OK, a halogen atom, OCH₃ or OC₂H₅), a CH₃ group, a C₂H₅ group, a C₃H₇ group, a (CH₃)₂-CH group or a (CH₃)₃-C group; and, in the presence of plural units, the foregoing stands independently for each unit.);



(wherein -SCoA represents a coenzyme A bonded to alkanoic acid, in which a combination of R1 and a is

at least one selected from a following group,
corresponding to the combination of R1 and a in the
monomer unit represented by the chemical formula [1]:

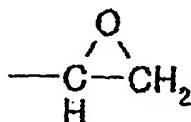
a monomer unit in which R1 is a hydrogen atom
(H) and a is any of integers from 3 to 10;

a monomer unit in which R1 is a halogen atom and
a is any of integers from 1 to 10;

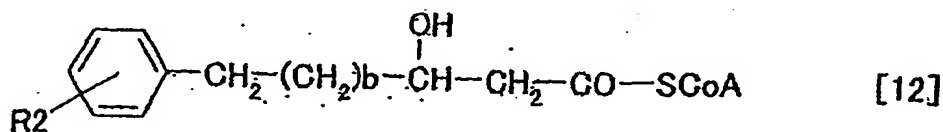
a monomer unit in which R1 is a chromophore and
a is any of integers from 1 to 10;

a monomer unit in which R1 is a carboxyl group
or a salt thereof and a is any of integers from 1 to
10; and

a monomer unit in which R1 is

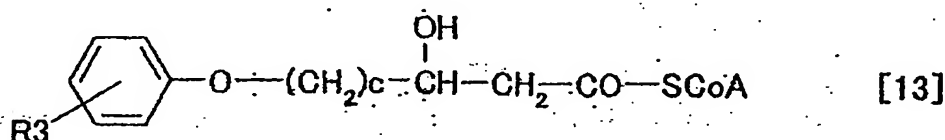


and a is any of integers from 1 to 7.);

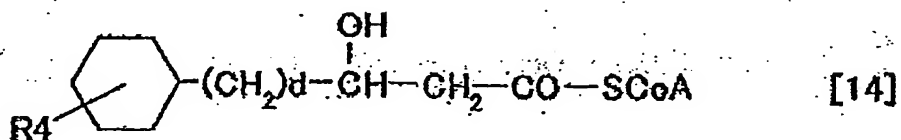


(wherein -SCoA represents a coenzyme A bonded to
alkanoic acid; b corresponds to b in the monomer unit
represented by the aforescribed chemical formula
[2] and represents any of integers from 0 to 7; and

R2 corresponds to R2 in the monomer unit represented by the chemical formula [2] and represents a hydrogen atom (H), a halogen atom, -CN, -NO₂, -CF₃, -C₂F₅, -C₃F₇, a CH₃ group, a C₂H₅ group, a C₃H₇ group, a vinyl group, an epoxy group or COOR₂₁ (R₂₁ representing an H atom, an Na atom or a K atom);

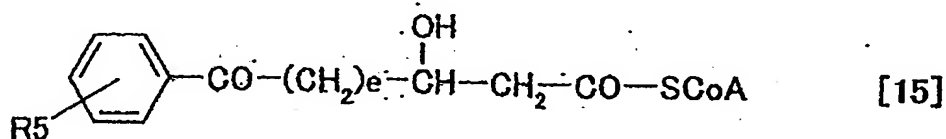


(wherein -SCoA represents a coenzyme A bonded to alkanolic acid; c corresponds to c in the monomer unit represented by the aforescribed chemical formula [3] and represents any of integers from 1 to 8; and R3 corresponds to R3 in the monomer unit represented by the aforescribed chemical formula [3] and represents any one selected from a group of a hydrogen atom (H), a halogen atom, -CN, -NO₂, -CF₃, -C₂F₅, -C₃F₇, a CH₃ group, a C₂H₅ group, a C₃H₇ group and a SCH₃ group.)



(wherein -SCoA represents a coenzyme A bonded to

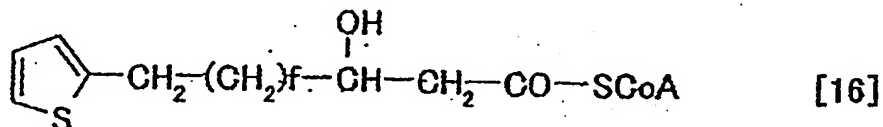
alkanoic acid; d corresponds to d in the monomer unit represented by the aforescribed chemical formula [4] and represents any of integers from 0 to 8; and R4 represents any one selected from a group of an H atom, a CN group, a NO₂ group, a halogen atom, a CH₃ group, a C₂H₅ group, a C₃H₇ group, a CF₃ group, a C₂F₅ group, and a C₃F₇ group.);



10

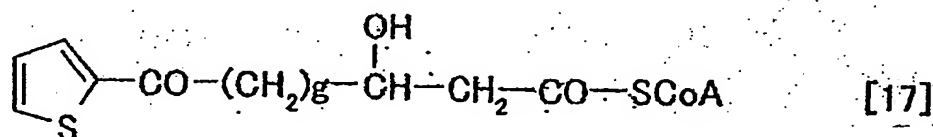
(wherein -SCoA represents a coenzyme A bonded to alkanoic acid; e corresponds to e in the monomer unit represented by the aforescribed chemical formula [5] and represents any of integers from 1 to 8; and R5 corresponds to R5 in the monomer unit represented by the aforescribed chemical formula [5] and represents any one selected from a group of a hydrogen atom (H), a halogen atom, -CN, -NO₂, -CF₃, -C₂F₅, -C₃F₇, -CH₃, -C₂H₅, and -C₃H₇.);

20

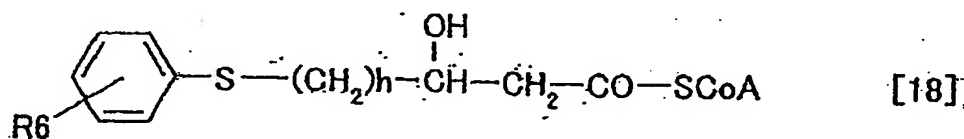


(wherein -SCoA represents a coenzyme A bonded to alkanoic acid; and f corresponds to f in the monomer

unit represented by the aforescribed chemical formula [6] and represents any one of integers from 0 to 7.);



(wherein, -SCoA represents a coenzyme A bonded to alkanolic acid; and g corresponds to g in the monomer unit represented by the aforescribed chemical formula [7] and represents any one of integers from 1 to 8.);



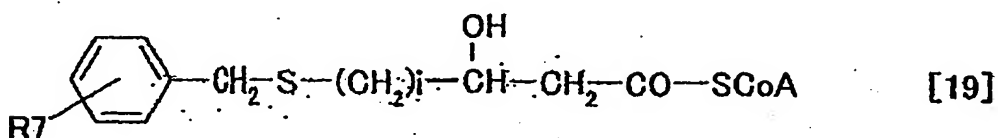
15

(wherein, -SCoA represents a coenzyme A bonded to alkanolic acid; h corresponds to h in the monomer unit represented by the aforescribed chemical formula [8] and represents any of integers from 1 to 7; R₆

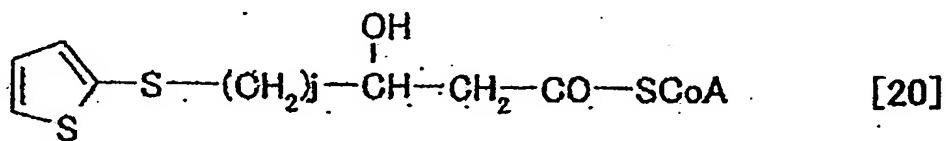
20

corresponds to R₆ in the monomer unit represented by the aforescribed chemical formula [8] and represents any one selected from a group of a hydrogen atom (H), a halogen atom, -CN, -NO₂, -COOR', -SO₂R'', -CH₃, -C₂H₅, -C₃H₇, -CH(CH₃)₂, and -C(CH₃)₃; R'

represents a hydrogen atom (H), Na, K, -CH₃, or -C₂H₅;
and R'' represents -OH, -ONa, -OK, a halogen atom,
-OCH₃, or -OC₂H₅);

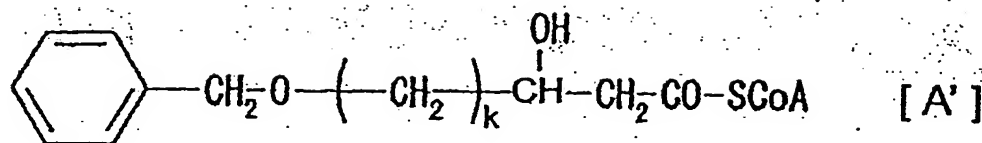


(wherein, -SCoA represents a coenzyme A bonded to
alkanoic acid, i corresponds to i in the monomer unit
represented by the aforescribed chemical formula
[9] and represents any of integers from 1 to 7; R7
corresponds to R7 in the monomer unit represented by
the aforescribed chemical formula [9] and
represents any one selected from a group of a
hydrogen atom (H), a halogen atom, -CN, -NO₂, -COOR',
and -SO₂R''; R' represents a hydrogen atom (H), Na, K,
-CH₃, or -C₂H₅; and R'' represents -OH, -ONa, -OK, a
halogen atom, -OCH₃, or -OC₂H₅);

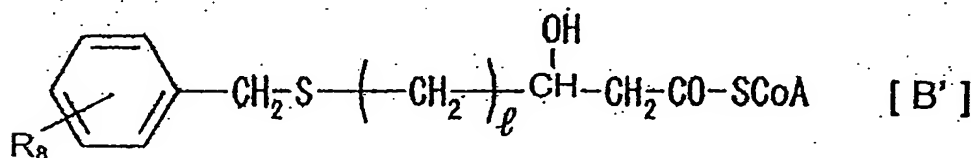


(wherein, -SCoA represents a coenzyme A bonded to
alkanoic acid, and j corresponds to j in the monomer
unit represented by the aforescribed chemical
formula [10] and represents any of integers from 1 to

9);

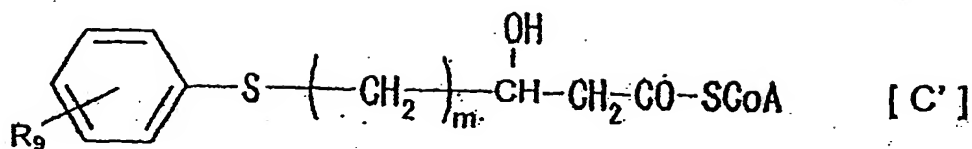


(wherein, -SCoA represents a coenzyme A bonded to
 5 alkanolic acid, and k corresponds to k in the monomer
 unit represented by the aforescribed chemical
 formula [A] and represents any of integers from 1 to
 8);

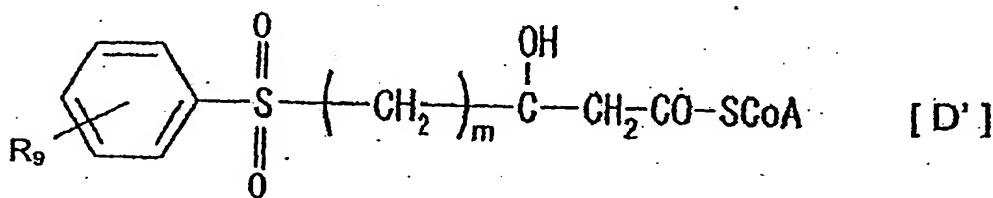


10

(wherein -SCoA represents a coenzyme A bonded to
 alkanolic acid; l corresponds to l in the monomer unit
 represented by the aforescribed chemical formula
 [B] and represents any of integers from 1 to 8; and
 15 R₈ corresponds to R₈ in the monomer unit represented
 by the aforescribed chemical formula [B] and
 represents any one selected from a group of a CH₃
 group, a C₂H₅ group, a C₃H₇ group, a CF₃ group, a
 (CH₃)₂-CH group and a (CH₃)₃-C group.);



(wherein -SCoA represents a coenzyme A bonded to
 alkanolic acid; m corresponds to m in the monomer unit
 5 represented by the aforescribed chemical formula
 [C] and represents any of integers from 1 to 8; and
 R₉ corresponds to R₉ in the monomer unit represented
 by the aforescribed chemical formulas [C] and [D]
 and represents an H atom, a halogen atom, a CN group,
 10 a NO₂ group, COOR₉₁, SO₂R₉₂ (R₉₁ representing H, Na, K,
 CH₃ or C₂H₅, and R₉₂ representing OH, ONa, OK, a
 halogen atom, OCH₃ or OC₂H₅), a CH₃ group, a C₂H₅ group,
 a C₃H₇ group, a (CH₃)₂-CH group or a (CH₃)₃-C group.);
 and.



15

(wherein -SCoA represents a coenzyme A bonded to
 alkanolic acid; m corresponds to m in the monomer unit
 represented by the aforescribed chemical formula
 20 [D] and represents any of integers from 1 to 8; R₉
 represents an H atom, a halogen atom, a CN group, a
 NO₂ group, COOR₉₁, SO₂R₉₂ (R₉₁ representing H, Na, K,

CH₃ or C₂H₅, and R92 representing OH, ONa, OK, a halogen atom, OCH₃ or OC₂H₅), a CH₃ group, a C₂H₅ group, a C₃H₇ group, a (CH₃)₂-CH group or a (CH₃)₃-C group).

13. A method for producing a construct
5 according to claim 10, further comprising a step of producing the polyhydroxyalkanoate synthesizing enzyme with a microorganism having a producing capacity for the enzyme.

14. A method for producing a construct
10 according to claim 10, further comprising a step of producing the polyhydroxyalkanoate synthesizing enzyme with a transformant in which a gene concerning a producing capacity of the enzyme is introduced.

15. A method for producing a construct
15 according to claim 14, wherein the gene is obtained from a microorganism having a producing capacity for the polyhydroxyalkanoate synthesizing enzyme.

16. A method for producing a construct
20 according to claim 13, wherein the microorganism having a producing capacity for the polyhydroxyalkanoate synthesizing enzyme is a microorganism belonging to *Pseudomonas* sp.

17. A method for producing a construct
25 according to claim 16, wherein the microorganism belonging to *Pseudomonas* sp. is at least a microorganism selected from a group consisting of *Pseudomonas putida* P91, FERM BP-7373, *Pseudomonas*

cichorii H45, FERM BP-7374, *Pseudomonas cichorii* YN2, FERM BP-7375, and *Pseudomonas gessenii* P161, FERM BP-7376.

18. A method for producing a construct
5 according to claim 13, wherein the microorganism having a producing capacity for the polyhydroxyalkanoate synthesizing enzyme is a microorganism belonging to *Burkholderia* sp.

19. A method for producing a construct
10 according to claim 14, wherein a host microorganism of the transformant having a producing capacity for polyhydroxyalkanoate synthesizing enzyme is *Escherichia coli*.

20. A method for separating a target component
15 contained in a specimen, comprising:

a step of preparing a carrier in which a molecule having a binding affinity to the target component is immobilized on a surface;

a step of mixing the carrier and the specimen;

20 a step of binding the target component contained in the specimen, to be mixed in the mixing step, with the molecule immobilized on the carrier surface and having the binding affinity in the mixing step; and

a step of separating the target component
25 together with the the carrier from the specimen through the coupling with the molecule having the coupling affinity; wherein the carrier is at least

partly coated with polyhydroxyalkanoate.

21. A method according to claim 20, wherein the carrier includes a magnetic material.

22. A method according to claim 21, wherein the carrier including the magnetic material is separated from the specimen by a magnetic field thereon.

23. A method for detecting a target component contained in a specimen, comprising:

a step of preparing a carrier in which a molecule having a binding affinity to the target component is immobilized on a surface;

a step of mixing the carrier and the specimen;

a step of binding the target component contained in the specimen with the molecule immobilized on the carrier surface and having the binding affinity in the mixing step; and

a step of selectively detecting the target component, immobilized on the carrier in the binding step through the binding with the molecule having the binding affinity;

wherein the carrier is at least partly coated with polyhydroxyalkanoate.

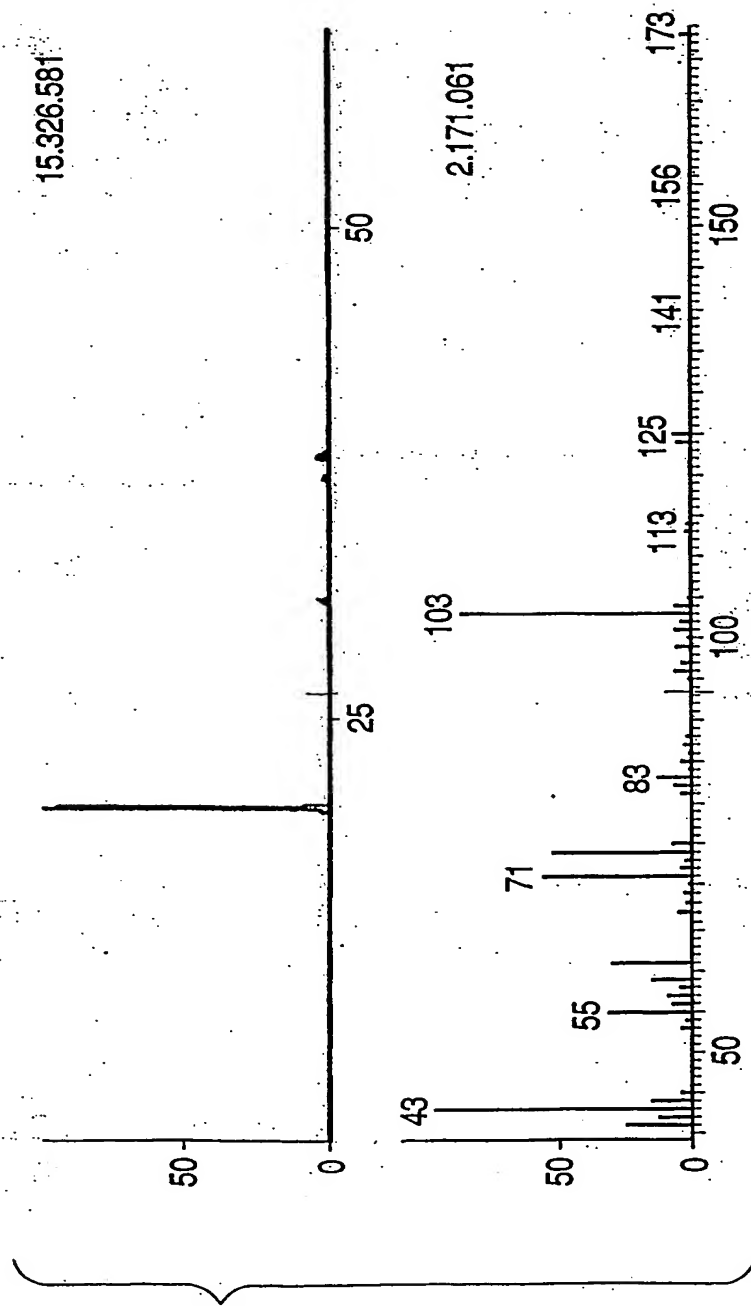
24. A method for screening a target component contained in a crude specimen comprising:

a step of preparing a carrier in which a molecule having a binding affinity to the target component is immobilized on a surface;

- a step of mixing the carrier and the specimen;
- a step of binding the target component contained in the specimen with the molecule immobilized on the carrier surface and having the binding affinity in
- 5 the mixing step; and
- a step of separating the target component together with the carrier from the specimen through the binding with the molecule having the binding affinity;
- 10 wherein the carrier is at least partly coated with polyhydroxyalkanoate.

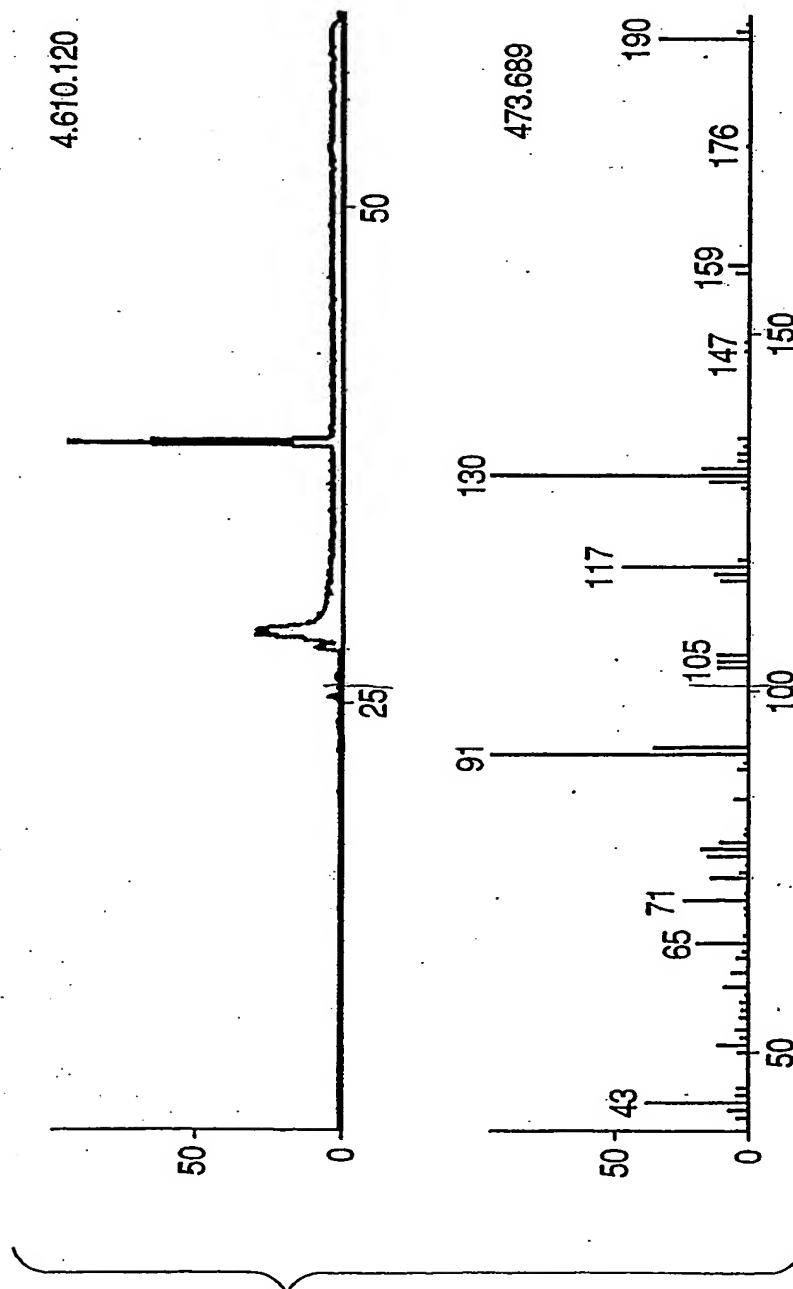
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FIG. 1



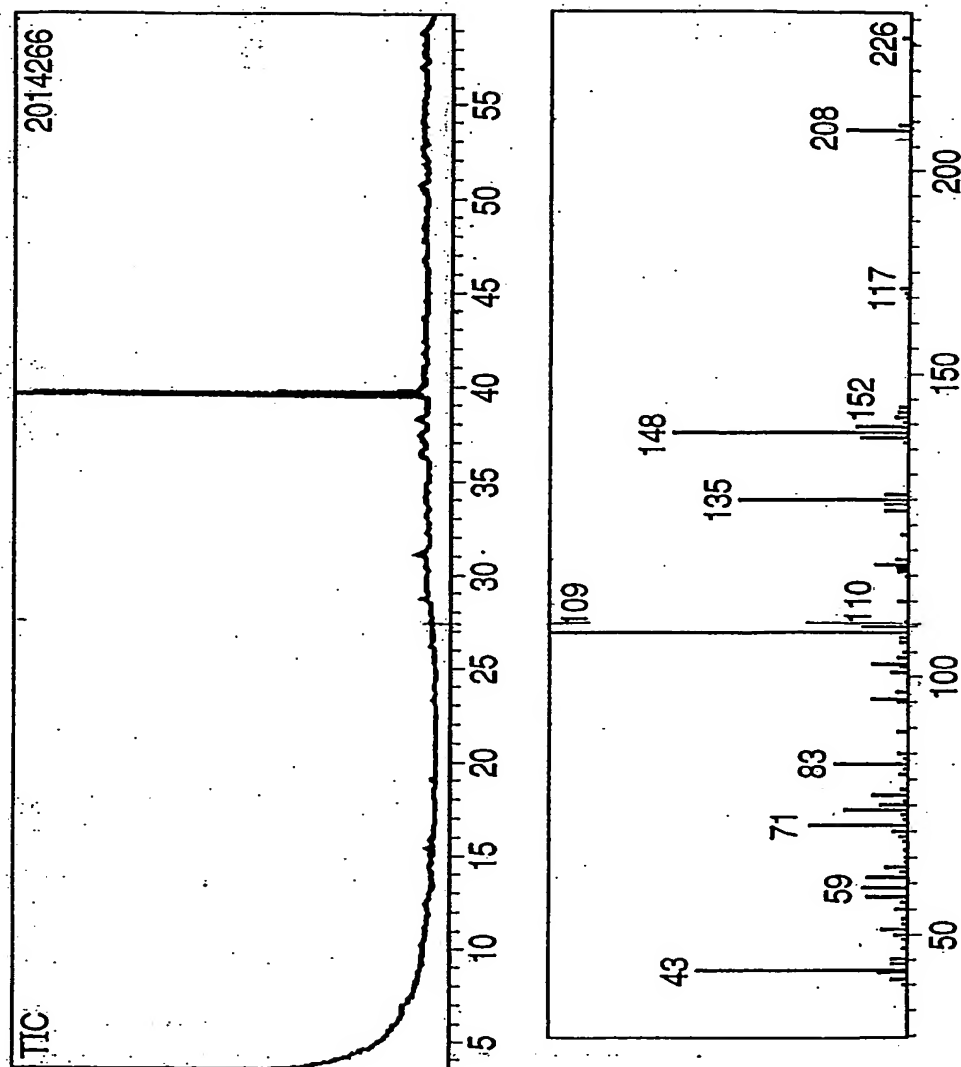
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FIG. 2



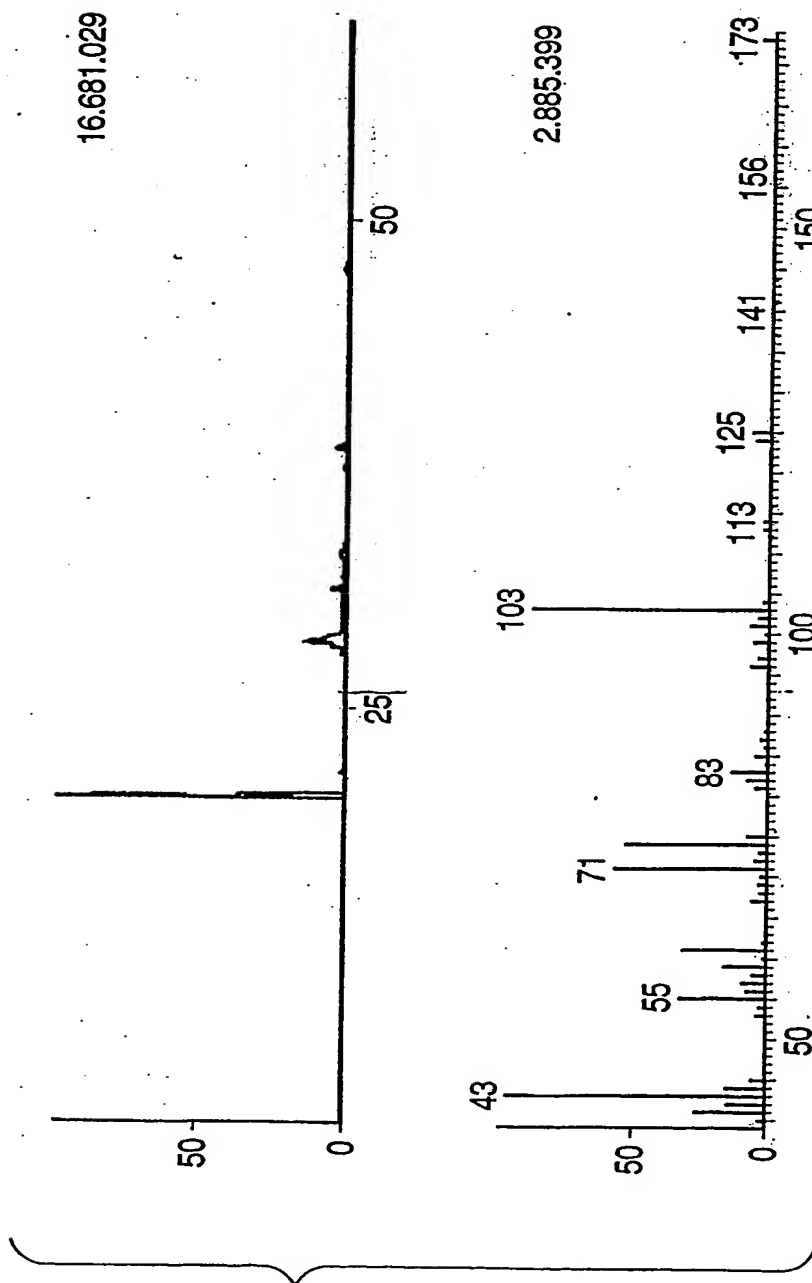
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FIG. 3



4/5

FIG. 4



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FIG. 5

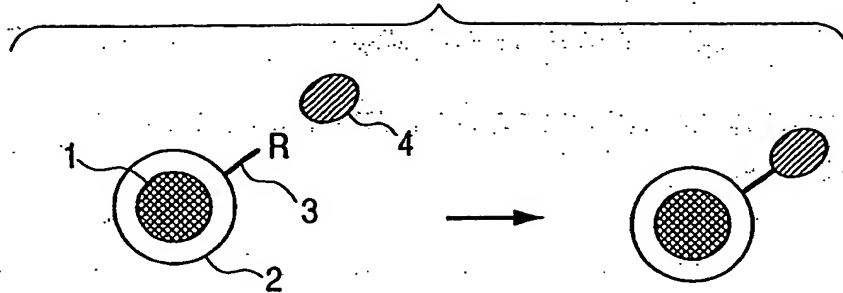


FIG. 6

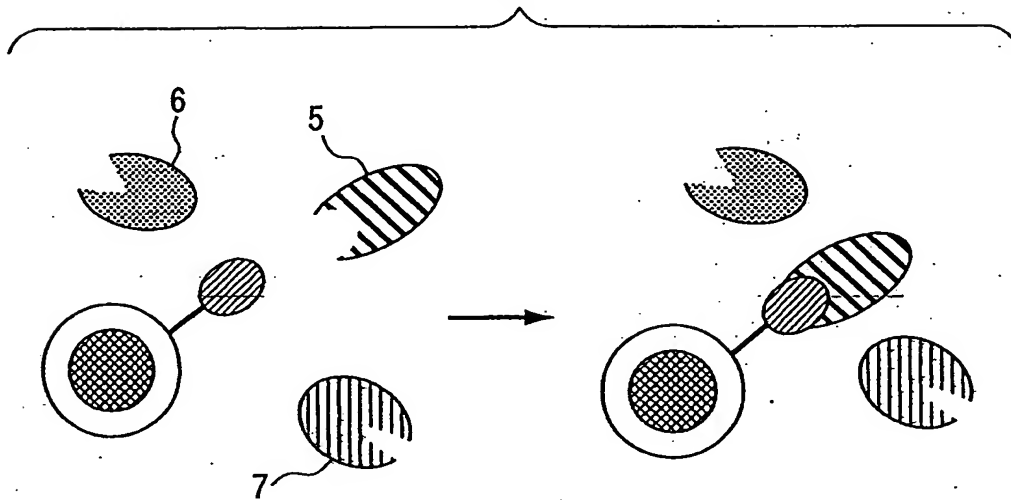
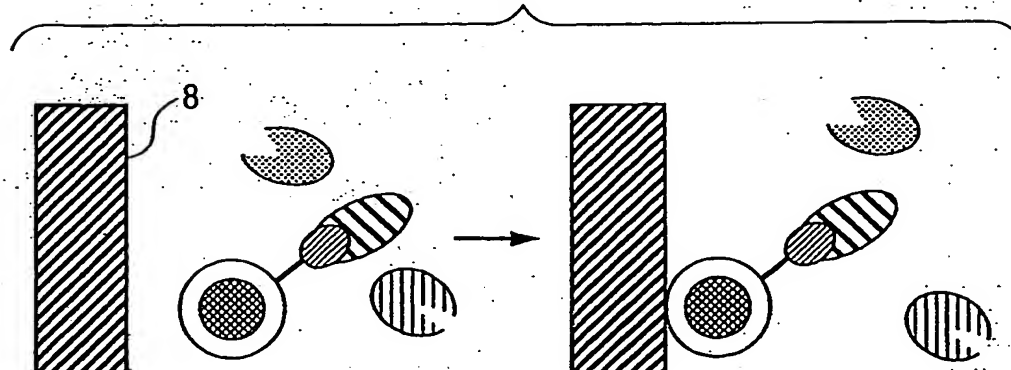


FIG. 7



INTERNATIONAL SEARCH REPORT

PCT/JP2004/006420

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 G01N33/543 G01N33/545 G01N33/551 C12P7/62 C12N11/00
C09D167/04

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 G03G C09D C12P C12N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

EPO-Internal, WPI Data, PAJ

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	EP 1 275 728 A (CANON KK) 15 January 2003 (2003-01-15) page 4, line 50 - line 55 page 6, line 42 - page 15, line 30; claims; examples	1-24
X	EP 1 253 476 A (CANON KK) 30 October 2002 (2002-10-30) page 3, line 40 - page 4, line 22 page 9, line 49 - page 10, line 3	1,2
X	EP 1 253 475 A (CANON KK) 30 October 2002 (2002-10-30) page 6, line 47 - page 10, line 34 page 11, line 10 - page 17, line 52 page 33, line 55 - line 58 page 37, line 29 - line 35	1-19
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☒ Further documents are listed in the continuation of box C.

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Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	US 6 133 047 A (SAUZEDDE FLORENCE ET AL) 17 October 2000 (2000-10-17) the whole document	20-24
A	US 5 648 124 A (SUTOR JAMES J) 15 July 1997 (1997-07-15) the whole document	20-24
A	WO 01/52612 A (ARKIS AHMED ; BIO MERIEUX (FR); DELAIR THIERRY (FR); ELAISSARI ABDELHA) 26 July 2001 (2001-07-26) page 3, line 1 - page 6, line 30; claims; examples	20-24
A	WO 90/15666 A (OMNI QUEST CORP) 27 December 1990 (1990-12-27) the whole document	20-24
A	US 6 521 341 B1 (ELAISSARI ABDELHAMID ET AL) 18 February 2003 (2003-02-18) column 3; claims; examples	20-24
A	US 5 736 349 A (YOSHIOKA KATSUAKI ET AL) 7 April 1998 (1998-04-07) column 5, line 52 - line 56; claims 4,7,8; examples	20-24
A	US 6 231 982 B1 (WANG CHENGRONG) 15 May 2001 (2001-05-15) column 2, line 64 - column 3, line 52; claims	20-24
A	EP 1 213 587 A (COULTER INTERNAT CORP) 12 June 2002 (2002-06-12) the whole document	20-24
A	US 6 048 515 A (LAWACZEK RUEDIGER ET AL) 11 April 2000 (2000-04-11) claims; figure 1; examples	20-24

INTERNATIONAL SEARCH REPORT

Information on patent family members

PCT/JP2004/006420

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
EP 1275728	A	15-01-2003	EP 1275728 A1	15-01-2003
			JP 2004065001 A	04-03-2004
			US 2003224494 A1	04-12-2003
EP 1253476	A	30-10-2002	JP 2002327046 A	15-11-2002
			EP 1253476 A2	30-10-2002
			US 2003096115 A1	22-05-2003
EP 1253475	A	30-10-2002	JP 2003015359 A	17-01-2003
			EP 1253475 A2	30-10-2002
			US 2003118931 A1	26-06-2003
US 6133047	A	17-10-2000	FR 2749082 A1	28-11-1997
			AT 205115 T	15-09-2001
			CA 2227439 A1	04-12-1997
			DE 69706501 D1	11-10-2001
			DE 69706501 T2	16-05-2002
			EP 0840648 A1	13-05-1998
			ES 2162301 T3	16-12-2001
			WO 9745202 A1	04-12-1997
US 5648124	A	15-07-1997	WO 9637313 A1	28-11-1996
WO 0152612	A	26-07-2001	FR 2804117 A1	27-07-2001
			AU 3558001 A	31-07-2001
			CA 2412536 A1	26-07-2001
			EP 1248679 A2	16-10-2002
			WO 0152612 A2	26-07-2001
			US 2003175691 A1	18-09-2003
WO 9015666	A	27-12-1990	WO 9015666 A1	27-12-1990
US 6521341	B1	18-02-2003	FR 2773416 A1	09-07-1999
			AU 1973499 A	26-07-1999
			CA 2316824 A1	15-07-1999
			EP 1046037 A1	25-10-2000
			WO 9935500 A1	15-07-1999
			JP 2002501175 T	15-01-2002
US 5736349	A	07-04-1998	JP 2979414 B2	15-11-1999
			JP 3115862 A	16-05-1991
			US 5320944 A	14-06-1994
			AU 641687 B2	30-09-1993
			AU 6328690 A	11-04-1991
			CA 2026515 A1	30-03-1991
			DE 69021309 D1	07-09-1995
			DE 69021309 T2	01-02-1996
			EP 0420186 A2	03-04-1991
			ES 2077618 T3	01-12-1995
			KR 206159 B1	01-07-1999
			DE 69327926 D1	06-04-2000
			DE 69327926 T2	20-07-2000
			DE 585868 T1	22-09-1994
			EP 0585868 A2	09-03-1994
			ES 2052465 T1	16-07-1994
			JP 6231957 A	19-08-1994
			JP 7092168 A	07-04-1995

INTERNATIONAL SEARCH REPORT

Information on patent family members

PCT/JP2004/006420

Patent document cited in search report		Publication date	Patent family member(s)	Publication date
US 6231982	B1	15-05-2001	DE 69815298 D1	10-07-2003
			DE 69815298 T2	06-05-2004
			EP 0961934 A1	08-12-1999
			ES 2200395 T3	01-03-2004
			JP 2001513203 T	28-08-2001
			WO 9930160 A1	17-06-1999
EP 1213587	A	12-06-2002	US 5466609 A	14-11-1995
			EP 1213587 A1	12-06-2002
			AU 5360394 A	09-05-1994
			CA 2146964 A1	28-04-1994
			DE 69331970 D1	04-07-2002
			DE 69331970 T2	19-12-2002
			EP 0665955 A1	09-08-1995
			JP 8502443 T	19-03-1996
			JP 3547437 B2	28-07-2004
			JP 2004003991 A	08-01-2004
			WO 9409368 A1	28-04-1994
			US 5639620 A	17-06-1997
			US 5776706 A	07-07-1998
			US 5707877 A	13-01-1998
US 6048515	A	11-04-2000	DE 4428851 A1	08-02-1996
			AU 703042 B2	11-03-1999
			AU 2921095 A	04-03-1996
			CA 2195318 A1	15-02-1996
			CN 1155844 A ,B	30-07-1997
			WO 9604017 A1	15-02-1996
			EP 0773796 A1	21-05-1997
			HU 77993 A2	28-04-1999
			IL 114713 A	17-02-2000
			IL 131562 A	31-08-2000
			JP 10503496 T	31-03-1998
			NO 970468 A	02-04-1997
			US 6576221 B1	10-06-2003
			US 2002141943 A1	03-10-2002
			ZA 9506005 A	22-02-1996

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